

Gastroprotective Activities of the n-Hexane Fraction of *Heliotropium indicum* on Gastric Ulceration

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ABSTRACT: *Heliotropium indicum* has been established to possess gastroprotective properties. We evaluated the gastroprotective properties of the non-polar (n-hexane) fraction of *H. indicum* to establish its anti-ulcer activities. Rats of Groups I and II received physiological saline and 80 mg/kg/body weight (bw) of Indomethacin respectively. Rats of Groups III – VI received orally 100, 200 and 400 mg/kg/bw of the n – hexane fraction of methanolic extracts of dried leaves of *H. indicum* and 50 mg/kg/bw of Ranitidine respectively before the induction of gastric ulceration. One hour after administration of extract doses and Ranitidine, 80 mg/kg/bw of Indomethacin (an ulcerogen) was administered orally to rats of Groups III – VI. Morphological analyses show that administrations of 100, 200 and 400 mg/kg/bw of extract and Ranitidine resulted in percentage ulcer reductions of 45%, 50%, 75% and 95% respectively. Histological analyses of the stomach in rats of Groups II and III show erosion of mucus-secreting cells, parietal cells, gastric pit and glands. Erosion of few mucus-secreting cells, gastric pit and parietal cells were observed in rats of Group IV. Normal cytoarchitectures of the mucosa layer were observed in rats of Groups I, V and VI. Our findings show that the n-hexane fraction of *H. indicum* possessed gastroprotective properties.

Keywords: *Heliotropium indicum*, Indomethacin, Ulceration.

INTRODUCTION

Peptic ulcer occurs in sections of the gastro intestinal tract exposed to gastric acid and pepsin, such as the stomach and duodenum. The etiology is not clearly known. It results probably from an imbalance between aggressive (acid, pepsin and *H. pylori* infection) and defensive (gastric mucus and bicarbonate secretion, prostaglandins, nitric oxide, innate resistance of the mucosal cells) factors; as well as factors such as genetic, psychosomatic, humoral and vascular derangements (Ramzi, 1999). Pathological examination of gastric ulcer could be evaluated in any part of the stomach, but is most commonly obtained from the lesser curvature. Histologically, there is a break in the superficial epithelium penetrating down to the muscularis mucosa with a fibrous base accompanied with increase in inflammatory cells (Ramzi, 1999)

Heliotropium indicum belongs to the family *Boraginaceae*. Other species of *Heliotropium* include *Heliotropium bacciferum*, *Heliotropium ovalifolium* and *Heliotropium pterocarpum* (Burkhill, 1985). It grows up to 80cm high with a pleasant aroma. Its stems and leaves are covered with a fine hairy layer, and its flowers are small and grow in clusters which curve in on themselves at the tips (Burkhill, 1985) It is found in tropical and non-tropical countries bearing various names, such as *Cock's comb* (Gambia), *Indian heliotrope*, *herbe a verrues* (France), *Heliotropio-in-Indiano* (Portugal) and *Agogo Igun* or *Ogbe Akuko* (Yoruba - Nigeria) (Burkhill, 1985) Its chemical components include

pyrolizidine alkaloids, indicine-N-oxide, tannins, saponins and heliotrine (Burkhill, 1985).

Its alkaloid component confers on it anti-inflammatory, wound healing, antiseptic/antimicrobial, febrifuge, secretagogue stimulation of gall bladder and menstruation activator properties (Burkhill, 1985; Sofowora, 1993). It is most important local application is for skin lesions, wounds, abscesses, gastric and varicose ulcerations, rashes and warts (Burkhill, 1985; Sofowora, 1993). It also shows anti leukemia activity, but hepatotoxicity negates its usefulness (Burkhill, 1985; Sofowora, 1993).

The crude aqueous extract of *H. indicum* has dose-dependent morphological gastroprotective effects on indomethacin – induced gastric ulcerated mucosa in male wistar rats (Akinlolu *et al.*, 2006). The present study, therefore, investigated the gastroprotective properties of the non-polar (n-hexane) fraction of the methanolic extract of *H. indicum* in order to establish its anti-ulcer activities.

MATERIALS AND METHODS

Collection and Authentication of *H. Indicum*

H. indicum of the family *Boraginaceae* was collected in Sagamu, Ogun State in the South West of Nigeria. The plant was identified and authenticated at the Forest Research Institute of Nigeria Ibadan. The authenticated plant has the Voucher number FHI 107093.

Care and Feeding of Animals

Adult male wistar rats weighing between 90 - 150g were obtained from the colony bred of the Physiology Department of Obafemi Awolowo University, Ile-Ife, Nigeria. Animals were fed during the experiment with growers feed from Bendel Feed and Flour Mill Ltd., Nigeria. The animals were caged under standard condition in the well ventilated animal house of the Faculty of Pharmacy, Olabisi Onabanjo University, Ogun State, Nigeria at room temperature (25°C). Water was supplied *ad libitum* to the rats. Ethical approval was sought and received from the Ethical Committee of the Faculty of Pharmacy of Olabisi Onabanjo University, Nigeria on the usage of animals for experimental studies.

Preparation of Plant Extracts

The leaves of *H. indicum* were air dried for nine days under shade and powdered using a laboratory blender. The powdered leaf (250g) was soaked in 1 (one) litre of methanol in a glass jar for the first three days, then soaked in another 1 (one) litre of methanol for the next two days and soaked lastly in 0.5 litres of methanol for another two days. The different methanolic extract fractions were collected, added together and concentrated using the Rotary evaporator at a temperature of 55°C. The methanolic extract was then finally dried on the hot plate at a reduced temperature. The methanolic extract was then dissolved in a methanol:water mixture ratio of 1:3. This was partitioned with n – Hexane to separate the non-polar fraction from the polar fraction of the extract. The partitioning gave the aqueous solution fraction and the n – hexane soluble fraction which were separately concentrated on a water bath to dryness and then kept under suitable conditions (10°C to 20°C) for further bioactivity studies. The yield was calculated for each of the fractions.

Phytochemical Evaluations: Test for Saponins

The powdered sample of the n-hexane fraction of *H. indicum* (1g) was boiled with 10mls distilled water for 10 minutes and filtered while hot, then allowed to cool. 2.5ml of the filtrate was diluted with 10ml of water and shaken properly, and then two drops of castor oil were added to the solution and shaken vigorously for 2 minutes. A stable emulsion was formed and there was frothing indicating that the result was positive (Evans and Trease, 2002).

Phytochemical Evaluations: Test for Tannins

The powdered sample (0.5g) of the n-hexane fraction of *H. indicum* was boiled with 10ml of distilled water for 5 minutes, filtered while hot and cooled. The filtrate was adjusted to 20ml with distilled water. One ml of the filtrate was further diluted with distilled water to 5ml, after which a

few drops of 0.1% ferric chloride solution were added. A bluish-black or greenish colour indicates a positive test (Evans and Trease, 2002; Rojas *et al.*, 2006).

Gastro-protective Activity of *H. indicum*

Feeding of the animals was terminated 24 hours before the commencement of experimental procedures. The animals were, however, allowed free access to water and were then randomly divided into six treatment groups of four rats each. Two (2) ml of normal saline was administered orally to rats of Control Group I, 80 mg/kg/bw of Indomethacin (Hovid, Nigeria) was administered orally to rats of Experimental Group II, n – hexane fraction of the methanolic extract of *H. indicum* at doses of 100, 200 and 400 mg/kg/bw were administered orally to rats of Experimental Groups III, IV and V respectively while 50 mg/kg/bw of Ranitidine (Glaxowellcome, Egypt) was administered orally to rats of Experimental Group VI as the standard control or positive group.

One hour after administrations of 100, 200 and 400 mg/kg/bw of *H. indicum* and 50 mg/kg/bw of Ranitidine; 80 mg/kg/bw of Indomethacin was administered orally to rats of Groups III - VI (Adesanwo *et al.*, 2003; Raji *et al.*, 2003). After four hours, the animals were sacrificed by cervical dislocation and the stomach of each rat was removed. The stomach was opened along the lesser curvature and washed with normal saline to remove any debris (Adesanwo *et al.*, 2003; Raji *et al.*, 2003). Assessment of gastric ulceration was done by means of “scoring technique” as earlier described (Raji *et al.*, 2001).

The ulcer scoring system is as shown below:

- 0.0 -Normal stomach
- 0.5 -Punctuate or pinpoint hemorrhagic ulcer
- 1.0 -Two or more hemorrhagic ulcers
- 2.0 -Ulcers greater than 3mm in diameter.

Macroscopic examination of the stomach was carried out with a hand lens of X2 magnification (Raji *et al.*, 2001).

Histopathological Examinations

The excised stomach of rats of Groups I - VI were kept in 10% formol-saline. Three to five μm thick sections of the stomach samples were cut on a Reichert-Jung 2050 rotary microtome (Cambridge Instruments, Germany). Sections were floated on water bath at 50°C and mounted on pre-washed, sterilized 25.4 X 76.2 mm glass slides (Pearls, China). Sections were stained with haematoxylin and eosin as described by Kiernan (1990) and modified by Akinlolu *et al.* (2008).

Statistical Analyses

Computed data were expressed as Mean \pm S.E.M. Computed Means were compared using paired sample students't- test. Data were analyzed using Microsoft Excel 2007 (Microsoft Corporation, USA).

RESULTS

Gross Morphological Analyses

Indomethacin induced a morphological ulcer score of 2.0 in all rats of Group II that received only 80 mg/kg/bw of Indomethacin. Gross morphological analyses show that the n-hexane fraction of the methanolic extracts (at doses of 100, 200 and 400 mg/kg/bw) of the dried leaves of *H. indicum* protected against 80 mg/kg/bw of Indomethacin-induced gastric ulceration in treated rats of Groups III – V in a dose-dependent manner. Gross morphological analyses show that administrations of 100, 200 and 400 mg/kg/bw of *H. indicum* and Ranitidine resulted in percentage ulcer reduction of 45%, 50%, 75% and 95% respectively.

Histological Analyses

Histological evaluations were carried out on the stomach of each rat using routine laboratory haematoxylin and eosin method (Kiernan, 1990) to evaluate the histogastroprotective effects of *H. indicum* on Indomethacin – induced gastric ulcerated mucosa of wistar rats. None of the

rats used in this study died while the experimental procedures lasted. Thus, all rats were histologically examined to evaluate the degree of ulceration, degree of gastro-protection and other intrinsic histopathological alterations on the mucosal lining of the stomach. Photomicrograph of histo-pathological examinations of the stomach sections of rats of Groups I – VI are presented in Figures 1 - 6.

Histological analyses of the stomach body walls in rats of Groups II and III that received 80 mg/kg/bw of Indomethacin only and 100 mg/kg/bw of *H. indicum* show erosion of the mucus-secreting cells, gastric pit, upper and middle parts of gastric glands and the parietal cells. Histological observations of the stomach body walls in rats of Group IV that received 200 mg/kg/body weight of *H. indicum* show erosion of few mucus-secreting cells, gastric pit and the parietal cells. Histological observations of the stomach body walls in rats of Groups I, V and VI that received 80 mg/kg/body weight of Normal Saline, 400 mg/kg/body weight of *H. indicum* and 50 mg/kg/body weight of Ranitidine respectively show normal morphological appearances of the different components of the mucosa layer. Therefore, the n-hexane fraction of the methanolic extracts of the dried leaves of *H. indicum* had dose - dependent gross morphological and histo-gastroprotective effects against Indomethacin- induced gastric ulceration.

Table 1: Doses of *H. indicum*/Drug Received by Experimental Rats and Ulcer Parameters, degree of ulceration and percentage of ulcer reduction in rats

Groups of Rats	Dosage of <i>H. indicum</i> Drug (mg/kg/bw)	Mean Ulcer Score (\pm SEM)	Degree of Ulceration	Ulcer Reduction (%)
I	2mls Normal Saline	0.00 \pm 0.00	0	-
II	80mg Indomethacin	2.00 \pm 0.00	40	0
III	100mg/kg/bw <i>H. indicum</i>	2.00 \pm 0.00	30	45
IV	200mg/kg/bw <i>H. indicum</i>	1.10 \pm 0.24	22	50
V	400mg/kg/bw <i>H. indicum</i>	1.00 \pm 0.27	20	75
VI	50mg/kg/bw Ranitidine	0.50 \pm 0.22	10	95

S.E.M. = Standard Error of Mean of 4 determination,, bw = body weight, *H. Indicum* = *Heliotropium indicum*.

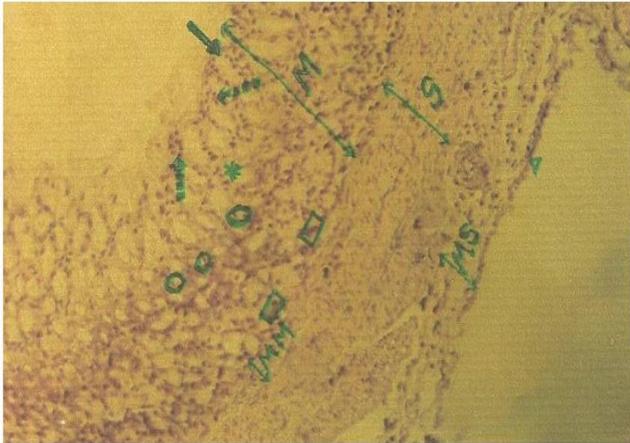


Figure 1: Photomicrograph sample of the stomach bodywall in rats of Group I which received only physiological saline. Haematoxylin and Eosin X 800. The Solid Arrow is on the gastric pit, the broken arrow is on the mucus secreting cells, the star sign is on the gastric gland, the circle sign is on parietal cells and the square sign is on peptic cells. M = Gastric Mucosa, MM = Muscularis Mucosae, S = Submuscularis and MS = Muscularis Layer and the triangle sign points to the Serosa. *Histological observations of the stomach body wall show normal morphological appearance of the different components of the mucosa layer.*

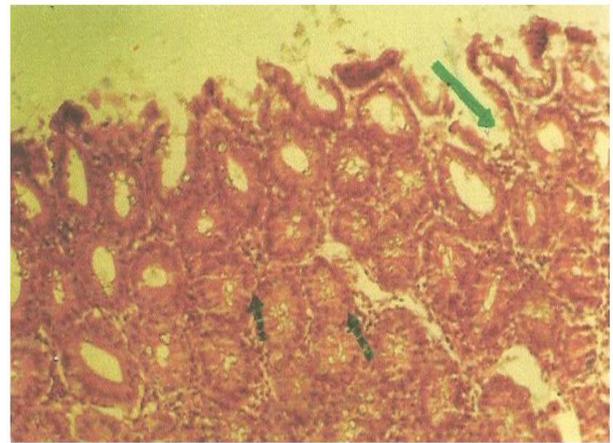


Figure 2: Photomicrograph sample of the stomach bodywall in rats of Group II which received oral administration of 80 mg/kg/bw of Indomethacin without prior oral administration of any dosage of extract of *Helicotropium Indicum*. Haematoxylin and Eosin X 800. The solid arrow is on the base of a gastric gland. The broken arrow on the peptic cells. *Histological analyses of the stomach bodywall show erosion of mucus-secreting cells, gastric pit, upper and middle parts of gastric glands and the parietal cells. The stomach mucosa is ulcerated.*

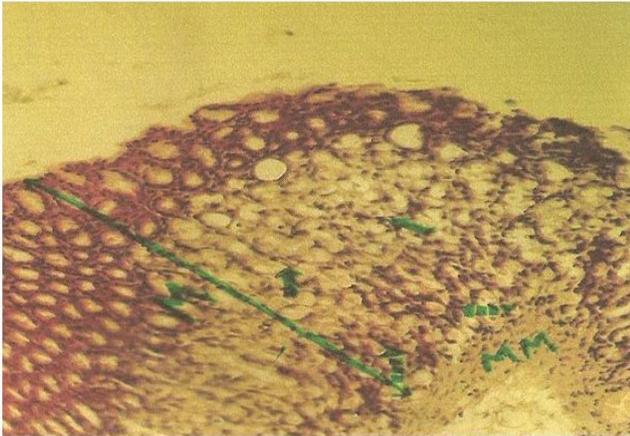


Figure 3: Photomicrograph sample of rats in Group III which received oral administration of 100 mg/kg/bw of extract of *Heliotropium indicum* prior to receiving further oral administration of 80 mg/kg/bw of Indomethacin. Haematoxylin and Eosin X 800. The solid arrow is on the parietal cells, the broken arrow is on the peptic cells. M = Gastric Mucosa, and MM = Muscularis Mucosae. *Histological analyses of the stomach body wall show erosion of the mucus-secreting cells, gastric pit and parietal cells. The stomach mucosa is ulcerated.*

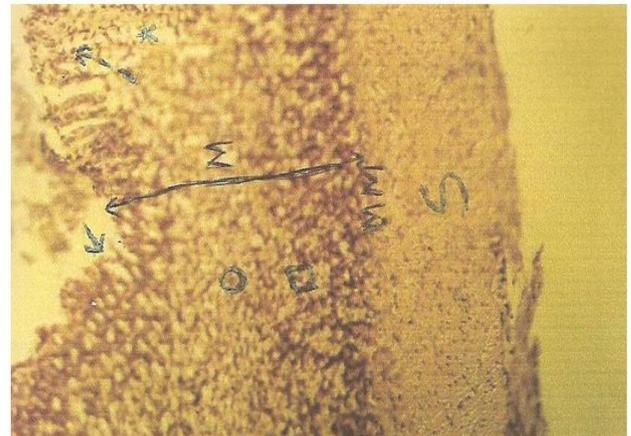


Figure 4: Photomicrograph sample of stomach bodywall of rats in Group IV which received oral administration of 200 mg/kg/bw of extract of *Heliotropium indicum* prior to receiving further oral administration of 80 mg/kg/bw of Indomethacin. Haematoxylin and Eosin X 800. The solid arrow is on the upper part of the gastric gland, the broken arrow is on the mucous secretion cells, the star sign is on the gastric gland, the circle sign is on parietal cells and the square sign is on peptic cells. M = Mucosa. MM = muscularis mucosa, and S = submuscularis *Histological observations of the stomach body wall show mild erosion of few mucus-secreting cells, gastric pit and parietal cells.*

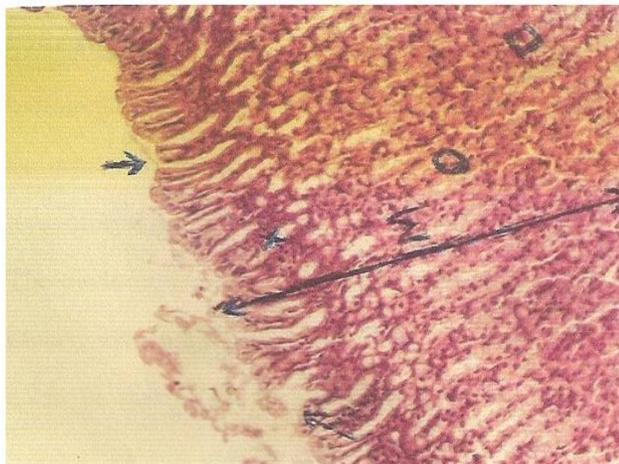


Figure 5: Photomicrograph sample of stomach bodywall of rats in Group V which received oral administration of 400 mg/kg/bw of extract of *Heliotropium indicum* prior to receiving further oral administration of 80 mg/kg/bw of Indomethacin. Haematoxylin and Eosin X 800. The solid arrow is on gastric pit, the broken arrow is on mucous secreting cells, the star sign is on the gastric gland, the circle sign is on parietal cells and the square signs is on peptic cell. M = Gastric Mucosa. *Histological observations of the stomach body wall show normal morphological appearance of the different components of the mucosa layer.*

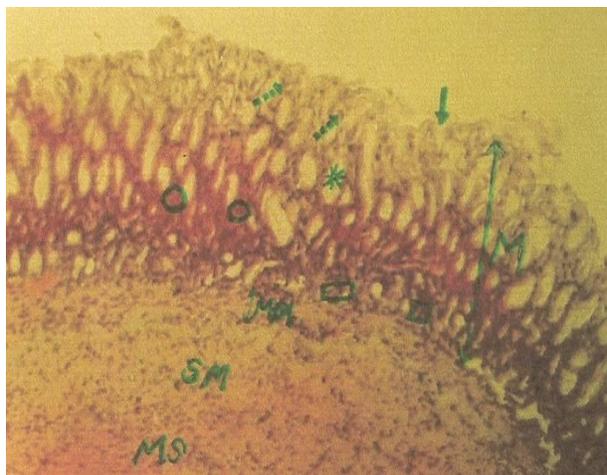


Figure 6: Photomicrograph sample of the stomach bodywall in rats of Group VI which received oral administration of 50 mg/kg/bw of Ranitidine prior to receiving further oral administration of 80 mg/kg/bw of Indomethacin. Haematoxylin and Eosin X 800. The solid arrow is on the gastric pit, the broken arrow is on mucous secreting cells, the star sign is on the gastric gland, the circle sign is on parietal cells and the square sign is on peptic cells. M = Gastric Mucosa, MM = Muscularis Mucosae, SM = Submuscularis and MS = Muscularis Layer. *Histological observations of the stomach body wall in rats show normal morphological appearance of the different components of the mucosa layer.*

DISCUSSION

Indomethacin is an established ulcerogen, especially in an empty stomach (Evbuonwan and Bolarinwa, 1991; Akinlolu *et al.*, 2008). The incidence of Indomethacin induced ulceration is mostly predominant in the glandular (mucosal) part of the stomach (Akinlolu *et al.*, 2008). Although, the mechanisms underlying the ulcerogenicity of indomethacin are not completely understood; it has been known that indomethacin induces gastric mucosa ulceration through inhibition of prostaglandins synthesis (John, 2000; Akinlolu *et al.*, 2008). This view is supported by the fact that several prostaglandins and prostaglandin analogues are potent anti-secretory and anti-ulcer agents. Prostaglandins normally serve protective functions in the stomach by maintaining gastric micro circulation via mucus and bicarbonate stimulation (John, 2000; Sabiha *et al.*, 2011).

Indomethacin stimulates catecholamines release from adrenal medulla, resulting in mucosal vasoconstriction (John, 2000; Sabiha *et al.*, 2011). It has also been established to uncouple mitochondrial respiration resulting in depletion of Adenosine Triphosphate and a reduced potential of gastric epithelial cells to coordinate normal cellular functions (John, 2000; Sabiha *et al.*, 2011). Hence,

Indomethacin reduces both the quality as well as amount of mucus secretion and changes in ionic permeability characteristics of gastric mucosa (John, 2000; Akinlolu *et al.*, 2008)

Indomethacin induced a morphological ulcer score of 2.0 in all rats of Group II that received only 80 mg/kg/bw of Indomethacin. Gross morphological analyses show that the n – hexane fraction of the methanolic extracts (at doses of 100, 200 and 400 mg/kg/bw) of leaves of *H. indicum* protected against indomethacin- induced gastric ulceration in treated rats of Groups III – V in a dose – dependent manner. Gross morphological analyses show that administrations of 100, 200 and 400 mg/kg/bw of *H. indicum* and Ranitidine resulted in percentage ulcer reduction of 45%, 50%, 75% and 95% respectively. Our findings are in agreement with earlier studies which noted percentage ulcer reductions in treatment with leaf extracts of *H. Indicum* (Akinlolu *et al.*, 2008; Ashoka *et al.*, 2011).

Histological analyses of the stomach body walls in rats of Groups II and III that received 80 mg/kg/bw of Indomethacin only and 100 mg/kg/bw of *H. indicum* show erosion of the mucus-secreting cells, gastric pit, upper and middle parts of

gastric glands and the parietal cells (Figures 2 and 3). Erosion of few mucus-secreting cells, gastric pit and the parietal cells were observed in rats of Group IV that received 200 mg/kg/bw of *H. indicum* (Figure 4). Normal morphological appearances of the different components of the mucosa layer were observed in rats of Groups I, V and VI that received 80 mg/kg/bw of Normal Saline, 400 mg/kg/bw of *H. indicum* and 50 mg/kg/bw of Ranitidine respectively (Figures 1, 5 and 6). Therefore, the n-hexane fraction of the methanolic extracts of the dried leaves of *H. indicum* had dose-dependent gross morphological and histo-gastroprotective effects against Indomethacin-induced gastric ulcerations. These observations are in agreement with our previous study which noted the gastroprotective activity of *H. Indicum* (Akinlolu *et al.*, 2008).

The precise mechanism of action of *H. indicum* is not clear, however, it has been proposed that mucosal protection induced by non-prostanoid compounds may be mediated through the mobilization of endogenous prostaglandins (Ima *et al.*, 2010; Sabiha *et al.*, 2011) Hence, it is possible that one of the mechanisms of anti-ulcerogenic effect of *H. indicum* might be due to its ability to mobilize endogenous prostaglandins in gastric mucosa.

The phytochemical analyses of the methanolic extract of the n-hexane fraction of *H. indicum* indicate the presence of tannins and saponins. Tannins being astringent with vasoconstricting effects (Ima *et al.*, 2010; Sabiha *et al.*, 2011; Ateeq *et al.*, 2013) might have precipitated microproteins on the sites of ulcers thereby forming a non-penetrable protective pellicle over the lining to prevent absorption of toxic substances and resist attack of proteolytic enzymes (Ima *et al.*, 2010; Sabiha *et al.*, 2011; Ateeq *et al.*, 2013). Tannins as phenolic compounds are known to enhance the status of oxidative stress biomarkers with the ability to scavenge free radicals, which could also have enhanced its anti-ulcer activity (Ima *et al.*, 2010; Sabiha *et al.*, 2011; Ateeq *et al.*, 2013). Saponins possess immunomodulatory, anti-inflammatory and vasoprotective effects which could have aided the observed anti-ulcer activity of *H. indicum* (Ima *et al.*, 2010; Sabiha *et al.*, 2011; Ateeq *et al.*, 2013).

The gastro - protective potential of n – hexane fraction of the methanolic extracts of the dried leaves of *H. indicum* against indomethacin-induced ulceration in rats, might in part, be due to its tannins and saponin constituents. Due to inadequate facilities in our laboratory, the active components responsible for the anti-ulcer activities of *H. indicum* could not be characterized and isolated. Further

experiments should be conducted to isolate the active components responsible for the anti-ulcer activities of *H. indicum*.

CONCLUSIONS

The findings of this study show that the n-hexane fraction of *H. indicum* possessed gastroprotective properties, probably due to the presence of tannins and saponins in its non-polar components.

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