■NIGERIAN JOURNAL OF BIOTECHNOLOGY

Nig. J. Biotech. Vol. 36 (1): 94-102 (June 2019) ISSN: 0189 1731 Available online at http://www.ajol.info/index.php/njb/index and www.biotechsocietynigeria.org DOI: https://dx.doi.org/10.4314/njb.v36i1.13



# Antagonistic effect of *Bacillus thuringiensis* for the control of bacterial wilt of tomato (*Lycopersicon esculentum* Mill)

<sup>1</sup>Akintokun, A. K., \*<sup>2</sup>Ojesola, C. O., <sup>3</sup>Akintokun, P. O. and <sup>1</sup>Oloyede, A. R. <sup>1</sup>Microbiology Department, Federal University of Agriculture, Abeokuta, Ogun State, Nigeria. <sup>2</sup>Biotechnology Centre, Federal University of Agriculture, Abeokuta, Ogun State, Nigeria. <sup>3</sup>Department of Plant Physiology and Crop Production, Federal University of Agriculture, Abeokuta, Ogun State, Nigeria.

Copyright resides with the authors in terms of the Creative Commons License 4.0. (See <u>http://creativecommons.org/licenses/by/4.0/</u>). Condition of use: The user may copy, distribute, transmit and adapt the work, but must recognize the authors and the Nigerian Journal of Biotechnology.

### Abstract

Tomato is affected by a lot of diseases including bacterial wilt caused by *Ralstonia* solanacearum (Rs), resulting in fruit yield loss. Chemical control of this disease is not advisable because of hazardous effects on human and environment, hence, the need to use biocontrol agents such as *Bacillus thuringiensis* (Bt). This study investigated the potential of Bt in the control of Rs under screenhouse condition. Six treatments with three replications in completely randomized design were carried out. Agronomic parameters were measured and data collected were analyzed using one - way analysis of variance. Means were then separated using Duncan's Multiple Range Test at  $p \le 0.05$ . Inoculation of Bt and Rs simultaneously significantly suppressed the incidence of bacterial wilt and promoted plant growth. Higher plant heights (72.70 - 111.00 cm), stem girth (1.63 - 1.90 cm) and number of leaves (103.33 - 125.33) were recorded for plants inoculated with Bt at 9th week after planting compared to plants inoculated with Rs alone. Highest disease incidence was recorded in plants inoculated with Rs alone (100 %), while plants inoculated with Bt had the highest disease reduction (67%). This study revealed that Bt may be used in the control of bacterial wilt of tomato.

Key words: Bacillus thuringiensis, screenhouse, Bacterial wilt, Ralstonia solanacearum, Tomato.

### \*Corresponding author: idit01@yahoo.com Tel: +2348027320685

### Introduction

Tomato (*Lycopersicon esculentum* Mill) belongs to the Solanaceae family, it is one of the most important vegetables consumed in the world (Olaniyi et al., 2010). The fruits are eaten raw as salads or ground into a paste and are an important source of antioxidants, minerals and vitamins (Frusciante et al., 2007. Soil-borne pathogens inflict a lot of diseases and infections on tomato (Babalola and Glick, 2012). Such diseases include Bacterial wilt, root-knot nematodes disease, early blight, late blight and *Fusarium* wilt (Ajilogba et al., 2013).

Bacterial wilt of tomato is a systemic vascular disease caused by *Ralstonia solanacearum* (Abeer and Hend, 2013). A soilborne plant bacterial pathogen notorious for its lethality, persistence, complex subspecies, wide host range, and broad geographic distribution (Elphinstone, 2005). Bacterial wilt is a common disease in tropical, subtropical and some temperate regions of the world (Fegan and Prior, 2005), it is endemic in most tomato-growing areas of Nigeria, causing 60 to 100% loss in yield (Popoola et al., 2015).

Soil fumigation, suppressive soil, short rotation, and resistant cultivars have been suggested as integrated control strategies for bacterial wilt (French, 1994). However, while chemical soil disinfection can temporarily eradicate most microbial flora, pesticideresistant pathogens may rebound, causing even more damage than those originally targeted for control (Gamliel et al., 2000). Therefore,

environment - friendly biopesticide and beneficial microorganisms are alternatives to chemical pesticides (Bailey and Lazarovits, 2003; Céline et al., 2007; Weller, 2007; Ongena and Jacques, 2008). Biological control of R. solanacearum has been achieved using several Beneficial Microorganisms which include Bacillus spp. (Ji et al., 2008), Pseudomonas spp. (Ramesh et al., 2009; Vanitha et al., 2009), avirulent mutants of R. solanacearum (McLaughlin and Sequeira, 1988; Frey et al., 1994, Acinetobacter. Enterobacter spp. (Xue et al., 2009), Stenotrophomonas maltophilia (Messiha et al., 2007) and Actinomycetes (Tan et al., 2006) under laboratory and/or greenhouse conditions. In vitro analysis revealed that four Bacillus isolates viz B. amyloliquefaciens, B. cereus, B. *pumilus* and *B. subtilis* inhibited the growth of Fusarium solani significantly (Ajilogba et al., 2013).

Bacillus thuringiensis (BT), a spore-forming bacterium is well known for its insecticidal properties associated with its ability to produce crystal inclusions during sporulation. These inclusions are proteins encoded by cry genes and have shown to be toxic to a variety of insects and other organisms like nematodes and protozoa (Konecka et al., 2007). Recently, in Japan, Bacillus thuringiensis attracted considerable attention as a potential biological control agent for the suppression of R. solanacearum growth and the development of wilt symptoms in tomato plants (Hyakumachi et al., 2013). Hence the need to investigate the antagonistic activity of native Bacillus thuringiensis in the control of bacterial wilt affecting tomato plants in Nigeria.

### **Materials and methods**

The study was conducted in the Screenhouse of College of Plant Science, Federal University of Agriculture, Abeokuta, Ogun State. The study location lies within the savanna agroecological zone of South-west Nigeria (Latitude 7°N, Longitude 3.5°E in Odeda Local Government Area of Ogun State).

### Preparation of seedlings and inoculum

Beske tomato seeds (susceptible to bacterial wilt disease of tomato) were raised in the nursery and transplanted after two weeks into 5kg soil (sterilized in a hot air oven at 120°C for 2 hours) in plastic buckets in the screenhouse. Ralstonia solanacearum (pathogen) was isolated from diseased tomato plant showing symptoms of bacterial wilt as described by Zubeda and Hamid (2011) and Bacillus thuringiensis (antagonist) was isolated from cultivated soil sample obtained from the Directorate of University Farms, Federal University of Agriculture, Abeokuta, according to Palma (2015). Pathogenicity test was carried out on *R. solanacearum* according to the methods described by Abeer and Hend, 2013; Hyakumachi et al., 2013 and Elsharkawy et al., 2015.

The two isolates were confirmed using biochemical and molecular methods. The pathogen and antagonist were grown separately on nutrient agar plates for two days at  $28\pm2^{\circ}$ C after which growth was scrapped into sterile distilled water in microcentrifuge tubes. The cells of the pathogen were removed by centrifugation at 7,000 rpm for 10 minutes; pellets were resuspended in sterile distilled water and adjusted to a final density of  $1 \times 10^{7}$  CFU / ml (Elsharkawy et al., 2015). Antagonist suspension was adjusted to a final density of  $1.8 \times 10^{8}$ CFU/ml. The suspensions were applied to 3-week old tomato seedlings. The experimental design used was Completely Randomized Design (CRD), replicated three times. Experimental treatments

Inoculation of *Bacillus thuringiensis* and *Ralstonia solanacearum* simultaneously (B+R)

Inoculation of *Bacillus thuringiensis* first and *Ralstonia solanacearum* a week after (Bf)

Inoculation of *Ralstonia solanacearum* first and Bacillus thuringiensis a week after. (Rf)

Inoculation of *Ralstonia solanacearum* only (Ro)

Inoculation of Bacillus thuringiensis only (Bo)

Inoculation with sterile water. (control)

The following agronomic data were collected at 3, 4, 5, 6, 7, 8, 9, 10, 11 and 12 weeks after planting: plant height (cm), stem girth (cm), fresh and dry root weights (g), number of flowers and number of leaves. Disease incidence and reduction were determined at the end of the experiment.

%DI = (NDP×100)/NPA

Where % DI is percentage disease incidence, NDP is the number of diseased plants and NPA is the number of plants assessed.

%DR =(C-T)/C

Where %DI is percentage disease reduction, C is percentage disease reduction in untreated plants, T is percentage disease incidence in treated plants

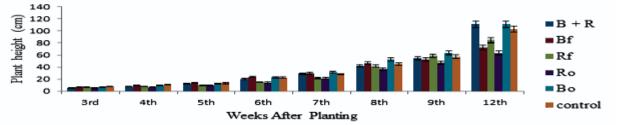
### Data analysis

Data obtained were analyzed using Statistical Package for Social Sciences (SPSS) version 20.0 for windows (SPSS, Chicago IL, USA). Descriptive statistics (mean and standard deviation) and analysis of variance (one-way) with means separated using Duncan multiple range test and level of significance was considered as  $p \le 0.05$ .

### Results

Effect of Bacillus thuringiensis on Tomato Plant height

Plant height increased with weeks after planting in all the treatments with highest plant height recorded in treatments B+ R (*Bacillus thuringiensis* and *Ralstonia solanacearum* applied at the same time) and Bo (*Bacillus thuringiensis* only) having 111.00 cm. The least plant height was recorded in treatment Ro that had only the pathogen with 63.67 cm (Figure 1).



### **Figure 1: Effect of treatments on tomato plants height at different weeks after planting**

B + R – Antagonist (Bt) and pathogen (Rs) applied simultaneously

Bf – Antagonist applied first, pathogen applied a week after

Rf – Pathogen applied first, antagonist applied a week after

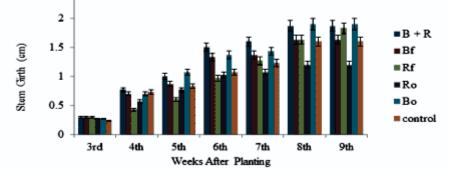
Ro – Pathogen only

Bo – Antagonist only

Control - Sterile water

Effect of Bacillus thuringiensis on Tomato Plant stem girth

Stem girth also increased with weeks after planting (WAP) in all the treatments until when the experiment was terminated. Treatment Bo where the antagonist was applied alone, had the highest stem girth of 1.90 cm and the least was recorded in treatment Ro where only the pathogen was inoculated having 1.20 cm (Figure 2)



### Figure 2: Effect of treatments on stem girth of tomato plants at different weeks after planting

KEY

B + R – Antagonist (Bt) and pathogen (Rs) applied simultaneously

Bf – Antagonist applied first, pathogen applied a week after

Rf – Pathogen applied first, antagonist applied a week after

Ro – Pathogen only

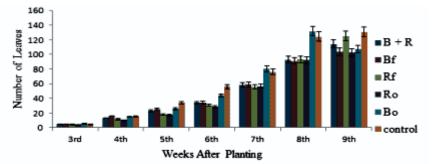
Bo – Antagonist only

Control - Sterile water

Effect of Bacillus thuringiensis on number of leaves of Tomato Plant

The number of leaves increased with weeks after planting across treatments. The highest number of leaves was recorded in control

(130) whereas the least was recorded in Ro having only pathogen (102) (Figure 3).



## Figure 3: Effect of treatments on the number of leaves of tomato plants at different weeks after planting KEY

B + R - Antagonist (Bt) and pathogen (Rs) applied simultaneously

Bf - Antagonist applied first, pathogen applied a week after

Rf – Pathogen applied first, antagonist applied a week after

Ro – Pathogen only

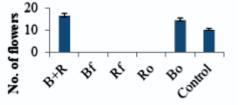
Bo – Antagonist only

Control - Sterile water

*Effect of Bacillus thuringiensis on number of flowers of tomato plants* 

At the end of the experiment, treatment B + R (*Bacillus thuringiensis* and *Ralstonia solanacerum* applied at the same time) had the

highest number of flowers (17) as shown in figure 4  $\,$ 



treatments

### **Figure 4: Effect of treatments on the number of flowers** KEY

B + R – Antagonist (Bt) and pathogen (Rs) applied simultaneously

Bf - Antagonist applied first, pathogen applied a week after

Rf – Pathogen applied first, antagonist applied a week after

Ro – Pathogen only

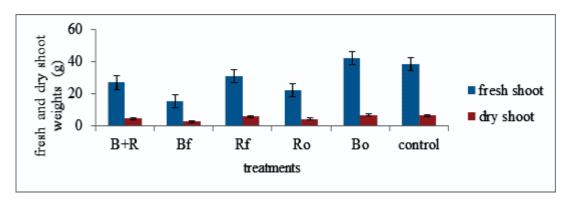
Bo – Antagonist only

Control - Sterile water

*Effect of Bacillus thuringiensis on Fresh and Dry Shoot Weight of tomato plant* 

Highest fresh and dry shoot weights were observed in treatment Bo having only

Bacillus thuringiensis while the least was observed in treatment Bf, where Bacillus thuringiensis was applied first and then Ralstonia solanaearum applied a week after (figure 5).



**Figure 5: Effect of treatments on fresh and dry shoot weights** KEY

B + R – Antagonist (Bt) and pathogen (Rs) applied simultaneously

Bf – Antagonist applied first, pathogen applied a week after

Rf – Pathogen applied first, antagonist applied a week after

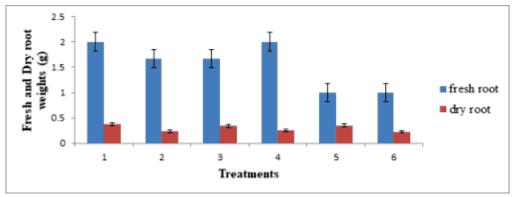
Ro – Pathogen only

Bo - Antagonist only

Control - Sterile water

*Effect of Bacillus thuringiensis on Fresh and Dry root Weight of tomato plant* 

Treatments B+R (*Bacillus thuringiensis* and *Ralstonia solanacearum* applied at the same time) and Ro with pathogen only had the highest fresh root weight while the least fresh root weight was observed in treatments Bo (*Bacillus thuringiensis* only) and control. The highest dry root weight was observed in treatment B+R (*Bacillus thuringiensis* and *Ralstonia solanacearum* applied at the same time) while the least was observed in control (Figure 6).



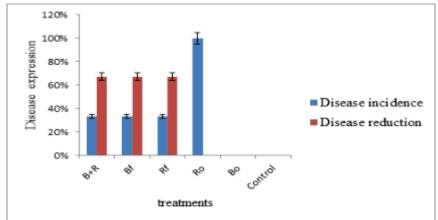
#### **Figure 6: Effect of treatments on fresh and dry root weights** KEY

B + R – Antagonist (Bt) and pathogen (Rs) applied simultaneously Bf – Antagonist applied first, pathogen applied a week after Rf – Pathogen applied first, antagonist applied a week after Ro – Pathogen only Bo – Antagonist only Control - Sterile water

### Disease Incidence and Reduction

Disease incidence was highest in treatment Ro (100 %) while the least was recorded in treatments Bo and control (0%). Percentage disease reduction was highest in

treatments having *Bacillus thuringiensis* (B+R, Bf and Rf) with 67% while the least was recorded in treatment having only *Ralstonia solanacearum* (Ro) with 0% (Figure 7).



#### **Figure 7: Disease Incidence and Reduction** KEY

B + R - Antagonist (Bt) and pathogen (Rs) applied simultaneously

Bf - Antagonist applied first, pathogen applied a week after

### Discussion

Bacterial wilt caused by *Ralstonia solanacearum* is a dangerous disease in tropical, subtropical and some temperate regions of the World (Elsharkawy et al., 2015). A species complex with a wide host range infecting more than 450 crop species belonging to more than 50 families (Swanson et al., 2005; Aliye et al., 2008). Control methods against various diseases caused by this phytopathogen include the use of resistant varieties, crop rotation, biological control e.t.c. (Elsharkawy et al., 2015).

*Bacillus thuringiensis* a Gram-positive spore-forming bacterium is commonly known as an important biocontrol agent for the control of many agricultural insect pests and vectors of human diseases (Chattopadhyay et al., 2004; Noura et al., 2007). This is owing to its ability to produce during sporulation characteristic proteinaceous crystalline toxins (deltaendotoxins) (Schnepf et al., 1998). It has also attracted considerable attention as a potential biocontrol agent for the suppression of plant diseases (Reyes-Ramirez et al., 2004; Zhou et al., 2008).

Results from this study revealed that inoculation of Bacillus thuringiensis and Ralstonia solanacearum into apparently healthy Beske tomato seedlings under screenhouse condition significantly reduced the manifestation of bacterial wilt disease. Higher growth rates were recorded for plants height, number of leaves, number of flowers and shoot weights across treatments treated with Bacillus thuringiensis. This is in agreement with the previous studies of Armada et al. (2015) and Kassogue et al. (2016) who found Bt to improve plant growth significantly. Disease suppression was also higher in treatments B+R (Bacillus thuringiensis applied first and Ralstonia solanacerum applied a week after), Bf (Bacillus thuringiensis applied first and *Ralstonia* solanacearum applied a week after), Rf (Ralstonia solanacearum applies first and Bacillus thuringiensis applied a week after) and Bo (only Bacillus thuringiensis applied). This is in agreement with Mitsuro et al. (2012) who found *Bacillus thuringiensis* to significantly suppress the growth of Ralstonia solanacearum and the development of wilt symptoms in tomato plants. Suppressive ability of Bacillus thuringiensis could be due to the production of several compounds, including b-exotoxins, antibiotics, degrading enzymes, bacteriocins, and a signal molecule in the bacterial quorumsensing system as opined by Dong et al. (2002), Cherif et al. (2003), Murphy et al. (2003), Cherif et al.(2008), Zhou et al.(2008) and Raddadi et al.(2009).

### Conclusion

From this study, it can be concluded that native *Bacillus thuringiensis* isolated from cultivated soil can be used in the biological control of the soil-borne plant bacterial pathogen, *Ralstonia solanacearum*, the causal agent of bacterial wilt of tomato and also to improve the overall health of tomato plants under screenhouse condition. Further study should be carried out on field trials under different local environmental conditions. After successful field trials, farmers should be encouraged to embrace the use of *Bacillus thuringiensis* in place of chemicals.

### References

Abeer, H.M. and Hend, A.H. (2013). Suppression of bacterial wilt disease of tomato plants using some bacterial strains. Life Sci. 10 (3):1732-1741.

Ajilogba, C. F., Babalola, O. O. and Ahmad, F. (2013). Antagonistic Effects of *Bacillus* species in Biocontrol of Tomato *Fusarium* Wilt. Ethno. Med. 7(3): 205-216.

Aliye, N., Fininsa, C. and Hiskias, Y. (2008). Evaluation of rhizosphere bacterial antagonists for their potential to bioprotect potato (*Solanum tuberosum*) against bacterial wilt (*Ralstonia solanacearum*). Biol. Control 47: 282–288.

Armada, E., Probanza, A., Roldan, A., and Azcon, R. (2015). Native plant growth-promoting bacteria *Bacillus thuringiensis* and mixed or individual mycorrhizal species improved drought tolerance and oxidative metabolism in Lavandula dentate plants. J. Plant Physiol.192:1-12.

Arora N., Ahmad T., Rajagopal R., and Bhatnagar R.K. (2003).A constitutively expressed 36 kDa exochitinase from *Bacillus thuringiensis* HD-1. Biochem.Biophy. Res. Commun. 307: 620–625.

Babalola, O.O. and Glick B.R. (2012). Indigenous African agriculture and plant-associated microbes: current practice and future transgenic prospects. Sci. Res. Essays. 7: 2431- 2439.

Bailey, K. L. and Lazarovits, G. (2003). Suppressing soil-borne diseases with residue management and organic amendments. Soil Till. Res. 72: 169–180.

Céline, J., Francois, V., Alabouvette, C., Véronique, E. H., Thierry, M. and Christian, S. (2007). Soil health through soil disease suppression: which strategy from descriptors to indicators. Soil Biol. Biochem. 39: 1–23.. Chattopadhyay, A., Bhatnagar, N.B. and Bhatnagar, R. (2004). Bacterial insecticidal toxins.Crit. Rev. Microbiol. 30: 33–54.

Cherif, A., Chehimi, S., Limem, F., Hansen, B.M., Hendriksen, N.B. Daffonchio, D. and Boudabous, A. (2003). Purification and characterization of the novel bacteriocin entomocine 9, and safety evaluation of its producer, *Bacillus thuringiensis* subsp. *entomocidus* HD9. J. Appl. Microbiol. 95: 990–1000.

Cherif A., Rezgui, W., Raddadi, N., Daffonchio, D. and Boudabous, A.(2008).Characterization and partial purification of entomocin 110, a newly identified bacteriocin from *Bacillus thuringiensis* subsp. *entomocidus* HD110. Microbiol. Res. 163:684–692.

Dawoud, M. E. Kamel, Z., Hanaa, A. and Farahat, M. G. (2012). Growth promotion and biocontrol of leaf spot and leaf speck diseases in tomato by *Pseudomonas* spp. Egypt. J. Exp. Biol. 8:61–70.

Dong, Y. H., Xu, J. L., Li, X. Z. and Zhang, L. H. (2000). AiiA, an enzyme that inactivates the acylhomoserine lactone quorum-sensing signal and attenuates the virulence of Erwinia carotovora. Proceedings of the National Academy of Science USA, 97: 3526–3531.

Dong, Y. H., Gusti, A. R., Zhang, Q, Xu, J. L. and Zhang, L. H. (2002). Identification o.f quorumquenching N-acyl-homoserine lactonases from *Bacillus* species. Appl. Environ. Microbiol. 68:1754–1759.

Elphinstone, J. G. (2005). The current bacterial wilt situation: a global overview. Pp. 9-28. In: Allen, C. Prior, P. and Hayward, A. C. (eds.) Bacterial Wilt disease and the *Ralstonia solanacearum* complex. Phytopathol. St. Paul, MN.

Elsharkawy, M. M., Hassan, N., Ali, M., Mondal, S. N. and Hyakumachi, M. (2014).Effect of zoysia grass rhizosphere fungal isolates on disease suppression and growth promotion of rice seedlings. Acta Agr. Scand. B- S.P.64:135–140.

Elsharkawy, M. M., Nakatani, M., Nishimura, M., Arakawa, T., Shimizu, M. and Hyakumachi, M. (2015). Control of tomato bacterial wilt and rootknot diseases by *Bacillus thuringiensis* CR-371 and *Streptomyces avermectinius* NBRC14893 Acta Agric. Scandi. Sec. B and Soil Pla. Sci.65 (6): 575-580.

Elsharkawy, M. M., Shimizu, M., Takahashi, H.,

Hyakumachi, M. (2012a).Induction of systemic resistance against Cucumber mosaic virus by *Penicillium simplicissimum* GP17-2 in arabidopsis and tobacco. Plant Pathol. J. 61:964–976.

Elsharkawy, M. M., Shimizu, M., Takahashi, H., Ozaki, K., Hyakumachi, M. (2013).Induction of systemic resistance against Cucumber mosaic virus in *Arabidopsis thaliana* by *Trichoderma asperellum* SKT-1.Plant Pathol. J. 29:193–200.

Elsharkawy, M. M., Takahashi, H., Hyakumachi, M. M. (2012b). The plant growth-promoting fungus *Fusarium equiseti* and the arbuscular mycorrhizal fungus *Glomus mosseae* induce systemic resistance against Cucumber mosaic virus in cucumber plants. Plant Soil. 361:397–409.

Fegan, M. and Prior, P. (2005). How complex is the "*Ralstonia solanacearum* species complex"? In: Allen, C., Prior, P., Hayward, A.C. (eds.). Bacterial wilt disease and the *Ralstonia solanacearum* species complex. APS Press, St. Paul, MN, USA.

French, E. R., (1994). Integrated control of bacterial wilt of potato. CIP Circular 20: 8–11.

Frey, P., Prior, P., Marie, C., Kotoujansky, A., Demery, D. T. and Trigalet, A., (1994).Hrp– Mutants of *Pseudomonas solanacearum* as potential biocontrol agents of tomato bacterial wilt. Appl. Environ. Microbiol.60:3175–3181.

Frusciante, L., Carli, P., Ercolano, M. R., Pernice, R., Di Matteo, A., Forgliano, V. and Pellegrini, N. (2007). Antioxidant nutritional quality of tomato. Mol. Nutr. Food Res. 51(5):609–617.

Gamliel, A., Austerweil, M., Kritzman, G., (2000).Non-chemical approach to soilborne pest management – Organic amendments. Crop Prot. 19: 847–853.

Haruna, S. G., Adebitan, S. A. and Gurama, A. U. (2012). Field evaluation of compost extracts for suppression of *Fusarium* wilt of tomato caused by *Fusarium oxysporum* fsp.*lycopersici*. Int. J. Agro. and Agric. Res. 2(4):7-17.

Hassan, N., Elsharkawy, M. M., Shivanna, M. B., Meera, M. S. and Hyakumachi, M. (2014). Elevated expression of hydrolases, oxidase, and lyase in susceptible and resistant cucumber cultivars systemically induced with plant growthpromoting fungi against anthracnose. Acta Agr. Scand. B- S. P. 64:155–164. Hyakumachi, M., Nishimura, M., Arakawa, T., Asano, S., Shigenobu, Yoshida, S., Tsuchima, S., and Takahashi, H. (2013). *Bacillus thuringiensis* Suppresses Bacterial wilt Disease Caused by *Ralstonia solanacearum* with Systemic Induction of Defense-Related Gene Expression in Tomato. Microbes and Environ. 28 (1): 128–134.

Ji, X., Lu, G., Gai, Y., Zheng, C., Mu, Z., (2008).Biological control against bacterial wilt and colonization of mulberry by an endophytic *Bacillus subtilis* strain.FEMS Micro. Eco. 65: 565–573.

Kassogue, A., Dicko, A. H., Traore, D., Fane, R., Valicente, F. H. and Banana, A. H. (2016). *Bacillus thuringiensis* strains isolated from Agricultural Soils in Mali Tested for their Potentiality on Plant Growth Promoting Traits. Br. Microbiol. Res. J.14(3):1-7.

Konecka, E., Kaznowski, A., Ziemnicka, J., Ziemnicki, K. (2007). Molecular and phenotypic characterization of *Bacillus thuringiensis* isolated during epizootics in *Cydia pomonella* L. J. Invertebr. Pathol. 94: 56-63.

Mc Laughlin, R. J. and Sequeira, L. (1988). Evaluation of an avirulent strain of *Pseudomonas solanacearum* for biological control of bacterial wilt of potato. American Potato J.65 (5):255-268.

Messiha, N., van Diepeningen, A., Farag, N., Abdallah, S., Janse, J., van Bruggen, A. (2007). *Stenotrophomonas maltophilia*: a new potential biocontrol agent of *Ralstonia solanacearum*, causal agent of potato brown rot. Eur. J. Plant Pathol. 118: 211–225.

Mitsuro, H., Mitsuyoshi, N., Tatsuyuki, A., Shinichiro, A., Shigenobu, Y., Seitya, T. and Hedeki, T. (2012). *Bacillus thuringiensis* suppresses Bacterial wilt caused by *Ralstonia solanacearum* with Systemic Induction of Defence-Related Gene Expression in Tomato. Microbes and Environ. 28(1):128-134

Murphy, J. F., Reddy, M. S., Ryu, C. M., Kloepper, J. W. and Li, R. (2003). Rhizobacteria-mediated growth promotion of tomato leads to protection against Cucumber mosaic virus. Phytopathology. 93:1301–1307.

Nguyen M. T., Ranamukhaarachchi, S. L. and Hannaway, D. B. (2011). Efficacy of antagonist strains of *Bacillus megaterium*, *Enterobacter cloacae*, *Pichia guilliermondii* and *Candida*  *ethanolica* against bacterial wilt disease of tomato. J. Phytopathol. 3: 1–10.

Noura, R., Ameur C., Abdellatif, B., and Daniele, D. (2007). Screening of plant growth-promoting traits of *Bacillus thuringiensis*. Ann. Microbiol. 58 (1): 57-52.

Olaniyi, J. O. Akanbi, W. B. Adejumo, T. A. and Akande, O. G.(2010). Growth, fruit yield and nutritional quality of tomato varieties. Afr. J. Food Sci. 4 (96): 398-402.

Ongena, M. and Jacques, P. (2008). *Bacillus lipopeptides*: versatile weapons for plant disease biocontrol. Trends Microbiol. 16:115-120.

Popoola, A. R., Ganiyu, S. A., Enikuomehin, O. A., Bodunde, J. G., Adedibu, O. B., Durosomo, H. A. and Karunwi, O. A. (2015). Isolation and Characterization of *Ralstonia solanacearum* Causing Bacterial Wilt of Tomato in Nigeria. Niger. J. Biotechnol. 29: 1 – 10.

Raddadi N., Cherif A., Ouzari H., Marzorati M., Brusetti L., Boudabous A., Daffonchio D. (2007). *Bacillus thuringiensis* beyond insect biocontrol: plant growth promotion and biosafety of polyvalent strains. Ann. Microbiol. 57: 481–494.

Raddadi, N., Belaouis, A., Tamagnini, I., Hansen, B. M., Hendriksen, N. B., Boudabous, A., Cherif, A. and Daffonchio, D. (2009).Characterization of polyvalent and safe *Bacillus thuringiensis* strains with potential use for biocontrol. J. Basic Microbiol. 49:293–303.

Ramesh, R., Joshi, A., Ghanekar, M., (2009).Pseudomonads: major antagonistic endophytic bacteria to suppress bacterial wilt pathogen, *Ralstonia solanacearum* in the eggplant (*Solanum melongena* L.). World J. Microbiol. and Biotechnol. 25: 47–55.

Reyes-Ramirez, A., B. I. Escudero-Abarca, G. Aguilar-Uscanga, P. M. Hayward-Jones, and J. Eleazar-Barbozacorona. (2004). Antifungal activity of *Bacillus thuringiensis* chitinase and its potential for the biocontrol of phytopathogenic fungi in soybean seeds. J. Food Sci.69:5:131-134.

Saldajeno, M. G. B., Naznin, H. A., Elsharkawy, M. M., Shimizu, M. and Hyakumachi, M. (2014). Enhanced resistance of plants to disease using *Trichoderma* spp. In: Gupta, V. K., Schmoll, M., Herrera-Estrella, A., Upadhyay, R. S., Druzhinina, I., Tuohy, M., editors. Biotechnology and Biology of Trichoderma 1st ed. Amsterdam: Elsevier BV; pp. 477–493. Schnepf, E., Crickmore, N., Van Rie, J., Lereclus, D., Baum, J., Feitelson, J., Zeigler, D. R. and Dean, D. H. (1998). *Bacillus thuringiensis* and its pesticidal proteins. Microbiol. and Mol. Biol. Rev. 62: 775–806.

Senthilrajz, G., Anand, T., Kennedy, J. S., Raguchander, T. and Samiyappan, R. (2013). Plant growth-promoting rhizobacteria (PGPR) and entomopathogenic fungus bioformulation enhance the expression of defense enzymes and pathogenesis-related proteins in groundnut plants against leaf *minuteer* insect and collar rot pathogen. Physiol. Mol. Plant Pathol.82:10–19.

Swanson, J. K., Yao, J., Tans-Kersten, J., and Allen C. (2005). Behavior of *Ralstonia solanacearum* race 3 biovars 2 during latent and active infection of geranium. J. Phytopathol. 95: 136–143.

Takahashi, H., Nakaho, K., Ishihara, T., Ando, S., Wada, T., Kanayama, Y., Asano. S., Yoshida, S., Tsushima, S. and Hyakumachi, M. (2014). Transcriptional profile of tomato roots exhibiting *Bacillus thuringiensis* - induced resistance to *Ralstonia solanacearum*. Plant Cell Rep. J. 33: 99–110.

Tan, H., Cao, L., He, Z., Su, G., Lin, B., Zhou, S. (2006). Isolation of endophytic Actinomycetes

from different cultivars of tomato and their activities against *Ralstonia solanacearum* in vitro.World J. Microbiol. and Biotechnol. 22: 1275–1280.

Vanitha, S. C., Niranjana, S., Mortensen, C. and Umesha, S. (2009). Bacterial wilt of tomato in Karnataka and its management by Pseudomonas fluorescence. Biocontrol 54:685-695.

Weller, D. M. (2007). *Pseudomonas* biocontrol agents of soilborne pathogens: looking back over 30 years. Phytopathol. 97: 250–256.

Xue, Q. Y., Chen, Y., Li, S. M., Chen, L. F., Ding, G. C., Guo, D. W. and Guo, J. H. (2009). Evaluation of the strains of *Acinetobacter* and *Enterobacter* as potential biocontrol agents against *Ralstonia* wilt of tomato. Biol. Control. 48: 252–258.

Zhou Y, Choi Y.L, Sun M., Yu, Z. (2008). Novel roles of *Bacillus thuringiensis* to control plant diseases. Appl. Microbiol. Biotechnol. 80: 563–572.

Zubeda, C. and Hamid, R. (2011). Isolation and Characterization of *Ralstonia solanacearum* from infected tomato plants of Soan Skesar Valley of Punjab. Pak. J. Bot. 43 (6): 2979-2985.