

# NIGERIAN AGRICULTURAL JOURNAL

ISSN: 0300-368X Volume 53 Number 1, April 2022 Pg. 380-387 Available online at: <u>http://www.ajol.info/index.php/naj</u> <u>https://www.naj.asn.org.ng</u>

Creative Commons User License CC:BY

# Haematological Values, Biochemical Parameters And Antioxidant Enzymes Concentration In Alloxan-Induced Diabetic Rats Treated With *Chrysophyllum albidum Leaf Extracts*

# Chinedu-Ndukwe, P.A., Amadi, A.N.C. and Obeta, C.E.

Department of Zoology and Environmental Biology, Michael Okpara University of Agriculture, P.M.B 7267, Umudike, Abia State \*Corresponding Author's email: chinedu-ndukwe.peace@mouau.edu.ng

#### Abstract

In this study, the anti-diabetic effects of three solvents (petroleum ether, ethanol and chloroform) extracts of Chrysophyllum albidum leaves were evaluated on alloxan induced diabetic rats. Thirty wistar rats of both sexes were assigned to 6 groups (A-F) of 5 rats each. Group A (normal rats) and groups B-F made diabetic via single dose administration of alloxan monohydrate (160 mg/kg). Group B (diabetic control) Groups C, D and E were treated with 500 mg/kg body weight of petroleum ether, ethanol and chloroform extract of Chrysopyllum albidum respectively. Group F was administered glibenclamide (3 mg/kg). All treatments were oral and lasted 14 days. Elevated blood glucose concentrations in the diabetic rats were significantly lowered following treatment with no significant difference observed in the activities of the different extracts (P>0.05). Treatment with the extracts caused further fall in the values of the already decreased red blood cell parameters (RBC, PCV and Hb) with the chloroform extract causing the highest fall and petroleum ether extract the least. WBC count was only significantly (P<0.05) higher in groups treated with petroleum ether and chloroform extracts. Elevations in AST, ALP, bilirubin, urea, creatinine, sodium, chloride, potassium, cholesterol and triglycerides concentrations observed in the diabetic rats were also significantly (P < 0.05) lowered following treatment with the extracts (P<0.05). All extracts also significantly improved the antioxidant strength of the diabetic treated rats. Chrysophyllum albidum may therefore be of value in the management of diabetes mellitus and its associated haematological and biochemical anormalies but should be used along with a haematinic agent.

Keywords: Chrysophyllum albidum, diabetes mellitus, rats, serum

# Introduction

Diabetes mellitus is a metabolic disorder characterized by chronic hyperglycemia due to anomalies in insulin secretion or action and causing impairments in the metabolism of glucose, lipid and protein with glycosuria, polydipsia and polyuria as most common clinical manifestations (Mayfiled, 1998 and Kim et al., 2006). Currently, it is estimated that about 2.8 % of global population is suffering from diabetes mellitus and projected that this prevalence rate may hit 4.4 % by the year 2030 with women and developing countries being more susceptible (Sarah et al., 2004 and Gill et al., 2009). The classification of diabetes mellitus into typed 1 and type 2 and their aetiologies are well documented (Akomas et al., 2014; Ijioma, 2015; Hussian and Theise, 2004). The failure of various organs including eyes, kidneys, nerves, heart and blood vessels in diabetics are thought to be associated with the oxidative stress and activities of free radicals generated from glucose autooxidation and protein glycosylation (Robertson, 2004

and Zozulinska et al., 1998). Currently available management strategies for diabetes mellitus have either been ineffective or too expensive for a larger number of affected individuals. The use of oral anti diabetic drugs is also limited by numerous adverse side effects which may include hematological, cutaneous and gastro intestinal reactions, hypoglycemic coma and impairment of liver and kidney functions. This may be the reason for the current continued search for alternative anti-diabetic agents and renewed interest in medicinal plants (Ijioma, 2015). The modulatory effects of medicinal plants against oxidative diseases like diabetes mellitus have been attributed to their antioxidant and free radical scavenging effects (Pourmorad et al., 2006). A number of these plants contain high amounts of naturally occurring antioxidants such as ascorbic acid, carotenoids, flavonoids and phenolic compounds (Duh et al., 1999). These substances inhibit lipid peroxidation and scavenge free radicals and reactive oxygen species

(Sundararajan et al., 2006); hence the evaluation of their host plants for possible anti-oxidative disease effects. At the moment, only few medicinal plants used in traditional medicine for the treatment of diabetes have received scientific validation (Fattanel, 2012). Chrysophyllum albidum (African star apple) is a rain forest fruit tree belonging to family sapotaceae. The plant is called Udara by the Ibos, Efik and Ibibio, agbalumo by the Yorubas and agwalumo by the Hausas, all of Nigeria (Amusa et al., 2013, Florence and Adiaha, 2005). Extracts from the leaves have been used in the treatment of malaria, high blood pressure, anaemia, stomach ache and diarrhea (Adisa, 2000 and Idowu et al., 2006). The anti-platelet and hypoglycemic effects of the leaf extract have been reported (Adebayo et al., 2010). Extracts from the plant's leaves has also been used in ethno-medicine to manage sprains, bruises, wounds, sterility, asthma and intestinal worms (Okunomo and Egho, 2010). The aim of this current study was to evaluate on comparative basis, the effect of petroleum ether, ethanol and chloroform leaf extracts of Chrysophyllum albidum on the haematology, biochemical and organ histology of all alloxan induced diabetic rats.

#### **Materials and Methods**

#### Sample collection

Fresh samples of *Chrysophyllum albidum* leaves were collected from Umuarigha Oboro village, Ikwuano LGA, Abia State. The plants were identified and authenticated by Dr. M.A. Jimoh, a botanist in the Department of Plant Science and Biotechnology, Michael Okpara University of Agriculture, Umudike, Abia State. A sample of the dried plant was assigned a voucher number MOUAU/ZEB/18/004 and was preserved in a herbarium of the Department of Zoology and Environmental Biology, in the same University.

#### Extract preparation and partitioning

The leaves were properly cleaned and air dried at room temperature on a laboratory bench in accordance with the technique described by Adebayo et al. (2010) and pulverized to powdered form. About 500g of the pulverized sample was macerated in 2.5 liter of absolute ethanol within 48 hours and was thoroughly shaken intermittently and filtered first with muslin cloth and with a whatman filter paper into a beaker. The filtrate was evaporated to dryness on a water bath at 55°C to yield a crude semisolid mass, which weighed 41.3 g and represented a yield of 8.26%. About 20 g of the crude extract was weighed and partitioned using 3 different solvents (chloroform, petroleum ether and methanol) in accordance with the method used by Egua et al. (2013) to obtain extracts of the three solvents and the dry concentrated extract was stored in a refrigerator at 4°C until required.

# **Experimental** Animals

Thirty Wistar albino rat of both sexes, age 12 weeks, weighing 20-50g were procured from Ogive Integrated Farms, Abayi Osisioma L.G.A. Abia State. They had unrestricted access to standard feed and water. The

animals were maintained under standard environmental conditions of temperature, relative humidity and dark and light cycle, in accordance with the guidelines of National Institute of Health Guide for the Care and Use of Laboratory Animals and animal ethics committee of the Department of Zoology and Environmental Biology. Body weight, food consumption and water intake were monitored throughout the period of administration.

#### Induction of diabetes

The mice were fasted overnight and then injected, intraperitoneally, with a single dose of 0.5ml of 160 mg/kg body weight (b/w) of Alloxan monohydrate (a product of Mekphar Chemical Pharmceutical Joint-Stock Company, Chiminh City, Vietnam), dissolved in freshly prepared normal saline, to induce T2D, The control animals (nondiabetic) were injected with 0.5ml of the vehicle (normal saline). Stable hyperglycemia was confirmed on the ninth day using glucometer (ACCU-Check, Roche Diagnostics). Rats with fasting blood glucose greater than 180 mg/dl were considered diabetic and used for this study.

#### Animal grouping and treatment

After 2 weeks of acclimatization, animals were randomly assigned to six groups (n=6/group) of 5 rats each. Group A (normal control) and B (diabetic control):received distilled water (vehicle) daily; Group C: (diabetic + 500mg/kg *C. albidum* extract): received 500mg/kg body weight of the petroleum ether *C. albidum* extract; Group D: (diabetic + 500mg/kg *C. albidum* extract): received 500mg/kg body weight of methanol *C. albidum* extract; Group E: (diabetic + 500mg/kg *C. albidum* extract): received 500mg/kg body weight of methanol *C. albidum* extract): received 500mg/kg body weight of chloroform *C. albidum* extract; while Group F: (diabetic + glibenclamide): received 3.0 mg/kg body weight of standard drug (glibenclamide). All treatments were through oral administration and lasted for 14 days.

# **Blood** collection

At the end of experiment, four (4) mice, from each group, were sacrificed and blood samples for hematological and biochemical assays were collected from each mouse through the orbital sinus with heparinized capillary tubes into EDTA treated bottles and plain bottles respectively, thereafter, mice were sacrificed. Hematological parameters were determined using standard procedures (Mukherjee *et al.*, 2007). Haematological parameters include red blood cell (RBC) count, haemoglobin (Hb) concentration, Packed cell volume (PCV), white blood count (WBC), platelet (PLT) count, reticulocytes count and WBC differential count were determined for each blood sample collected in accordance with the standard techniques described by Baker *et al.* (1998) and Cheesbrough (2000).

# **Biochemical analysis**

Alanine aminotransferase (ALT), aspartate aminotransfarase (AST), alkaline phosphatase (ALP), total biliribin, total protein, urea, creatinine, albumin, serum potassium, and serum sodium.

#### Statistical analysis

Data generated from the study were analyzed using statistical package SPSS version 22.0 (2020) Group comparisons were done using the analysis of variance (ANOVA). Significant differences between control and experimental were assessed by least significant difference (LSD). All data were expressed as mean  $\pm$  SEM. P-values less than 0.05 were considered to be significant.

# **Results and Discussion**

#### Results

#### Effect of administration of 3 solvent extract of Chrysophyllum albican on body weight changes in alloxan-induced diabetic mice

Body weight gain was significantly lower in the diabetic control group when compared with the normal control group (P<0.05). However, treatment with extracts also significantly increased body weight gains in the test group, ameliorating the effect of induction of diabetes on body weight. Comparative evaluation of the effect of the extracts on body weight showed that petroleum ether extract improved body weight more and was followed by chloroform extract while ethanol extract performed least. The three extracts however performed better than glibenclamide, the standard drug used in terms of body weight improvement (Table 1).

### *Effect of administration of 3 solvent extract of Chrysophyllum albican on blood glucose concentrations in alloxan-induced diabetic mice*

Treatment of the diabetic rats with the extracts significantly lowered their elevated blood glucose concentrations and successfully returned the values to normal by end of the treatment period. No significant difference was observed between the activities of the different extracts (P>0.05).

# *Effect of the Chrysophyllum albican leaf extract on hematological parameters in alloxan-induced diabetic rats*

The induction of diabetes mellitus caused significant decrease in red blood cell parameters (RBC, PCV and Hb) when compared with the normal rats (P < 0.05). Treatment with the extracts caused further decrease in the values of these parameters with the chloroform extract causing the highest fall and petroleum ether extract the least. The group treated with glibenclamide however significantly improved red blood cell parameters when compared with the diabetic untreated group (P < 0.05) and did not significantly differ from the values in the normal control group (P>0.05). WBC count was only significantly higher in groups treated with petroleum ether and chloroform extracts. Elevated platelets counts following the induction of diabetes was also significantly lowered following treatment with the extracts and the standard drug (Table 3).

# Effect of the Chrysophyllum albidum leaf extract on liver function parameters in alloxan-induced of diabetic rats

The slight elevations in AST and ALP values observed

in the diabetic rats following induction of diabetes mellitus was only significantly lowered in group treated with the standard drug (glibenclamide), but elevations in ALT values were significantly lowered following treatment with ethanol extract, chloroform extract and glibenclamide (P<0.05). All extracts also significantly lowered elevated bilirubin and improved the low total protein values in the diabetic rats (Table 4).

# Effect of the Chrysophyllum albidum leaf extract on renal function parameters in alloxan-induced of diabetic rats

The elevated urea and creatinine concentrations in the diabetic rats were significantly lowered following chloroform extract treatment (P<0.05). Elevated sodium concentrations in the diabetic rats were also significantly lowered in all test groups following treatment with the extracts (P<0.05). Raised serum potassium concentration was also lowered after treatment with petroleum ether and chloroform extracts. Chloride concentration was only lowered in diabetic rats treated with ethanol extract and glibenclamide. The concentrations of bicarbonate did not significantly change in all test groups when compared with the diabetic untreated rats (P<0.05). These results are presented in Table 5.

# Discussion

Higher extract yield (12.2%) obtained with ethanol after extractions with the three different solvents suggest that ethanol may have collected more of the bioactive compounds Chrysophyllum albidum than the other solvents (petroleum ether and chloroform) used which gave a yield of 9.70% each. The fact that ethanol is a polar solvent further suggests that the phytocomponents in Chrysophyllum albidum are more soluble in polar solvent than non-polar ones. This result agrees with Do et al., (2014), who reported that extraction solvents usually affect the quality, quantity and pharmacology of plant extracts generated during extraction and that a polar solvent like ethanol is likely to give higher extract yield than non-polar ones. Improvements observed in the test groups following treatment also suggest that the various extracts used may contain active components with anti-diabetic properties. The higher weight gains in groups treated with petroleum ether and chloroform extracts may be due to the presence of more lipids in these extracts which may have augmented that which is present in the animal's body leading to increase in body weight. This probably indicates that the extract might not exert its hyperglycaemic effect through weight reduction. The presence of tannins in the extracts may have also contributed to the weight gains observed in the extract treated groups (Adewoye et al., 2012). The fall in blood glucose levels in the diabetic rats gives credence to the hypoglycaemic effect of *Chrysophyllum albidum* leaf extract and may have been achieved via one or a combination of mechanisms including reactivation of destroyed beta cells with subsequent increase in insulin production and secretion, decreased glucose absorption in the gastrointestinal tract and increased mobilization of glucose molecules into cells to be metabolized

(Ijioma et al., 2014). These results obtained for Chrysophyllum albidum leaf extracts agree with an earlier report of Olorunnisola et al. (2008) and Adebayo et al., 2010, on the anti-hyperglycemic and hypoglycaemic effects of ethanol extract of C. albidum. The Assessment of haematotogical parameters can be used to explain blood related functions of a plant extract (Yakubu et al., 2007). Hence, analysis of blood parameters is relevant in risk evaluation as changes in the haematological system have higher predictive value for toxicity and state of health (Olson et al., 2000). The fall in red blood cells number, haemoglobin concentration and packed cell volume following induction of diabetes gives credence to the report that anaemia is one of the clinical manifestations of diabetes mellitus and is attributable to the destruction of RBC and reduced rate of its production in the bone marrow due to the oxidative effect of alloxan and increased in lipid peroxidation of the erythrocyte cell membrane (Akomas et al., 2014) all as a result of alloxan treatment. The further fall in the level of these blood parameters observed in the diabetic rats treated with the extracts suggest that the extracts may have some toxicity effects on blood, not minding its blood glucose lowering activity. Adewoye et al. (2012) had reported that methanol extract of C. albidum bark caused haemorrhagic anaemia in experimental rats. This RBC lowering effect of the extract may also be due to the antibiotic effects of the extract. Leaf extract of Chrysophyllum albidum is known to be a potent agent against malaria parasites (Adisa, 2000; Idowu et al., 2006; Florence and Adiaha, 2015; Adewoye et al., 2011). Agents with such effects have greatly been implicated in post treatment anaemia due to their destructive effects on red blood cells (Girdwood, 1976). The mild rise in WBC values, particularly in the groups treated with petroleum ether and chloroform extracts may be a normal reaction of the animals to foreign substances. It is established that leuococytosis may be a physiological response to a stimulated immune system aimed at protecting the body against infections caused by chemical and secondary infections (Celik and Suzek, 2008). There was a significant increase in the platelet count of diabetic control compared to the normal control. The increased platelet count in the diabetic rats suggests that thrombocytopaenia may be another clinical manifestation in diabetes mellitus, which may be why diabetics are usually prone to blood clotting disorders such as thrombosis (Ijioma, 2015). In diabetics, declined insulin release may cause loss of anti-platelets aggregation activity and defective endothelial production leading to bleeding disorders. Accumulation of products of advanced glycosylation in diabetics coupled with reduced membrane fluidity of platelets may also contribute to platelet hyper function as observed in this study. The lowering effects of the extracts on these platelets values suggest that the extracts may have some level of anti-platelet activity and may therefore be of further value in the management of blood clotting disorders and their associated cardioavascular challenges in diabetes mellitus rise in serum Aspartate aminotransferase (AST) and Alanine

aminotrasferase (ALT) values beyond the upper limits of the normal ranges are usually indicative of liver toxicity and/or damage, while increase in alkaline phosphatase (ALP) may suggest biliary tract obstruction. Results of this study have shown no obvious changes in the values of these parameters following induction of diabetes and treatment with the extracts and may be explained by the shortness of the period of study. Liver damage has indeed been associated with chronic diabetes mellitus (Mohammed et al., 2016). The rise in bilirubin concentrations in the diabetic rats may be due to increased RBC haemolysis in the diabetic animals even as reductions in the values of this parameter in the extract treated groups suggest possible modulation and membrane stabilizing activities on the part of the extracts. Abiodun et al. (2011) had reported the hepato-protective effects of leaf extracts of C. albidum against carbon tetrachloride  $(CCl_{4})$  induced liver damage in wistar rats. The increase in urea, creatinine and sodium concentrations in the diabetic rats may indicate threat on the renal system due to the oxidative effect of alloxan and also the oxidative stress which usually accompany diabetes mellitus (Pari and Uma, 2000). On these, the chloroform extract of C. albidum offered significant ameliorative effect, indicating possible area of usefulness of the agent.

#### Conclusion

If the results obtained in the present study can be extrapolated to humans, then the three solvent extracts of *Chrysophyllum albidum* could be of value in the management of diabetes mellitus having shown significant levels of anti-hyperglycaemia, improvements of haematological activities and good modulatory effects on liver and renal functions in diabetic rats. However the use of a haematinic agent alongside treatment may be encouraged due to the adverse effects of the extract on red blood cells.

#### References

- Abiodun, H. A., Amos, O. A., Roseline, K., Olayemi, O. A. and Titilayo, B. O. (2011). Antioxidant activities of the Chrysophyllum albidum. Pakistan Journal of Pharmaceutical Sciences, 24 (5): 545-551.
- Adebayo, A. H., Abolaji, A. O., Kela, R., Ayepola, O. O., Olorunfemi, T. B. and Taiwo, O. S. (2011). Antioxidant activities of the leaves of Chrysophyllum albidum. Pakistan Journal Pharmacitical Science. 24(4):545-51.
- Adebayo, H. A., Abolaji, A. O., Opata, T. K. and Adegbenro, I. K. (2010). Effects of ethanolic leaf extract of *Chrysophyllum albidum G*. on biochemical and haematological parameters of albino wistar rats. *African Journal of Biotechnology*, 9(14): 2145-2150.
- Adewoye, E. O., Salami, A. T and Emikpe, B. O. (2012). Effect of methanolic extract of *Chrysophyllum albidum* bark on hematological indices in mice with experimental hemorrhagic anemia. *African Journal of Biomedical Research*, 15:85-91.
- Adisa, S. A. (2000). Vitamin C, protein and mineral content of African apple *Chrysophyllum albidum* in

the proceedings of the  $18^{th}$  annual conference of NIST (Eds) 141-146.

- Akomas, S.C., Okafor, A.I., and Ijioma S.N., (2014) Hypoglycemic, Hematologic and Hypolipidemic Activity of *Mucuna pruriens* ethanol leaf extract in alloxan induced diabetic rats Annual Research and Review in Biology, 4(24): 4284-4292.
- Amusa, N. A., Ashaya, O. A. and Olaadapo, M. O. (2003). Biodeterrioration of the African star apple (*Chrysophyllum albidum*) in storage and the effect on its food value. *African Journal of Biotechnology*, 22: 56-59.
- Baker, F. J., Silverton, R. E. and Pallister, C. J. (1998). Baker and Silverton's Introduction to Medical Laboratory Technology. Seventh ed. Pp. 356-360.
- Celik, I. and Suzek, H. (2008). The hematological effects of methyl parathion in rats. *Journal of Hazardous Material*, 153:1117-21
- Cheesbrough, M. (2000). District laboratory practices in tropical countries part-2. Low price edition, Pp 267-334.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S. and Ju, Y. (2014). Effect of extraction solvent on total phenol content, total flavonoid content and antioxidant activity of *Limnophila aromatic*. *Journal of Food and Drug Analysis*, 22(3): 296-302.
- Doumas, B. T., Watson, W. A., Biggs, H. G. (1971). Albumin standards and the measurement of serum albumin and bromecresol green. *Clinical Chemistry*, 31: 87-96.
- Duh, P. D., Tu, V. V. and Yen, G. C. (1999). Antioxidant activity of aqueous extract of Harnjyur (*Chrysanthemum morifolium*). *Lebenswiss Technology*, 32: 269-277.
- Egua, M. O., Etuk, E. O., Bello, S. O. and Hassan, S. W. (2013). Antidiabetc potential of liquid-liquid portion fractions of ethanolic seed extract of *Corchorus olitorious. Journal of Pharmacolognosy and Phytotherapy*, 6(1): 4-9.
- Fattanel, H. D., Mohammed, K., Asieh, S. and Mehri, A. F. (2012). Pre4senting anti-diabetic plants in Iranians traditional medicine. *Journal of Diabetes* and Endocrinology, 3: 514-564.
- Gill, G. U., Mbanya, J. C., Ramaiya, K. I. and Tesfays, S. (2009). A sub-Saharan African perspective of diabetes. *Diabetologia*, 52:8-16.
- Girdwood, R. H. (1976). Drug-induced anaemias. *Drugs*, 11(5): 394-404.
- Hussain, M. A. and Theise, N. D. (2004). System cell Therapy for Diabetes mellitus. *Lancet*, (364): 205-208
- Idowu, T. O., Iwalewa, E. O., Aderogba, M. A., Akinpelu, B. A. and Ogundaini, A. O. (2006). Biochemical and behavioural effects of elegnine from *Chrysophyllum albidum*. Journal of Biological Sciences, 6: 1023-1034.
- Ijioma, S. N., Okafor, A. I., Ndukuba, P. I., Nwankwo, A. A. and Akomas, S. C. (2014). Hypoglycemic, hematologic and lipid profile effects of *Chromolaena odorata* ethanol leaf extract in

alloxan induced diabetic rats. *Annals of Biological Sciences*, 2(3): 27-32.

- Kim, S. J., Ju, B. J., Choi, W. C. and Kin, C. S. (2006). Hypoglycemic and antihyperlipidemic effect of four medicinal plants in alloxan induced diabetic rats. *American Journal of Biochemistry and Biotechnology*, 2(4): 154-160.
- Luke, U. O., Ebong, P. E., Eyong, E. U., Robert, A. E., Ufot, S. U. and Egbung, G. E (2013) Effect of ethanolic root and twig extracts of *vernonia amygdalina* (etidot) on liver function parameters of streptozotocin induced hyperglycaemic and normal wistar rats. *European Scientific Journal*, 9(30): 1857–7881.
- Mayfield, J. (1998). Diagnosis and classification of diabetes mellitus. New criteria. *American Academy and family Physician*, 58(6): 1355-1358.
- Mohammed, J., Nazratun, N., Zariyantey, A. H. and Budin, S. B. (2016). Mechanisms of Diabetesinduced liver damage: The role of oxidative stress and inflammation. *Sultan Qaboos University Meducal Journal*, 16(2): 132-141.
- Olorunnisola, D. S. I., Amao, I. S., Ehigie, D. O., Ajayi, Z. A. F. (2008). Anti-hyperglycemic and hypolipdemic effect of ethanolic extract of *Chrysophyllum albidum* seed cotyledon in alloxan induced. Diabetic rats. *Research Journal of Applied Science*, 3(2): 123-127.
- Olson, H., Betton, G., Robinson, D., Thomas, K., Monro, A., Kolaja, G., Lilly, P., Sander, J., Sipes, G., Bracken, W., Dorato, M, Deun, K. V., Smith, P. Berger, B. and Heller, A. (2000). Concordance of toxicity of pharmaceuticals in humans and in animals. *Regulatory Toxicological and Pharmacology*, 32: 56-67.
- Omoboyowa, D. A., Igara, E. C., Christian, G. O. and Olugh, K. D. (2016). Anti-diabetic activity of methanolic extract of seed cotyledon of *Chrysophyllum albidum* in alloxan induced diabetic rats. *Nigerian Society for Experimental Biology*, 28(2): 88-95.
- Oyebade, B. A., Ekeke, B. A. and Adeyemo, F. C. (2011). Fruits categorization and diagnostic analysis of *Chrysophyllum albidum* (G. Don) in Nigeria. *Advance in Applied Science Research*, 2(1): 7-15.
- Pari, L. and Uma, M. (2000) Antihyperglysemic activity of *Musa sapientum* flowers: Effect on lipid peroxidation in alloxan induced diabetic rats. *Phytotherapy Research*, 14(2): 136-138.
- Pourmorad, F., Hosseinmelu, S. J. and Shahabimaid, N. (2006). Antioxidant activities, phenols, flavonoids contents of selected Iranian medicinal plants. *Science African Journal of Biotechnology*, 5: 1142-1145.
- Robertson, R. P. (2004). Chronic oxidative stress as a central mechanism for glucose toxicity in pancreatic islet beta cells in diabetes. *Journal of Biological Chemistry*, 279(41): 42351-423554.
- Sarah, W., Gojka, R., Anders, G., Richard, S. and Hilary, K. (2004). Global prevalence of diabetes estimates for the year 2000 and protections for 2030.

*Diabetes Care*, 27(5):1047

- Searcy, R. L., Reardon, J. E. and Foreman, J. A. (1967). Enzymatic serum urea determination. *American Journal of Medical Technology*, 33: 15-20.
- Shae, J. E., Siecree, R. A. and Zimmet, P. Z. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Research. Clinical. Practice*, 87:4-14.
- Shah, M., Patel, P. and Phadke, M. (2002). Evaluation of the effect of aqueous extract from powders of root, stem, leaves and whole plant of *Phyllanthus debilis* against CCl<sub>4</sub> induced rat liver dysfunction. *Indian Drugs*, 39: 333-337.
- Shahidi, F., Janitha, P. K. and Wanasundara, P. D. (1992). Phenolic antioxidants. *Critical Review of Foodscience and Nutrition*, 32 (1):67-103.
- Lowel, B. B. and Shulman, G. I. (2005). Mitochondrial dysfunction and type 2 diabetes. *Sciences*, 307(5708):384-387.
- Sharma, S. B., Hasir, A., Prabhu, K. M., Murthy, P. S. and Dez, G. (2003) Hypoglycemic and hypolipidemic effect of ethanolic extract of seeds of

*Eugenia jambolona*in alloxan-induced diabetic rabbits. *Journal of Ethnopharmacologyl*, 85: 201-6.

- Spiller, H. A. and Sawyer, T. S. (2006). Toxicology of oral antidiabetic medication. *American*
- Sundararajan, R. N., Venkatesan, N. A., Mukherjan, K., Saha, B. P., Bandyopadhyan, A. and Mukherree, P. K. (2006). Cytisus scoparius link- a natural antioxidants. *BMC complement and Alternative Medicine*, 6(8): 1-7.
- Yakubu, M. T., Akanji, M. A., Oladiji, A. T. (2007). Haematological evaluation in male albino rats following chronic administration of aqueous extract of *Fadogia agrestis* stem. *Pharmacological Magazine*, 3: 34.
- Zozulinska, D., Wierusz-wysocka, B., Byks, H., Majchrazak, A., Grykiel, K. and Wysocki, H. (1998). Glicazide reduces plasma hydrogen peroxide level in patient with type II diabetes. *Medicinal Science Monograph*, 4(1): 66-71.

Table 1: Effect of administration of 3 solvent extract of *chrysophyllum albican* leaves on body weight in alloxan-induced diabetic mice

Groups	Body weight (g) initial	Body weight (g) during treatment	Body weight (g)
A (Normal control)	27.95	32.65	35.55
<b>B</b> (Untreated Diabetic control)	29.15	31.05	33.80
<b>Petroleum Ether extract</b>			
C( 500mg/kg)	28.40	28.70	33.80
Ethanolic extract			
<b>D</b> (500mg/kg)	28.25	28.55	28.45
Chloroform extract			
<b>E</b> (500mg/kg)	30.55	30.40	31.45
F (Glibenclamide)	27.45	26.80	26.60

Group A(Normal control) Group B (Untreated Diabetic control), Groups C, D and E were treated with 500 mg/kg body weight of petroleum ether, ethanol and chloroform extract of Chrysopyllum albidum respectively while Group F (Glibenclamide)

Groups	blood glucose level baseline (mg/al)	une (mg/aı)	DIOOU GIUCOSE IEVEI IIIIUAI (IIIg/UI)		Dioou glucose level fillal (Ilig/ul)
A (Normal control)	117.50		93.50		96.50
B (Untreated Diabetic control)	92.00		368	38	380.00
Petroleum Ether extract					
C(500mg/kg)	69.50		314	98	68.00
Ethanolic extract					
<b>D</b> (500mg/kg)	98.00		388	76	76.00
Chloroform extract					
E (500mg/kg)	61.00		334.00	69	69.00
F (Glibenclamide)	74.00  mg/dl		244.00 mg/dl	81	81.00 mg/dl
and chloroform extract of Chrysopyllum albidum respectively while Group F (Glibenclamide) Table 3. Effect of the Chassenhalling albidum loof extract on becaustological values in alloven induced of diabetic vets	vllum albidum respectively wh	iile Group F (C	<i>llibenclamide)</i> soluce in olloven induced	d of diahotio wats	
Groups	RBC (X10 <sup>12</sup> /L)	PCV (%)	Hb(g/dL)	WBC (X10 <sup>9</sup> /L)	L) Platelet (X10 <sup>9</sup> /L)
A (Normal control)	$7.85\pm0.03^{a}$	43.50±0.87 <sup>cd</sup>	$15.60\pm0.64^{bc}$	$11.05\pm0.20^{\circ}$	
<b>B</b> (Untreated Diabetic control)	$680+0.06^{\circ}$	42 50+0 29 <sup>d</sup>	14 25+0 09°	$11 30+0.35^{\circ}$	570.00+11.55°
Pet Ether extract					
C( 500mg/kg)	$6.75 \pm 0.03^{b}$	$41.50{\pm}0.87^{\rm b}$	$14.20{\pm}0.35^{\rm ab}$	$12.80{\pm}0.35^{\rm b}$	$470.00{\pm}40.41^{\circ}$
Ethanolic extract					
D (500mg/kg)	5.35±0.03°	$40.50\pm0.29^{b}$	$13.65 \pm 0.49^{ab}$	$11.20\pm0.35^{\circ}$	$465.00\pm 20.21^{\rm b}$
Chloroform extract					
E (500mg/kg)	$4.20{\pm}0.13^{d}$	$38.50\pm0.13^{\rm bc}$	$13.30\pm0.13^{bc}$	$14.60\pm0.13^{a}$	$430.00\pm10.13^{\circ}$
F (Glibenclamide)	$7.60{\pm}0.17^{a}$	$44.00\pm0.58^{a}$	$14.90\pm0.98^{a}$	$11.60\pm0.23^{\circ}$	$545.00 \pm 14.43^{a}$
Values are expressed as abc Means in the same row with different superscript are significantly different $p \le 0.05$ Table 4: Effect of the <i>Chrysophyllum albidum</i> leaf extract on liver function parameters in alloxan-induced of diabetic rats	<i>in the same row with differen um albidum</i> leaf extract on liv	<i>ut superscript a</i> ver function pa	<i>e significantly different p</i> rameters in alloxan-indu	$0 \le 0.05$ iced of diabetic ra	ts
Groups	AST (IU/L)	ALT (IU/L)	ALP (IU/L)	Bilirubin (mg/dl)	(dl) Protein (g/dl)
A (Normal control)	$42.15 \pm 3.95^{b}$	$30.25\pm2.28^{\circ}$	$21.30\pm1.73^{bc}$	$1.55 \pm 0.03^{ab}$	$5.05{\pm}0.09^{a}$
B (Untreated Diabetic control)	$48.30{\pm}0.58^{a}$	$40.60{\pm}0.64^{a}$	$25.00{\pm}2.60^{ab}$	$1.70{\pm}0.06^{a}$	$4.25\pm0.03^{b}$
Pet Ether extract					
C( 500mg/kg)	$44.00 \pm 6.52^{a}$	$39.15\pm 2.40^{ab}$	$23.10 \pm 1.79^{bc}$	$1.10\pm0.06^{\circ}$	$4.00{\pm}0.12^{b}$
<b>Ethanolic extract</b>					
<b>D</b> (500mg/kg)	$47.35\pm5.40^{a}$	$30.10{\pm}0.06^{\circ}$	$25.75\pm0.49^{a}$	$1.30{\pm}0.06^{\mathrm{bc}}$	$5.10 \pm 0.29^{a}$
Chloroform extract					
E (500mg/kg)	$47.35\pm8.46^{a}$	$33.25\pm4.36^{bc}$	$21.45\pm0.66^{ m bc}$	$1.55 \pm 0.14^{ab}$	$4.85 \pm 0.20^{a}$
F (Glibenclamide)	41 70+0 13 <sup>b</sup>	28 00+0 13°	$18\ 90+0\ 13^{\circ}$	$1 50+0 13^{a}$	$4 60+0 13^{\circ}$

-----

---

Table 5: Effect of the <i>Chrysophyllum albican</i> leaf extract on kidney function parameters in alloxan-induced of diabetic rats	vllum albican leaf	extract on kidney fund	ction parameters	in alloxan-indu	iced of diabetic r	ats	
Groups	Urea(mg/dl)	Urea(mg/dl) Creatinine (mg/dl) Na <sup>+</sup> (mEq/L) K <sup>+</sup> (mEq/L) Cl <sup>+</sup> (mEq/L) HCO <sub>3</sub> -(mmol/L) Ca <sup>2+</sup>	Na <sup>+</sup> (mEq/L)	K <sup>+</sup> (mEq/L)	Cl <sup>+</sup> (mEq/L)	HCO <sub>3</sub> <sup>-</sup> (mmol/L)	Ca <sup>2+</sup>
A (Normal control)	$25.50\pm1.44^{b}$	$0.76{\pm}0.01^{ m b}$	$198.00\pm1.15^{d}$ $2.50\pm0.06^{d}$	$2.50{\pm}0.06^{d}$	$120.12\pm0.12^{a}$	$17.00{\pm}2.06^{a}$	$6.10 \pm 0.17^{a}$
B (Untreated Diabetic control)	$54.00 \pm 1.15^{a}$	$1.76{\pm}0.01^{a}$	242.00±2.89 <sup>b</sup>	$5.45\pm0.03^{a}$	125.56±4.03 <sup>a</sup>	$17.30{\pm}3.06^{a}$	7.10±0.29 <sup>a</sup>
C(500mg/kg) Fthenolic extract	$49.00{\pm}4.04^{a}$	$1.65\pm0.02^{a}$	$156.50{\pm}2.60^{f}$	$3.70{\pm}0.23^{b}$	$120.81{\pm}6.06^{a}$	$17.50{\pm}4.06^{a}$	$7.00{\pm}0.46^{a}$
D (500mg/kg)	$51.50 \pm 4.33^{a}$	$1.64{\pm}0.02^{a}$	$208.50\pm 2.02^{a}$	$5.25{\pm}0.32^{a}$	$115.20\pm 5.03^{a}$	$17.91 \pm 3.03^{a}$	$6.90{\pm}0.35^{a}$
E (500mg/kg)	$23.00{\pm}1.02^{b}$	$0.82 \pm 0.02^{b}$	256.00±2.22ª	3.00±0.22 <sup>cd</sup>	$121.71\pm 5.02^{a}$	17.72±2.02 <sup>a</sup>	6.40±0.22 <sup>a</sup>
F (Glibenclamide)	$48.00{\pm}3.46^{a}$	$1.63{\pm}0.02^{a}$	$183.50\pm 2.60^{\circ}$	$183.50\pm 2.60^{\circ}$ $3.10\pm 0.00^{\circ}$	$115\pm4.03^{-1}$	$16.5\pm 2.03^{a}$	$6.90\pm0.35$ <sup>a'</sup>
Values are expressed as Means SEM. Means marked $*$ in the same column are significantly different from diabetic control at $p \leq 0.05$	SEM. Means mark	ed * in the same colum	n are significanti	v different from	diabetic control	$at \ p \le 0.05$	