ANTIULCER AND GASTRIC ANTISECRETORY EFFECTS OF LANDOLPHIA OWARIENSIS EXTRACTS IN RATS

SAMUEL. B. OLALEYE, VICTOR B. OWOFULE and A. O. ODUKANMI

Gastrointestinal Research Unit, Department of Physiology, College of Medicine, University of Ibadan, Ibadan, Nigeria

Department of Physiology, College of Health Sciences, University of Ilorin, Ilorin, Nigeria

E-mail: olaleye_ui@hotmail.com, sb.olaleye@mail.ui.edu.ng

Summary: Water, Methanol and Chloroform Extracts of Landolphia owariensis were investigated for their effects on gastric acid secretion and ulceration in male albino rats. Two models of gastric lesion induced in experimental Wistar rats- HCl/ethanol- induced gastric lesions and Pylorus ligation-induced gastric lesions – were employed. In both models, the antiulcer activity of LA was compared with that of cimetidine (100 mg kg p.o.). In the HCl/Ethanol model, ulcer index and mucus production was determined. In pylorus ligated rat, ulcer index, mucus production, total volume of gastric juice and gastric acidity level were measured. Pre-treatment of animals with the aqueous extracts (100mg/kg compared to control group, to levels comparable to that produced by cimetidine. The results indicate that the leaf extracts of LO contains antiulcer principles.

Keywords: Landolphia owariensis, antiulcer, gastric secretion, rat

Introduction

Peptic ulcer is a common disease throughout the world and in the past one or two decades there has been a phenomenal increase in the contribution to knowledge on the treatment of the disease (Odaibo, 1988). The protective effects of some medicinal plant species have been investigated for anti-ulcer properties. Such plants include Egletes viscosa (Rao et al, 1997), Garcinia kola seeds (Ibironke et al, 1997), Picrorhiza kurroa (Anaden et al, 1999).

Landolphia owariensis P. beavu. (family Apocynaceae) commonly called vine rubber and known locally by various names in Nigeria-Nwalikali or Mba (Ibo); Panukuoro or Ibo akitipa (Yoruba) and Ciwo or Kuranga (Hausa) - is widely used locally for the treatment of many ailments. The decoction of leaves is used as a purgative and to cure malaria. In the southwestern Nigeria, the leaf decoction is employed in the treatment of stomach pain and ulcer. The root is soaked in local gin for about a week and the extract given two full wine glass a day to cure gonorrhea (Gil, 1992). Lewis and Lewis (1977) also reported the use of stem bark as vermifuge. The latex is drunk or used in French Equatorial Africa as an enema for intestinal worms and in parts of Ivory Coast the latex forms an ingredient arrow poison (Irvine 1961). It is used to make native bear and beverage in Senegal and upper Nile land respectively (Dalziel, 1937).

Very few reports are available in literature to establish the anecdotal uses of this plant. Ebi and Ofoefule (1997) have validated the folkloric use of Landolphia Owariensis as an antimicrobial agent while the anti-inflammatory and analgesic properties have been reported (Owoyele et al, 2001). In this study the effect of various extracts of the leaves of Landolphia owariensis on experimental ulceration was investigated.

Materials and Methods

Plant Material:

L. Owariensis leaves were collected from their natural habitat at Gambari forest reserve, Oyo State Nigeria and authenticated by a Taxonomist of the forestry research institute of Nigeria (FRIN) Ibadan. Identification of the plant took place at FRIN by the same Taxonomist. A voucher specimen (FHI 105678) has been deposited in the Herbarium of the same institution.

Extract Preparation:

Air dried and powdered leaves of L. Owariensis were extracted successfully with water, Methanol and Chloroform at 80°C, 40°C and room temperature respectively. The dried extract was stored at 40°C until they were ready for use. The extract yields of the plant were 1.2g, 3.0g and 2.0g from 20.0g, 30.0g and 20.0g of powdered leaves in 150ml water, 300ml methanol and 250ml of chloroform respectively. The chloroform extract (CELO) was dissolved in 0.9% saline while the methanol extract (MELO) and chloroform extract (CELO) were each dissolved in 2.5% Tween 80 and subsequently in normal saline.

Animals

Adult male albino rats weighing between 120-150g obtained from the animal house of the College of Medicine, University of Ibadan, Nigeria were used.
They were housed in cages at room temperature with free access to rat cubes (Ladokun Feeds Nig. Limited, Ibadan, Nigeria).

**Pepsin binding activity:**

The method described by Rifat-uz-Zaman *et al* (2002) was essentially followed. Briefly, 500mg powdered AELO, MELO and CELO were added to 2mL of pepsin solution (2 mg/mL) each in the separate test tubes. 4mL of 0.2 N HCl buffered with 1 mL of 0.2 N sodium citrate solution was then added. Thereafter, one milliter of bovine serum albumin (5 mg/mL) was added to treat the excess pepsin except control test tubes, pre-incubation at 37°C for 30min. The contents were further incubated at 37°C for 30 min following shaking. Remaining protein in each tube was treated with 1.0 mL of Biuret reagent and alkalized by adding 5 mL 0.2 N NaOH solution. Their absorbances were read at 546 nm. The values obtained were expressed as percentage binding of pepsin.

**HCl/Ethanol-Induced Ulcers**

After a 2-week administration of 100mg/kg and 200mg/kg of the extracts of *Landolphia owariensis* (AELO, MELO and CELO) and vehicle (0.1ml, 25% Tween 80), all the groups had their food withdrawn 24hours but allowed free access to water. Afterwards, 1ml of HCl/ethanol mixture containing 0.15N HCl in 70% V/V ethanol was administered orally. The rats were sacrificed 1 hr later by ether overdose and the stomachs removed and observed for lesions in the glandular portion. Ulcers were independently assessed and scored by two observers using the method of Rao *et al.* (1997) as follows: 0 = no ulcers (normal stomach); 1 = up to 5 petechial hemorrhages, 2 = up to 5 petechial hemorrhages with erosions of depth 1mm, 3 = up to 10 petechial hemorrhages with erosions of depth 1mm; 4 = up to 10 petechial hemorrhages with erosion of depth above 1mm.

**Pylorus-ligated gastric secretion and ulceration**

Oral administration of 100mg/kg and 200mg/kg of the extract of *Landolphia owariensis* (AELO, MELO and CELO) and vehicle (0.1ml, 25% Tween 80) was done for two weeks. After treatments, all the animals were subjected to surgery under light ether anaesthesia according to Brodie (1966) The 4h gastric juice collection was drained into a graduated test tube and centrifuged at 2000rpm for 10min. The supernatant volume and pH were recorded. The total acid content of the gastric juice was also determined by titration to pH 7.0 with 0.05N NaOH, using phenolphthalein as indicator. Ulcers formed in the glandular portion of the stomachs were scored as previously described.

**Measurement of mucus production**

Gastric mucus production was measured in the rats that were subjected to HCl/ethanol induced lesions. After estimating the degree of lesion formation, the gastric mucosa of each rat was gently scraped using a glass slide. The gastric barrier mucous was estimated by the method of Corne *et al.* (1974). Briefly, the excised stomachs from the rats were soaked for 2 hours in 0.1% Alcian blue dissolved in buffer solution containing 0.1M sucrose and 0.05M Sodium acetate (pH adjusted to 5.8 with hydrochloric acid). After washing the stomach twice in 0.25M sucrose (15 and 45 min), the dye complexed with mucous was eluted by immersion in 10ml aliquots of 0.5M MgCl₂ for 2 hours. The resulting blue solution was shaken with equal volumes of diethyl ether and the optical density of the aqueous phase measured at 605nm using a spectrophotometer.

**Statistical Analysis**

All values presented in tables are expressed as means ± Standard error of Means (SEM). Comparisons between groups were made using student’s t-test. The difference between the groups is taken to be significant at P< 0.05.

**Results**

**Effect of Extracts of Landolphia owariensis on protein binding**

The in-vitro binding activities of the extracts are shown in Fig. 1. The Aqueous, methanol and chloroform extracts of *Landolphia owariensis* afforded 24.22%, 30.53% and 36.42% pepsin binding capacities respectively. No noticeable binding was observed in the standard tube.

**HCl/Ethanol induced lesion**

Treatment of animals with the HCl/ethanol mixture (0.15N HCl in 70% V/V ethanol) induced visible gastric ulceration characterized by elongated bands parallel to the greater axis of the stomach. As shown in Table 1, animals (pre-treated with normal saline) given HCl/Ethanol mixture had mean ulcer score of 23.70 ± 3.15 and an ulcer index of the same value. Pre-treatment of animals with the aqueous extracts (100mg/kg and 200mg/kg) significantly reduced formation of ulcers induced by HCl/ethanol mixture, the percentage inhibition being 43.8% and
55.27% respectively. The Chloroform extract afforded the least protection with 23.07% and 14.77% inhibition. In this model of ulcer, the standard drug, Cimetidine (100mg/kg) afforded a 54.5% protection on ulcer. There was also a dose dependent increase in mucus secretion in the extract treated animals.

Pylorus-ligated gastric secretion and ulceration

Pylorus ligation for 4 h resulted in the accumulation of gastric secretion and increase in the titratable acidity. Extracts produced dose-dependent significant decrease in the volume of gastric juice, titratable acidity and ulcer index as compared to control (Table 2). However, mucous production was significantly increased in the extract treated animals.

Discussion

Peptic ulcer is the most common chronic illness of the digestive system which causes symptoms of upper abdominal pain, fullness, gas and bloating. In some patients, it may also give rise to serious complications such as bleeding, bowel perforation, and obstruction. Ulcer bleeding, a common reason for emergency hospital admission, is the basis for most laboratory evaluation of antulcer activity of local plants. Many plants are known to have a long history of use for soothing inflamed and injured mucous membranes in the digestive tract. For example, Licorice may protect the stomach and duodenum by increasing production of mucin, (Goso et al, 1996). Moreover, some active components of plants have been shown to confer antulcer properties on certain plants. Consequently, reports of laboratory research have shown that compounds like flavonoids may inhibit growth of H. pylori, in addition to their direct cytoprotective effects (Beil et al, 1995).

The results of the present study have shown that the extracts of the investigated plant exhibit potent and dose related anti-ulcerogenic activities. Moreover, the aqueous extract produced a more effective cytoprotection to the mucosa. These activities may be linked to the reported presence of flavonoidal compounds (Owoyele et al, 2001) in L. Owariensis. Many plants with flavonoidal activities have been shown to be promising in the development of potent antulcer drugs (Olaleye and Farombi, 2006).

The high in-vitro pepsin binding activities of the extracts may suggest a possible and considerable in-vivo binding to substrate proteins by a non-specific hydrophobic interaction to form complexes which are less vulnerable to peptic hydrolysis. However, this is just an hypothesis to which further studies may throw more light on. There was a strong correlation between the protection afforded against experimental ulcers by the extracts and the mucous secretion. It is known that the layer of mucus which lines the gastrointestinal tract serves not only as a lubricant but equally more important, as a protective barrier between the mucosal surface and the luminal contents. In conditions where the integrity of the wall of the gastrointestinal tract is threatened, gastric mucous production may be increased. In addition, the biochemical characteristics of the component mucin molecules may change (Jass and Walsh, 2001). The extracts of L.O. may therefore be affording their gastroprotective effects via an increase in the defensive mechanisms of the stomach. Further studies are however needed to elucidate the unknown facts about the gastroprotective effects this plant. In conclusion, the overall finding of this study is that L. owariensis possesses antulcer properties and may be a promising material for treatment of ulcerogenesis and gastric mucosal injury.

### Table 1: Effect of Extracts of Landolphia owariensis on gastric lesions induced by HCl/Ethanol in rats

<table>
<thead>
<tr>
<th>Treatment and dose (mg kg body weight, orally)</th>
<th>Mucus production (mg)</th>
<th>Ulcer score Mean ± SE</th>
<th>Ulcer index Mean % inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (1 ml)</td>
<td>20.50 ± 1.55</td>
<td>23.7 ± 4.91</td>
<td>23.70 -</td>
</tr>
<tr>
<td>Vehicle (Tween 80)</td>
<td>20.8 ± 1.24&lt;sup&gt;NS&lt;/sup&gt;</td>
<td>22.20 ± 1.69</td>
<td>22.20 6.33</td>
</tr>
<tr>
<td>AELO (100mg/kg)</td>
<td>27.1 ± 1.35&lt;sup&gt;**&lt;/sup&gt;</td>
<td>13.30 ± 2.58&lt;sup&gt;**&lt;/sup&gt;</td>
<td>13.30 43.80</td>
</tr>
<tr>
<td>AELO (200mg/kg)</td>
<td>30.6 ± 1.14&lt;sup&gt;***&lt;/sup&gt;</td>
<td>10.60 ± 1.02&lt;sup&gt;**&lt;/sup&gt;</td>
<td>10.60 55.27</td>
</tr>
<tr>
<td>MELO (100mg/kg)</td>
<td>25.3 ± 1.79&lt;sup&gt;*&lt;/sup&gt;</td>
<td>13.60 ± 2.56&lt;sup&gt;**&lt;/sup&gt;</td>
<td>13.60 42.62</td>
</tr>
<tr>
<td>MELO (200mg/kg)</td>
<td>27.1 ± 2.02&lt;sup&gt;**&lt;/sup&gt;</td>
<td>11.60 ± 2.25&lt;sup&gt;**&lt;/sup&gt;</td>
<td>11.60 50.89</td>
</tr>
<tr>
<td>CELO (100mg/kg)</td>
<td>25.2 ± 2.00&lt;sup&gt;***&lt;/sup&gt;</td>
<td>18.10 ± 3.23&lt;sup&gt;NS&lt;/sup&gt;</td>
<td>18.10 23.63</td>
</tr>
<tr>
<td>CELO (200mg/kg)</td>
<td>25.9 ± 1.77&lt;sup&gt;*&lt;/sup&gt;</td>
<td>16.00 ± 1.90&lt;sup&gt;**&lt;/sup&gt;</td>
<td>16.10 32.49</td>
</tr>
<tr>
<td>Cimetidine (100mg/kg)</td>
<td>33.2 ± 1.20&lt;sup&gt;***&lt;/sup&gt;</td>
<td>7.63 ± 0.98&lt;sup&gt;**&lt;/sup&gt;</td>
<td>7.63 67.81</td>
</tr>
</tbody>
</table>

<sup>*</sup><p><0.05 <sup>p</sup> < 0.01 and <sup>***</sup> < 0.001 Student’s t-test, All treated groups were statistically compared to control group, Six rats were used in each group</p>
### Table 2: Ulcer index and Gastric juice parameters in rats with pylorus ligated stomach

<table>
<thead>
<tr>
<th>Treatment and dose (mg kg body weight, orally)</th>
<th>Ulcer index Mean ± SE</th>
<th>Gastric juice Vol (ml/4hr)</th>
<th>pH Acid output (x10^4 mmol/4hr)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>10.30 ± 2.91</td>
<td>4.6 ± 0.1</td>
<td>2.30 ± 0.22</td>
</tr>
<tr>
<td>Vehicle (Tween 80)</td>
<td>10.60 ± 1.69</td>
<td>4.3 ± 0.1</td>
<td>2.55 ± 0.13</td>
</tr>
<tr>
<td>AELO (100mg/kg)</td>
<td>7.30 ± 2.58**</td>
<td>2.8 ± 0.2**</td>
<td>3.45 ± 0.32</td>
</tr>
<tr>
<td>AELO (200mg/kg)</td>
<td>5.60 ± 1.02**</td>
<td>2.2 ± 0.2***</td>
<td>3.80 ± 0.12</td>
</tr>
<tr>
<td>MELO (100mg/kg)</td>
<td>7.60 ± 2.56**</td>
<td>3.3 ± 0.2*</td>
<td>3.20 ± 0.11</td>
</tr>
<tr>
<td>MELO (200mg/kg)</td>
<td>6.20 ± 2.25**</td>
<td>3.1 ± 0.1*</td>
<td>3.00 ± 0.15</td>
</tr>
<tr>
<td>CELO (100mg/kg)</td>
<td>9.40 ± 3.23NS</td>
<td>3.5 ± 0.3NS</td>
<td>2.85 ± 0.28</td>
</tr>
<tr>
<td>CELO (200mg/kg)</td>
<td>7.60 ± 1.90*</td>
<td>3.2 ± 0.3*</td>
<td>2.75 ± 0.37</td>
</tr>
<tr>
<td>Cimetidine (100mg/kg)</td>
<td>4.50 ± 0.98**</td>
<td>2.1 ± 0.2***</td>
<td>3.85 ± 0.22</td>
</tr>
</tbody>
</table>

*<005 p<0.01 and ***<0.001 Student’s t-test, All treated groups were statistically compared to control group, Six rats were used in each group.

References