

The Chemical Constituents Extractable From Teak Tree (*Tectona Grandis Linn*) Obtained From Fountain University, Osogbo

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ABSTRACT

A branch of Teak tree was cut, dried and grounded with mortar and pestle before using blender. The powder was extracted with four different solvents namely *n*-hexane, benzene, chloroform and distilled water; the extracts were concentrated using rotary evaporator. Gas chromatography- mass spectroscopy (GC-MS) was used to identify various constituents in the samples. Bis(2-ethylhexyl)phthalate was the major constituent common to Chloroform (35.50 %) and benzene (26.78 %) while *n*-hexadecanoic acid was the major constituent extracted by *n*-hexane (21.76 %) and water (24.16 %). Other phthalic acid derivatives extracted by both chloroform and benzene are Phthalic acid, di(2-propylpentyl)ester and di(oct-3-yl)ester. All the extracts were screened for termites repellence; all of them except water extract repelled termites in various degrees.

Keywords: Bis (2-ethylhexyl)phthalate, *n*-hexadecanoic acid, rotary evaporator, termites.

INTRODUCTION

Tectona grandis Linn otherwise known as “teak” belongs to verbanaceae family. The plant *Tectona agrandis* is probably the most widely cultivated high value hardwood in the world and it originates from India, Myanmar and South- East Asian countries. Researches have confirmed that teak contains many quinones (Lukmandaru, 2012). It is also proved that the quinone and its derivatives exhibit antitermitic activities.

It is now one of the most important species of tropical plantation forestry. The whole plant is highly medicinal and many reports claimed it cures several diseases. The research has shown that the plant is used in the treatment of bronchitis, cold and headache, urinary discharge, used as a laxative and sedative, as diuretic, anti-diabetic, analgesic and anti-inflammatory (Neha and Sangeeta, 2013).

Teak is an obvious choice for different activities because it is used on a large scale in many countries especially its timber which has a high wood value. Also, teak is easily established in plantations, which allows introduction of improved genetic material.

It is a large deciduous tree, 10-20 m tall; densely clothed with yellowish grey tomentum. Leaves are opposite, ovate-elliptic to ovate, 30-50 x 15-20 cm, cuneate at base. Flowers are small, whitish and bisexual. They appear in large panicles containing up to a few thousand flower buds, which open only few at a time during flowering period of 2-4 weeks. Calyx in flower is 2.5-3 cm long, in fruit enlarged to 2-2.5 cm or more, bladderly, enclosing the fruit. Fruit is a drupe with 4 chambers; round, hard and woody, enclosed in an inflated, bladder-like covering; pale green at first, then brown at maturity. Each fruit contains 0 to 4 seeds. Seeds are oblong, brown, enclosed in bony endocarp (Neha and Sangeeta, 2013).

The high natural durability of teak is an important characteristic that has been attributed to various extractive compounds identified in teak. Quinones and their derivatives have been confirmed to have termiticidal effects (Lukmandaru, 2011). Deference in natural durability may be related to the concentration of toxic extractable substances of wood accumulation during the formation of heartwood.

Teakwood is also known for its natural resistance against non-native termites (Rudi *et al.*, 2013). This advantage can be attributed to extractives present in teakwood. There are numerous bioactive compounds isolated from the teakwood. A variety of compounds have been isolated from almost every part of teak with varied degree of structures, belonging to different classes such as flavonoids, steroidal compounds, glycosides, quinones and phenolic acids (Ohmura *et al.*, 2000).

Its wood is known internationally for its beauty, weightlessness, durability and weather resistance and it is used in the building of ships, furniture, house floors and walls, and carpentry works (Miranda *et al.*, 2011; Lukmandaru and Takahashi, 2008). Currently, the wood market has a great interest in teak extractives such as naphthoquinones and anthraquinones, which have shown remarkable antifungal and antitermitic effects (Guerrero-Vásquez, 2013; Healey and Gara, 2003). Additionally, teak populations serve significant environmental roles, as they can be used in agroforestry systems and forest recovery (Healey and Gara, 2003). These characteristics make teak one of the most widely grown and economically profitable trees around the world (Hallett *et al.*, 2011).

Termites are social insects found in a wide range of terrestrial environments and are distributed throughout the warmer regions of the world. They are very important organisms ecologically as they significantly contribute to the organic decomposition process either by direct consumption of decomposing plant materials, by physical and chemical conditioning the soil they inhabit and by nitrogen fixation. They feed on a very wide variety of organic detritus like dry grass, decaying leaves, animal dung, humus and living or dead wood.

The destructive effects of termites to man are enormous including damages done to timbers used in buildings and other purposes (Su and

Scheffrahn, 2000). Mounds formed by termites make cultivation of land difficult, they also interferes with traffic, if formed on the tracks or along the roads. The mounds can also form a source of termite infestation of field crops (Piper, 2007). Biological deterioration of wood is of concern to the timber industry due to the economic losses caused to wood in service or in storage. They attack different components of wood at different rates giving rise to a particular pattern of damage (Sirmah, *et al.*, 2009). Degradation is influenced by environment whether in storage or in use. The degraded wood material enhances soil fertility for plantation (Silva, *et al.*, 2007).

Since termites are destructive and a constant threat to properties, efforts are been made to control their activities. Various control methods including application of termiticides, graded stones, glass splinters, stainless steel and chemical barriers are used (Culliney and Grace, 2000).

The present research work used four different solvents to extract the teak tree obtained from Fountain University's premises; the extracts were analyzed with GC-MS to determine the constituents extractable by each solvent and evaluated the termiticidal effect of the extracts. Bis(2-ethylhexyl)phthalate that is known to be toxic according to literature is the major constituent extracted by benzene and chloroform (Seth, 1982).

MATERIALS AND METHODS

Collection and Preparation of Samples

The sample was collected and identified at the Department of Botany Herbarium, University of Ibadan, Ibadan, Nigeria. About 5.5 inches diameter of teak wood were cut in Fountain University, Osogbo and chopped into various sizes and was allowed to dry under the sun for 76 days. The chopped wood was pounded using a mortar and pestle and blended to powdery form using a master chef blender.

Extraction of Wood Sample

The blended wood sample (100 g) was weighed into white handkerchief and was kept in glass container with lid. 500 ml of distilled water, benzene, *n*-hexane and chloroform were poured on each sample. The samples were then shaken for 24 hours on an orbital shaker at 100 oscillations (cold extraction). The content was decanted after 24 hours and they were concentrated using rotary evaporator. The weights of the extracts were determined and recorded.

Infrared Spectrophotometric Analysis

About 1mg of each of the four samples (in solid form) was finely grounded in a small mortar with added pure potassium bromide. The mixture was pressed into a disc using a special mould and a hydraulic press. The functional group was determined using infrared spectrophotometer (FTIR-8201A single beam laser Shimadzu Infrared Spectrophotometer).

GC-MS Analysis

Extracts of the samples were subjected to GC-MS analysis (model: 7890A (GC) Agilent Technologies) interfaced with Mass Selective Detector (Model: 5975C (MSD)). The electron ionization was at a 70v with an ion source temperature at 250°C. Helium gas (99% purity) was used as a carrier gas in a HP-5 column (30mm X 0.25mm X 0.320µm). The oven temperature was set at 80°C held for 1 minute and ramped to 240°C at the rate of 10°C/minute while holding for 5 minutes. Sample injected was 1µL.

Repellency Test

Repellency tests were conducted following the method proposed by Talukder and Howse (1993). Filter-paper circles of 9 cm in diameter were cut in half. One mL (25 mgL⁻¹) of samples was applied uniformly on one half using a pipette. The treated half-circles were air-dried until the solvent was totally evaporated. The treated and the untreated half-circles were placed

contiguously on the petri dishes and 10 adult termites were released on each dish. Termites present in each half circle were counted at hourly intervals for 5 hours after treatment. Data were converted to express percentage repulsion (PR) using the following formula:

$$PR (\%) = (Nc - 50) \times 2$$

Where Nc is the percentage of termites present in the control half. Positive values (+) indicated repellency and negative values (-) attractancy. Three replications were made for each treatment and the average calculated.

RESULTS AND DISCUSSION

Table 1 presents IR absorption peaks of all the samples. The IR absorption frequencies at 3009, 2942, 3014, 3013 cm⁻¹ of *n*-hexane, benzene, chloroform and water extracts respectively correspond to the C-H of aromatic compound. It has been reported that the aromatic C-H stretches are to the left of 3000 cm⁻¹ and the alkyl C-H stretches are to the right of 3000 cm⁻¹ (Scott, *et al.*, 2013; Serdar, 2005). Absorptions at 1785, 1782, 1781 cm⁻¹ in the *n*-hexane, benzene, chloroform extracts were characteristic absorption frequencies of carbonyl group (C=O) but it is absent in distilled water extract. The two sets of peaks suggest that the major constituents of the samples are aromatic with carbonyl functional group perhaps derivatives of phthalic acid. The absorption peaks at 3500 cm⁻¹ which is common to all the spectra indicate the presence of hydroxyl (O-H) group as shown in column 3 of Table 1. Absorptions at 1785, 1782, 1781 cm⁻¹ in the *n*-hexane, benzene, chloroform extracts were characteristic absorption frequencies of carbonyl group (C=O) but it is absent in distilled water extract. The two sets of peaks suggest that the major constituents of the samples are aromatic with carbonyl functional group perhaps derivatives of phthalic acid. The absorption peaks at 3500 cm⁻¹ which is common to all the spectra indicate the presence of hydroxyl (O-H) group. The absorption peaks at 3500 cm⁻¹ which is common to all the spectra

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Hexane extracted phthalic acid and its derivatives- 1, 2-benzenedicarboxylic acid (19.43 %) as shown in Table 2, while distilled water neither extracted phthalic acid nor its derivatives. Both benzene and chloroform extracted phthalic acid and its derivatives at a very high degree (26.78 %) and (35.05 %) respectively as presented in Tables 3 and 4. All solvents were observed to have extracted *n*-Hexadecanoic acid

with *n*-hexane having the highest percentage (21.76 %), followed by distilled water (11.23 %), chloroform (10.35 %) and benzene (10.20 %). Other prominent chemical constituents extracted are Octadecanoic acid which was extracted by distilled water (12.65 %) and *n*-hexane (15.32 %) only. Constituents of other extracts as detected in GC-MS are presented in Tables 3, 4 and 5 with their corresponding percentages.

Table 1: FTIR absorbance regions of the extracts of *Tectona grandis*

Extracts	C-H stretches	OH stretches	C=O bond	C-H Stretches in the aromatic ring	In plane C-H bending
<i>n</i> -Hexane	3009	3500	1785	1594	1025
Benzene	2942	3500	1782	1595	904
Chloroform	3014	3500	1781	1596	1036
Water	3013	3500	-	1569	1063

Table 2: GC-MS analysis of Constituents of *n*-hexane Extract

Rt (min)	Constituents	Percentage (%)	MF	MW
20.67	Hexadecanoic acid, methyl ester	4.45	C ₁₇ H ₃₄ O ₂	270.45
21.09	<i>n</i> -Hexadecanoic acid	21.76	C ₁₆ H ₃₂ O ₂	256.42
22.44	9,12-Octadecadienoic acid, methyl ester	11.86	C ₁₉ H ₃₄ O ₂	294.47
22.49	9-Octadecenoic acid (Z)-, methyl ester.	15.32	C ₁₉ H ₃₆ O ₂	296.49
27.48	1,2-Benzenedicarboxylic acid, diisooctyl ester.	19.43	C ₂₄ H ₃₄ O ₄	390.56
	Phthalic acid, 2-ethylhexyl isohexyl ester.		C ₂₂ H ₃₄ O ₄	362.50
27.86	Dodecanoic acid, ethyl ester.	4.69	C ₁₄ H ₂₈ O ₂	228.37
	Tetradecanoic acid, ethyl ester.		C ₁₆ H ₃₂ O ₂	256.42
	Nonanoic acid, ethyl ester.		C ₁₁ H ₂₂ O ₂	186.29

MF = Molecular formula; MW = Molecular weight

Table 3: Chemicals contained in chloroform Extract as obtained from GC-MS

Rt (min)	Constituents	Percentage (%)	MF	MW
21.17	<i>n</i> -hexadecanoic acid,	10.355	C ₁₆ H ₃₂ O ₂	256.42
	Tridecanoic acid		C ₁₃ H ₂₆ O ₂	214.34
22.65	9,10-Anthracenedione,2-methyl	8.104	C ₁₅ H ₁₀ O ₂	222.24
27.51	Bis(2-ethylhexyl)phthalate,	35.502	C ₂₄ H ₃₆ O ₄	388.54
	Phthalicacid,di(2-propylpentyl)ester,		C ₂₄ H ₃₈ O ₄	390.56
	Phthalicacid,di(oct-3-yl)ester		C ₂₄ H ₃₈ O ₄	390.56

MF = Molecular formula; MW = Molecular weight

Table 4: Chemicals contained in benzene Extract as obtained from GC-MS

Rt (min)	Constituents	Percentage (%)	MF	MW
15.947	Dodecanoic acid	5.08	C ₁₂ H ₂₄ O ₂	200.32
18.797	Tetradecanoic acid	2.27	C ₁₄ H ₂₈ O ₂	228.00
19.563	Cyclononasiloxane,octadecamethyl 1,1,1,5,7,7,7-Heptamethyl-3-3- bis(trimethylsiloxy)tetrasiloxane	2.01		
21.148	n-Hexadecanoic acid, Tetradecanoic acid	10.20	C ₁₃ H ₂₆ O ₂ C ₁₄ H ₂₈ O ₂	256.42 228.37
24.170	Cyclononasiloxane,octadecamethyl 1,1,1,5,7,7,7-Heptamethyl-3-3- bis(trimethylsiloxy)tetrasiloxane, Hexasiloxane,tetradecamethyl	3.071	C ₁₄ H ₄₂ O ₅ Si ₆	458.99
25.692	Cyclononasiloxane,octadecamethyl, Cyclodecasiloxane,eicosamethyl	3.339	C ₁₈ H ₅₄ O ₉ Si ₉ C ₂₀ H ₆₀ O ₁₀ Si ₁₀	667.39 741.54
27.506	Bis(2-ethylhexyl)phthalate, Phthalicacid, di(2-propylpentyl)ester, Phthalic acid, di(oct-3-yl)ester	26.786	C ₂₄ H ₃₆ O ₄ C ₂₄ H ₃₈ O ₄ C ₂₄ H ₃₈ O ₄	388.54 390.56 390.56
27.855	Undecanoic acid,2-methyl-,methyl ester Dodecanoic acid, ethyl ester Undecanoic acid, ethyl ester	1.428		

MF = Molecular formula; MW = Molecular weight

Table 5: Chemicals contained in distilled water Extract as obtained from GC-MS

Rt (min)	Constituents	Percentage (%)	MF	MW
20.925	n-hexadecanoic acid, Pentadecanoic acid	11.232	C ₁₆ H ₃₂ O ₂ C ₁₅ H ₃₀ O ₂	256.42 242.40
21.148	n-Hexadecanoic acid, Tridecanoic acid	24.168	C ₁₆ H ₃₂ O ₂ C ₁₃ H ₂₆ O ₂	256.42 214.34
23.128	Octadecanoic acid	12.654	C ₁₈ H ₃₆ O ₂	284.48

MF = Molecular formula; MW = Molecular weight

Repellancy Activity

Results of repellency of the various extracts of *Tectona grandis* are presented in Figure 1. Positive bars indicate repellency while negative bars indicate attractancy. Therefore, the Figure showed distilled water and *n*-hexane fractions poorly repelled the termites. In contrast, there

was significantly less number of termites in contact with the chloroform and benzene fraction discs when compared with the numbers of termites in control. Thus, chloroform and benzene extracts repelled the termites better than distilled water and *n*-hexane extracts.

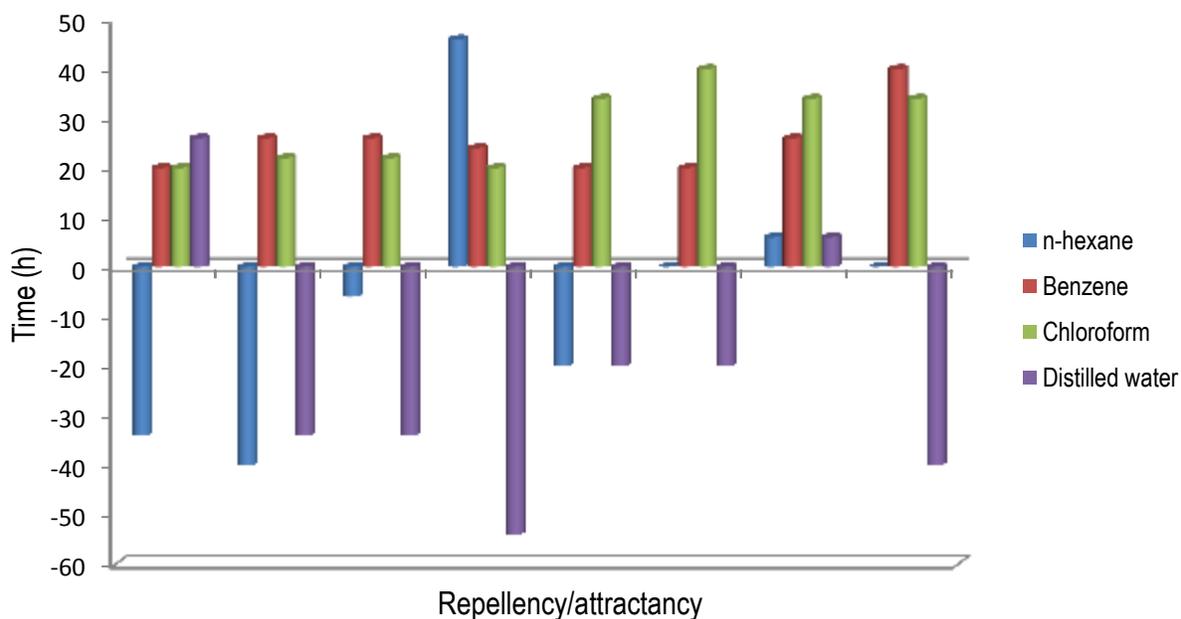
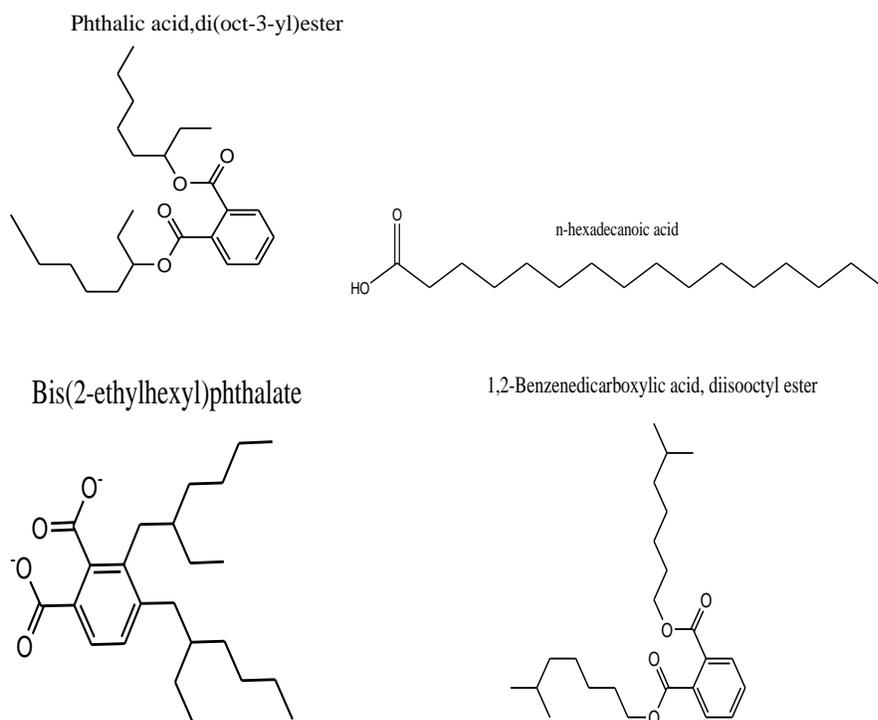


Figure 1: Repellency potential of *TectonagrandsL.f* extracts against termites



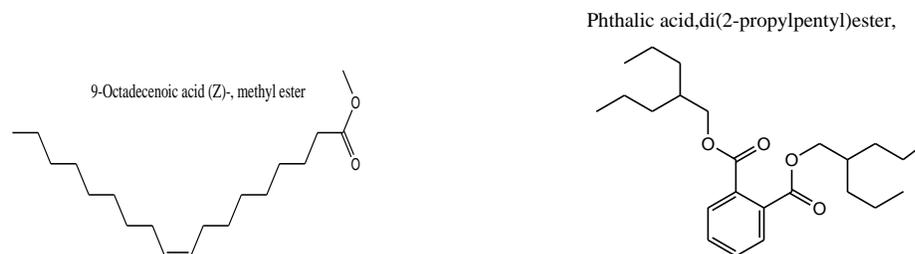


Figure 2: Chemical Structures of the major constituents of extractives *Tectona grandis*

CONCLUSION

It was discovered that Bis (2-ethylhexyl) phthalate that is toxic is extractable from *Tectona grandis* especially by benzene and chloroform (moderately polar solvents). Only extracts that contain high percentage of Bis (2-ethylhexyl) phthalate repelled the termite (benzene and Chloroform) while others appeared to attract the insects. The presence of Bis (2-ethylhexyl) phthalate in *Tectona grandis* may be responsible for its high resistance and durability properties.

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