

Effects of Vitamin C on Kidney and Bone of Rats Exposed to Low Doses of Cadmium

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ABSTRACT: In this study, the effect of vitamin C on cadmium-induced toxicity was investigated. Wister rats were exposed to cadmium (as CdSO₄.8H₂O), by sub-cutaneous injection, at doses of 1.0, 2.0 and 3.0 µg/kg body weight, with or without vitamin C supplementation, for four weeks. Serum alkaline phosphatase activity of the group of rats not supplemented vitamin C (group III) significantly (p<0.05) increased, all the groups supplemented with vitamin C also had significantly (p<0.05) increased serum alkaline phosphatase. The bone protein level and serum calcium of the vitamin C untreated group of rats, significantly (p<0.05) decreased relative to control. The bone calcium of the vitamin C treated rats significantly (p<0.05) decreased (group IIIc from 2896.30 ± 344.64 mg Ca/dl to 1049 ± 101.43 mg Ca/dl) while the bone phosphate of this same group of rats, significantly (p<0.05) increased. For some parameters evaluated, such as serum calcium and bone protein concentration, the effects of cadmium on the vitamin C treated rats were less pronounced, indicating that vitamin C may be protective against cadmium-induced toxicity.

Keywords: Cadmium, Vitamin C, Toxicity, Kidney, Bone

INTRODUCTION

For non-smoking, non-occupationally exposed population, food, especially of vegetable origin, is the main source of cadmium exposure (Kierstin, 2003). It is estimated that 98% of the ingested cadmium comes from terrestrial foods, while only 1% comes from aquatic foods such as fish and shellfish, and 1% arises from cadmium in drinking water (Van Assche, 1998). Acute ingestion of cadmium produces severe gastrointestinal irritation, which is manifested as severe nausea and vomiting, abdominal cramps and diarrhea. A lethal dose of cadmium for ingestion is estimated to be between 350 and 8900 milligrams (Fauci *et al.*, 1998). The chronic effects of cadmium are dose-dependent and also depend on the route by which the metal enters the body. Chronic inhalation causes emphysema and obstructive airways disease, and these occur before kidney damage is seen (Timbrell, 1995; Fauci *et al.*, 1998; Baldwin and Marshall, 1999). Long term ingestion causes kidney damage, which is first seen as proteinuria and β₂microglobulinuria (Fauci *et al.*, 1998; Williams *et al.*, 1999). In prolonged cadmium exposure, disorders of calcium metabolism occur, causing osteomalacia (Fauci *et al.*, 1998; Williams *et al.*, 1999). This leads to painful fractures, hence the name given to the chronic exposure disease in Japan: Itai-itai disease (literally "ouch!-ouch!" disease) (Fauci *et al.*, 1998; Williams *et al.*, 1999).

Cadmium is also known to be carcinogenic, and studies have linked it with cancers of the lungs and prostate (Timbrell, 1995; Fauci *et al.*, 1998; Williams *et al.*, 1999).

Studies have shown that vitamin C supplementation has varied effects on induced toxicity (Netke *et al.*, 1997). Ascorbic acid has been found to interact with several elements in such a way as to render them less available for animals (Hill, 1980). Grosicki (2004) reported a decrease in the carcass cadmium burden and the cadmium contents in the liver, kidney, testicles and muscles of cadmium exposed rats (1.0-1.2mgCd/kgb.w) given water supplemented with vitamin C (1.5mg/L) for 28days.

Apart from accidental and occupational exposure to cadmium, the general population in southern Nigeria is facing an increasing risk of cadmium exposure. A comprehensive and continuous monitoring of Warri River (Nigeria) between 1986 and 1991, showed that the average level of cadmium was 0.3mg/litre (Egborge, 1994); this far exceeds the minimum allowable level of cadmium in drinking water (0.005mg/litre) (WHO, 1984). This study was designed to evaluate the protective effects of vitamin C in rats exposed to low doses of Cd, i.e. doses that are frequently encountered

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in the natural environment, as a means of ameliorating its toxic effects.

MATERIALS AND METHODS

Chemicals and Reagents

Hydrated Cadmium sulphate ($\text{CdSO}_4 \cdot 8\text{H}_2\text{O}$) and other analytical grade chemicals used for this study were produced by E. Merck Darmstadt, Germany and BDH Chemical Limited, Poole, England. Pharmaceutical grade vitamin C tablets were obtained from Emzor Pharmaceuticals, Lagos, Nigeria. Alkaline phosphatase and calcium kits were products of Quimica Clinica Applicada SA (QCA), Spain, purchased from Equator Medics, Benin City, Nigeria.

Experimental Animals and Management

Forty-eight post weaned healthy albino rats (*Rattus norvegicus*) of average weight 84.20g were obtained from the Animal Unit of Lagos University Teaching Hospital (LUTH), Nigeria. Treatment of the animals was in accordance with the Principles of Laboratory Animal Care (NIH Publication 85-93, revised 1985). They were divided into eight (8) groups of six (6) rats each. All the rats were maintained on commercial feeds, product of Bendel Feeds and Flour Mill, BFFM, Ewu, Nigeria. A set of four groups (I, II, III, and IV) corresponding to control, 1.0, 2.0 and 3.0 $\mu\text{gCd/kg}$ body weight were given water, while another set of four groups (Ic, Iic, IIic and IVc) corresponding to control, 1.0, 2.0 and 3.0 $\mu\text{gCd/kg}$ body weight were given 5% vitamin C solution in place of water.

Once a day, five days a week, the rats were given 0.25ml of the appropriate cadmium solution or distilled water per 100g body weight by subcutaneous injection. Each rat was weighed weekly. At the end of the fourth week, the rats were anaesthetized in a chloroform saturated chamber and while under the influence of the anaesthesia, the kidneys and femur bones were collected. The tissues were homogenized in ice cold physiological saline (1:4 w/v of saline), blood was collected by heart puncture, allowed to clot on ice and then centrifuged at 5,000rpm for 5minutes. The supernatant of the centrifuged homogenates and the sera was kept frozen until analyses.

One of the femur bones from each rat was ashed at 600 °C for 12 h in a Gallenkamp muffle furnace (model FR614, Gallenkamp & Co., England). The ashed bones (dissolved in 2M HCl) were used for the determination of bone calcium and inorganic phosphate.

Biochemical Analysis

Serum, kidney and bone alkaline phosphatase (ALP) activities were assessed using QCA alkaline phosphatase kits. Briefly, the method measures spectrophotometrically, the intensity of the pink colour of phenolphthalein which is obtained by the hydrolysis of a colourless substrate phenolphthalein monophosphate by ALP (Babson *et al.*, 1966). Serum, kidney and bone protein levels were assayed by Biuret method (Gornall *et al.*, 1949) method. Serum and bone calcium concentrations were measured spectrophotometrically at 565nm, using QCA calcium kits. At alkaline pH, calcium forms a coloured complex with O-cresolphthalein, 8-hydroxyquinolein is added to the reagent as a chelating agent of magnesium ions which can interfere with the reaction (Baginski *et al.*, 1973). Serum and bone inorganic phosphate were measured spectrophotometrically at 625nm, according to the method described by Plummer (1978). In this method, ammonium molybdate- H_2SO_4 reagent reacts with inorganic phosphate to form complexes which is reduced to give a blue colour that can be read spectrophotometrically by adding 0.2% ascorbic acid.

Statistical Analyses

Values are expressed as means of 5 or 6 determinations \pm SEM. The results obtained for the vitamin C treated and untreated groups were analyzed separately (within the column) by the Independent Samples T-test, each group was compared to its respective control on SPSS 11.0, SPSS Inc., Chicago, Illinois, USA.. A $p < 0.05$ was considered statistically significant.

RESULTS AND DISCUSSION

Table 1 shows a general steady increase in body weight gain in the vitamin C untreated and treated groups of rats, however, while the final body weight of all tests groups of the vitamin C untreated rats decreased compared to control, only Group Iic of the vitamin C treated groups showed a significant ($p < 0.05$) decrease in final body weight compared to control. Group IVc actually recorded a significantly ($p < 0.05$) higher final body weight compared to control. Asagba *et al.* (2006) reported that rats given 3 mg Cd/ kg body weight (as CdCl_2) for 4 weeks, had lower final body weight ($p < 0.05$) compared to control. Ribas *et al.* (2004) also reported that the pups of female rats supplied with drinking water containing 300 mg/l of CdCl_2 during lactation, showed significantly lower ($p < 0.01$) body weight than control pups. Grosicki and Kowalski (2002), reported that rats exposed for 4 weeks to CdCl_2 (labeled with cadmium-109) at 10 mg/ kg diet, showed a steady increase in average body weight gain

throughout the experiment, however the final body weight of the rats exposed to Cd was markedly lower in comparison to the control rats. This observation correlates with the results obtained for the vitamin C untreated rats in this study.

Table 2 shows the general decreases observed in the bone- and kidney-body weight ratios for both set of rats, however, group IIc bone-body weight ratio was similar to its corresponding control. The kidney-body weight ratio of this same group was significantly ($p < 0.05$) higher than its control (Ic).

Since chemical toxicity can be assessed by changes in the body weight and organ-body weight ratio (Timbrell, 1991), the significant alterations of these parameters in the Cd-treated rats, especially the vitamin C untreated

rats implies Cd toxicity. The fact that only group IIc of the vitamin C treated rats showed a significant ($p < 0.05$) decrease in final body weight, while all the vitamin C untreated rats had significantly ($p < 0.05$) decreased final body weights compared to their corresponding control, implies that vitamin C had a protective effect against the Cd-induced body weight reduction.

Since the kidney and bone are implicated in non-acute cadmium exposure, the serum, kidney and bone ALP were determined in this study. The bile canaliculi of the liver, osteoblasts in the bone, proximal tubules in the kidney and mucosal cells of the small intestine, are rich sources of alkaline phosphatase (Verley, 1967). Damage to any of these organs or tissues would lead to elevated activity of the ALP isoform in the serum (Lin and Wang, 1986; Ngaha *et al.*, 1989).

Table 1: Body Weight of rats administered doses of cadmium

Group	Body Weight (g)				
	Week 0 (Initial)	Week 1	Week 2	Week 3	Week 4 (Final)
I	78.60 ^a ± 5.036	80.10 ^a ± 5.675	90.70 ^a ± 5.682	94.80 ^a ± 5.472	108.10 ^a ± 6.346
II	85.67 ^b ± 5.137	81.50 ^a ± 5.130	90.83 ^a ± 5.320	101.67 ^b ± 5.664	102.25 ^b ± 5.470
III	69.42 ^b ± 3.348	66.25 ^b ± 3.007	82.50 ^b ± 3.601	96.67 ^a ± 3.712	103.67 ^b ± 3.803
IV	75.00 ^a ± 4.645	83.75 ^b ± 4.727	87.25 ^a ± 4.765	93.75 ^a ± 5.028	102.17 ^b ± 5.143
Ic	86.12 ^{a*} ± 6.360	94.7 ^{a*} ± 6.878	99.39 ^{a*} ± 7.328	110.00 ^{a*} ± 7.292	111.88 ^{a*} ± 7.920
IIc	81.21 ^{a*} ± 6.084	72.4 ^{b*} ± 5.597	85.30 ^{b*} ± 5.98	89.20 ^{b*} ± 5.245	88.10 ^{b*} ± 4.488
IIIc	104.50 ^{b*} ± 8.24	95.10 ^{a*} ± 8.141	102.60 ^{a*} ± 8.95	115.30 ^{b*} ± 10.14	124.80 ^{b*} ± 10.99
IVc	93.10 ^{b*} ± 5.382	96.10 ^{a*} ± 4.831	99.30 ^{a*} ± 5.408	102.80 ^{a*} ± 5.126	108.80 ^{a*} ± 4.596

Values are mean of five or six determinations ± standard error of the mean. Values carrying b and b* are significantly ($p < 0.05$) different from control (a and a*).

Table 2: Organ- Body Weight Ratio of rats administered doses of cadmium.

Group	Organ-Body Weight Ratio (X 10 ⁻³)	
	Bone	Kidney
I	7.5 ^a ± 0.42	8.8 ^a ± 0.28
II	7.2 ^b ± 0.23	8.1 ^b ± 0.30
III	6.6 ^b ± 0.05	8.4 ^b ± 0.27
IV	7.9 ^b ± 0.30	8.1 ^b ± 0.25
Ic	8.8 ^{a*} ± 0.78	8.1 ^{a*} ± 0.62
IIc	8.7 ^{a*} ± 0.32	8.7 ^{b*} ± 0.10
IIIc	7.7 ^{b*} ± 0.30	7.3 ^{b*} ± 0.26
IVc	7.0 ^{b*} ± 0.14	7.3 ^{b*} ± 0.28

Values are mean of five or six determinations ± standard error of the mean. Values carrying b and b* are significantly ($p < 0.05$) different from control (a and a*).

I = Rats fed with 0 µgCd/kg Body weight
 II = Rats fed with 1.0 µgCd/kg Body weight
 III = Rats fed with 2.0 µgCd/kg Body weight
 IV = Rats fed with 3.0 µgCd/kg Body weight

Ic = Rats fed with 0 µgCd/kg Body weight + 5% vitamin C solution
 IIc = Rats fed with 1.0 µgCd/kg Body weight + 5% vitamin C solution
 IIIc = Rats fed with 2.0 µgCd/kg Body weight + 5% vitamin C solution
 IVc = Rats fed with 3.0 µgCd/kg Body weight + 5% vitamin C solution

Table 3 shows significant ($p < 0.05$) decreases observed in tissue ALP of almost all the groups of the vitamin C untreated and treated rats. Axelsson and Piscator (1966), reported a reduction in the ALP activity of renal

cortex in rabbits given 0.5mgCd/kg body weight. Saillenfait *et al.* (2006) also reported that pups from pregnant rats intraperitoneally injected with CdCl₂ at 2.5 mg/kg body weight for up to 14 days of gestation,

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exhibited significant decreases in renal ALP activity. The significantly high serum ALP (Table 3) observed in this study could be as a result of the leakage of this enzyme from damaged tissues, rich in ALP, into the blood stream. This is particularly evident in the vitamin C treated groups where both bone and kidney ALP were significantly ($p < 0.05$) decreased while the serum enzyme significantly ($p < 0.05$) increased compared to control (Ic). Itokawa *et al.* (1978) observed an increase in the activity of serum ALP coincident with changes in the skeleton after 120 days of oral exposure to 50 ppm cadmium. Akesson *et al.* (2006), using a population based women health survey in Southern Sweden to investigate the association between low-level cadmium

exposure and osteoporosis, reported that urinary cadmium displayed a near-significant ($p < 0.06$) association with serum bone-alkaline phosphatase (bALP). Since the kidney and bone, are established target organs of cadmium toxicity (Ahn and Park, 1995; Kido *et al.*, 1995); in addition to the decreased tissue ALP observed in this study, it is plausible to conclude that the increase in serum ALP is a reflection of the decreased bone and kidney ALP.

Serum total protein level is a rough measure of protein status but reflects major functional changes in kidney and liver functions (Pachathundikandi and Verghese, 2006).

Table 3: Serum, kidney and bone alkaline phosphatase of rats administered doses of cadmium.

Group	Alkaline Phosphatase U/L		
	Bone	Kidney	Serum
I	215.16 ^a ± 2.56	466.32 ^a ± 14.69	99.51 ^a ± 2.22
II	193.78 ^b ± 15.14	411.28 ^b ± 12.26	94.62 ^a ± 2.63
III	192.51 ^b ± 13.48	467.19 ^a ± 8.51	146.09 ^b ± 6.11
IV	182.63 ^b ± 12.77	435.01 ^b ± 14.71	118.73 ^a ± 2.40
Ic	286.60 ^a ± 15.83	494.15 ^a ± 15.30	74.26 ^a ± 10.15
IIc	195.81 ^b ± 10.21	436.84 ^b ± 16.49	123.22 ^b ± 3.10
IIIc	203.80 ^b ± 12.55	448.67 ^b ± 5.10	123.22 ^b ± 3.44
IVc	240.57 ^b ± 16.77	478.27 ^b ± 12.07	124.17 ^b ± 2.88

Values are mean of five or six determinations ± standard error of the mean. Values carrying b and b* are significantly ($p < 0.05$) different from control (a and a*).

I = Rats fed with 0 µgCd/kg Body weight

II = Rats fed with 1.0 µgCd/kg Body weight

III = Rats fed with 2.0 µgCd/kg Body weight

IV = Rats fed with 3.0 µgCd/kg Body weight

Ic = Rats fed with 0 µgCd/kg Body weight + 5% vitamin C solution

IIc = Rats fed with 1.0 µgCd/kg Body weight + 5% vitamin C solution

IIIc = Rats fed with 2.0 µgCd/kg Body weight + 5% vitamin C solution

IVc = Rats fed with 3.0 µgCd/kg Body weight + 5% vitamin C solution

Table 4 shows the serum, kidney and bone protein levels. Low doses of cadmium administration for 4 weeks appears to have a general lowering effect on the bone, kidney and serum protein levels of the vitamin C treated groups. Also, the significant reduction in the bone protein level of group IV of the Vitamin C untreated rats, show that cadmium interferes with bone protein levels. The reductions observed in the bone, kidney, and serum protein levels of all test groups of the vitamin C treated rats are puzzling, however, since statistical analysis show that vitamin C alone significantly ($p < 0.05$) increased the bone, kidney and serum protein levels of group Ic (Table 6), the reductions observed can be attributed to cadmium-induced toxicity. Proteinuria due to kidney impairment in cadmium toxicity may be the cause of protein loss in the test groups since inhibitory role of cadmium in protein synthesis has not been established.

The reduction in serum calcium in the vitamin C untreated groups is a reflection of the effect of cadmium on calcium metabolism (Table 5). The decreased Ca²⁺ absorption and negative calcium balance in cadmium exposed rats could result from the inhibitory effect of cadmium on the activation of vitamin D in renal cortical cells (Feldman and Cousins, 1973). Nogawa *et al.* (1987) reported that serum 1,25-dihydroxycholecalciferol levels were lower in Itai-Itai disease patients and cadmium exposed subjects with renal damage. A reduction in serum calcium level (hypocalcaemia) stimulates the release of calcium from bone, this correlates well with the reduced bone calcium observed in the test groups of the vitamin C untreated and vitamin C treated rats. Itokawa *et al.* (1978) reported that simultaneous administration of cadmium with a low-protein, low-calcium diet led to a decrease in calcium and zinc content of the bone.

Cadmium can affect calcium, phosphorus and bone metabolism (Mukunoki and Fujimoto, 1996). Reductions in serum 1, 25-dihydroxycholecalciferol in cadmium-exposed subjects, are closely related to serum concentrations of parathyroid hormone (PTH), β_2 -microglobulin and the percentage tubular reabsorption of phosphate (Nogawa *et al.*, 1987), suggesting that cadmium-induced bone effects may also be due to disturbances in vitamin D and PTH metabolism (Kjellstrom, 1992). Phosphorus is important for the modulation of calcium mobilization from the bone and the regulation of plasma calcium (Felsenfeld and Rodriguez, 1999). Increase in serum calcium is

associated with decrease in serum phosphorus and increased urinary phosphorus excretion and *vice versa* (Felsenfeld and Rodriguez, 1999).

The results obtained for serum calcium and phosphate in this study (Tables 5) do not sufficiently reflect the reciprocal relationship between calcium and phosphorus, however, group IVc, of the vitamin C treated rats, shows an increased calcium level while its phosphate level is decreased. The results for bone calcium and phosphate levels of the vitamin C treated rats, correlate better with the reciprocal relationship between calcium and phosphorus.

Table 4: Serum, kidney and bone protein levels of rats administered doses of cadmium.

Group	Protein Levels mg/ml		
	Bone	Kidney	Serum
I	5.38 ^a ±0.41	12.13 ^a ±0.74	47.80 ^a ±1.04
II	6.00 ^b ±0.33	14.50 ^b ±0.67	52.50 ^b ±0.48
III	6.00 ^b ±0.42	12.50 ^a ±0.44	48.90 ^a ±1.26
IV	2.70 ^b ±0.27	11.50 ^b ±0.54	47.75 ^a ±0.95
Ic	6.67 ^{a*} ±0.52	15.00 ^{a*} ±0.90	52.75 ^{a*} ±2.31
IIc	4.80 ^{b*} ±0.51	8.88 ^{b*} ±0.71	42.80 ^{b*} ±1.15
IIIc	5.63 ^{b*} ±0.28	9.10 ^{b*} ±0.49	49.80 ^{b*} ±0.55
IVc	6.88 ^{a*} ±0.43	11.25 ^{b*} ±1.32	46.75 ^{b*} ±1.48

Values are mean of five or six determinations ± standard error of the mean. Values carrying b and b* are significantly (p<0.05) different from control (a and a*).

Table 5: Calcium and Phosphate Levels in Serum and Bone of Rats Administered Doses of Cadmium

Group	Calcium mg Ca/dl		Phosphate Levels µg/ml	
	Bone	Serum	Bone	Serum
I	1641.98 ^a ±119.19	12.23 ^a ±0.54	1397.50 ^a ±37.68	253.75 ^a ±7.37
II	1323.46 ^b ±94.25	11.16 ^b ±0.18	1310.00 ^b ±19.68	196.67 ^b ±0.96
III	1033.33 ^b ±129.00	8.24 ^b ±0.49	1362.50 ^a ±33.38	188.75 ^b ±9.96
IV	1739.46 ^b ±28.21	8.48 ^b ±0.10	1479.17 ^b ±20.98	240.00 ^a ±5.93
Ic	2896.30 ^{a*} ±344.64	10.10 ^{a*} ±0.14	1200.00 ^{a*} ±15.10	318.33 ^{a*} ±26.94
IIc	1017.28 ^{b*} ±40.65	8.46 ^{b*} ±0.20	1295.00 ^{b*} ±30.57	70.00 ^{b*} ±2.83
IIIc	1049.99 ^{b*} ±101.43	8.99 ^{b*} ±0.38	1400.00 ^{b*} ±30.77	176.25 ^{b*} ±12.15
IVc	1370.37 ^{b*} ±108.56	12.2 ^{b*} 0 ±0.20	1347.50 ^{b*} ±19.24	211.67 ^{b*} ±14.84

Values are mean of five or six determinations ± standard error of the mean. Values carrying b and b* are significantly (p<0.05) different from control (a and a*).

I = Rats fed with 0 µgCd/kg Body weight
 II = Rats fed with 1.0 µgCd/kg Body weight
 III = Rats fed with 2.0 µgCd/kg Body weight
 IV = Rats fed with 3.0 µgCd/kg Body weight

Ic = Rats fed with 0 µgCd/kg Body weight + 5% vitamin C solution
 IIc = Rats fed with 1.0 µgCd/kg Body weight + 5% vitamin C solution
 IIIc = Rats fed with 2.0 µgCd/kg Body weight + 5% vitamin C solution
 IVc = Rats fed with 3.0 µgCd/kg Body weight + 5% vitamin C solution.

Analysis of groups I and Ic, show that vitamin C alone positively influenced most of the parameters evaluated (Table 6). For some of the parameters evaluated, recognizable differences were observed between the vitamin C untreated and the vitamin C treated groups of

rats, These include: The final body weights of groups IIIc and IVc of the vitamin C treated rats did not decrease relative to control, while all the groups of the vitamin C untreated rats decreased compared to their control. The bone protein concentration of group IV of

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the vitamin C untreated groups of rats show a significant ($p < 0.05$) decrease compared to its control, while there was no significant difference in group IVc of the vitamin C treated groups, compared to its control. A significant ($p < 0.05$) decrease in serum calcium was observed in all the vitamin C untreated groups, group IVc of the vitamin C treated groups, show a significant increase compared to its control.

Cadmium may also exert its toxicity via oxidative damage. Valko *et al.* (2005) reported that the primary route for cadmium toxicity is depletion of glutathione and binding to sulfhydryl groups of protein. It is therefore not surprising that vitamin C, an antioxidant, would play a protective role against cadmium toxicity.

Table 6: Effect of Vitamin C on all Parameters Evaluated

Parameter	Group I	Group Ic
Final Body Weight (g)	108.10 ± 6.346 ^a	111.88 ± 7.920 ^a
Bone-Body Weight Ratio	7.5 ± 0.42 ^a	8.8 ± 0.78 ^b
Kidney-Body Weight Ratio	8.8 ± 0.28 ^a	8.1 ± 0.62 ^b
Bone ALP U/L	215.16 ± 2.56 ^a	286.60 ± 15.83 ^b
Kidney ALP U/L	466.32 ± 14.69 ^a	494.15 ± 15.30 ^b
Serum ALP U/L	99.51 ± 2.22 ^a	74.26 ± 10.15 ^b
Bone Protein Level mg/ml	5.38 ± 0.41 ^a	6.67 ± 0.52 ^b
Kidney Protein Level mg/ml	12.13 ± 0.74 ^a	15.00 ± 0.90 ^b
Serum Protein Level mg/ml	47.80 ± 1.04 ^a	52.75 ± 2.31 ^b
Bone Calcium Level mgCa/dl	1641.98 ± 119.19 ^a	2896.30 ± 344.64 ^b
Serum Calcium Level mgCa/dl	12.23 ± 0.54 ^a	10.10 ± 0.14 ^a
Bone Phosphate level µg/ml	1397.50 ± 37.68 ^a	1200.00 ± 15.10 ^b
Serum Phosphate level µg/ml	253.75 ± 7.37 ^a	318.33 ± 26.94 ^b

Values are mean of five or six determinations ± standard error of the mean. Values carrying b and b* are significantly ($p < 0.05$) different from control (a and a*).

I = Rats fed with 0 µgCd/kg Body weight

Ic = Rats fed with 0 µgCd/kg Body weight + 5% vitamin C solution

CONCLUSION

The findings from this study have shown that subcutaneous administration of low doses of cadmium to rats either with or without vitamin C supplementation caused an increase in serum ALP and bone phosphate. It also caused a decrease in bone protein concentration, serum calcium and bone calcium, indicating that under the conditions of this study, the bone is the major target of cadmium toxicity. Vitamin C may play a protective role against cadmium toxicity and hence vitamin C can be used to ameliorate the toxic effect of low doses of cadmium.

REFERENCES

Ahn, D.W. and Park, Y.S. (1995). Transport of inorganic phosphate in renal cortical brush-border membrane vesicles of cadmium intoxicated rats. *Toxicology and Applied Pharmacology*, **133**(2): 239 - 243.

Akesson, A., Bjellerup, P., Lundh, T., Lidfeldt, J., Nerbrand, C., Samsioe, G., Skeefving, S. and Vahter, M. (2006). Cadmium-induced effects on bone in a population-based study of women.

Environmental Health Perspective, **114**(6): 830-834.

Asagba, S.O., Adiakpoh, M.A., Kadiri, H. and Obi, F.O. (2006). Influence of aqueous extract of *Hibiscus sabdariffa* L. petals on cadmium toxicity in rats. *Biology of Trace Element Resource*, 115: 47-57.

Axelsson, B. and Piscator, M. (1966). Renal damage after prolonged exposure to cadmium: An experimental study. *Archive of Environmental Health*, **12**: 360-373.

Babson, A.L., Greeley, S.J., Coleman, C.M. and Phillips, G.E. (1966). Phenolphthalein monophosphate as a substrate for serum alkaline phosphatase. *Clinical Chemistry*, **12**: 482-490.

Baginski, E.S., Marie, S.S., Clark, W.L. and Zak, B. (1973). Direct microdetermination of serum calcium. *Clinical Chimica Acta*, **46**: 49-54.

Baldwin, D. R. and Marshall, W. J. (1999). Heavy metal poisoning and its laboratory investigation (Review Article). *Annals of Clinical Biochemistry*, **36**: 267-300.

- Egborge, A.B.M. (1994). *Water pollution in Nigeria. Biodiversity and Chemistry of Warri River*. Ben Miller Books Nig. Nigeria. Pp 32.
- Fauci, A.S., Braunwald, K., Isselbacher, K.J., Wilson, J.D., Martin, J.B., Kasper, D.L., Hauser, S.L. and Longo, D.L. (eds). (1998). *Heavy metal poisoning. in: Harrison's Principles of Internal Medicine*. 14th ed. McGraw-Hill, New York. Pp2564-2569.
- Feldman, S.L. and Cousins, R.G. (1973). Influence of cadmium on the metabolism of 25-hydroxycholecalciferol in chicks. *Nutrition Reports International*, **8**: 251-259.
- Felsenfeld, A.J. and Rodriguez, M. (1999). Phosphorus, regulation of plasma calcium, and secondary hyperthyroidism: A hypothesis to integrate a historical and modern perspective. Review Article. *Journal of American Society of Nephrology*, **10**: 878-890.
- Gornall, A.G., Bardawill, J.C. and David, M.M. (1949). Determination of serum proteins by means of Biuret reaction. *The Journal of Biological Chemistry*, **177**: 751-760.
- Grosicki, A. and Kowalski, B. (2002). Whole body and organ retention of cadmium after repeated administration to rats. *The Bulletin of the Veterinary Institute in Pulawy*, **46**: 143-147.
- Grosicki, A. (2004). Influence of vitamin C on cadmium absorption and distribution in rats. *Journal of Trace Element and Medical Biology*, **18(2)**: 183-187.
- Hill, C.H. (1980). Interactions of vitamin C with lead and mercury. *Annals of NY Academic of Science*, **355**: 262-266.
- Itokawa, Y., Nishino, K., Takashima, M., Nakata, T., Katto, H., Okamoto, E., Daijo, K. and Kawamura, J. (1978). Renal and skeletal lesions in experimental cadmium poisoning of rats. Histology and renal function. *Environmental Research*, **15**: 206-217.
- Kido, T., Kobayashi, E., Hayano, M., Nogawa, K., Tsuritani, I., Nishijo, M., Tabata, M., Nakagawa, H., Nuyts, G.D. and Debroe, M.E. (1995). Significance of elevated urinary human intestinal alkaline phosphatase in Japanese people exposed to environmental cadmium. *Toxicology Letter*, **80(1-3)**: 49 - 54.
- Kierstin, P. G. (2003). Lactational transfer of cadmium in rodents - CNS: Effects in the offspring. Acta Universitatis Agriculturae Sueciae. Veterinaria Vol. 150. Doctoral Thesis. Pp 10-11.
- Kjellstrom, T. (1992). Mechanism and epidemiology of bone effects of cadmium. IARC Sci. Publ. 118; 301-310.
- Lin, J. K. and Wang, C. J. (1986). Protection of crocin dyes in the acute hepatic damage induced by aflatoxin b1 and dimethylnitrosamine in rats. *Carcinogenesis*, **7(4)**: 595 - 599.
- Mukunoki, J. and Fujimoto, K. (1996). Collection and recycling of used Ni-Cd batteries in Japan, sources of cadmium in the environment, Inter-Organisation Programme for the Sound Management of Chemicals (IOMC), Organisation for Economic Co-operation and Development (OECD), Paris.
- Netke, S. P., Roomi, M. W., Tsao, C. and Niedzwiecki, A. (1997). Ascorbic acid protects guinea pigs from acute aflatoxin toxicity. *Toxicology and Applied Pharmacology*, **143**: 429-435.
- Ngaha, E. O., Akanji, M. A. and Madusuolumo, M. A. (1989). Studies on correlation between chloroquine - induced tissue damage and serum enzyme changes in the rat. *Experientia*, **45**: 143 - 146.
- Nogawa, K., Tsuritani, I., Kido, T., Honda, R., Tamada, Y. and Ishizaki, M. C. (1987). Mechanism for bone disease found in inhabitants environmentally exposed to cadmium: Decreased serum 1 & 1, 25-dihydroxy vitamin D level. *International Archived Occupational and Environmental Health*, **59**: 21-30.
- Pachathundikandi, S.K. and Verghese, E.T. (2006). Blood Zinc Protoporphyrin, Serum total protein, and total cholesterol levels in automobile workshop workers in relation to lead toxicity: Our experience. *Indian Journal of Clinical Biochemistry*, **21(2)**: 114-117.
- Plummer, D.T. (1978). *An Introduction to Practical Biochemistry*. McGraw Hill Ltd. Mardenhead Berkshire, England. Pp. 109-185.
- Ribas, J.P., Lopes, R.A., Sala, M.A., Ribas, L.M.R., de Mattos, M.C., Semprini, M., Watanabe, I. and Regalo, S.C.H. (2004). Effect of cadmium on rat maxillary molar junctional epithelium during lactation. *International Journal of Morphology*, **22(4)**: 257-262.
- Saillenfait, A.M., Payan, J.P., Ban, M. and de Ceaurriz, J. (2006). Indirect and lactation-associated changes in renal alkaline phosphatase of newborn rats prenatally exposed to cadmium chloride. *Journal of Applied Toxicology*, **12(3)**: 205-210.

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- Timbrell, J. A. (1995). *Cadmium. In: Introduction to Toxicology*. 2nd edition. Taylor and Francis, London. Pp 76-77.
- Timbrell, J.A. (1991). *Principles of Biochemical Toxicology*. 2nd Edition Taylor & Francis, London, Washington DC. Pp. 369-370.
- Valko, M., Morris, H. and Cronin, M. T. (2005). Metals, toxicity and oxidative Stress. *Current in Medicinal Chemistry*, **12(10)**: 1161-1208.
- Van Assche, F.J. (1998). A stepwise model to quantify the relative contribution of different environmental sources to human cadmium exposure, Paper presented at *NiCad '98*, Prague, Czech Republic, 1998; September 21-22.
- Verley, H. (1967). *Practical Clinical Biochemistry*, 4th edition, Heinemann Medical Books Ltd and New York Interscience Books, Inc, New York. Pp 891 – 921
- WHO (1984). Guidelines for Drinking-water Quality: Health Criteria and Other Supporting information. World Health Organization, Geneva. Vol. 2 Pp. 84-90.
- Williams, F., Robertson, R. and Roworth, M. (1999). *Scottish Centre for Infection and Environmental Health. Detailed Profile of 25 Major Organic and Inorganic Substances*. 1st ed. Glasgow: SCEIH.