EFFECT OF SUB CHRONIC ADMINISTRATION OF ETHANOLIC LEAF EXTRACT OF CROTON ZAMBESICUS ON HEMATOLOGICAL PARAMETERS OF RATS

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Summary: Ethanolic leaf extract of *Croton zambesicus* was administered to rats at doses of 100 – 400mg / kg for 21 days to investigate its effect on the haematological indices of rats. Haematological indices, namely packed cell volume (PCV), Haemoglobin concentration (Hb) Red blood cell count (RBC), Mean cell Haemoglobin concentration (MCHC), Mean cell volume (MCV), and Mean Corpuscular Haemoglobin (MCH) were assessed from whole blood obtained from the treated animals as well as those in the control group. The extract at the doses administered was found to caused reductions in PCV, HB, RBC, MCH, MCH and WBC, in a dose – dependent fashion. However elevation of MCV was observed. This results indicate that the extract has the potential of suppressing haemopoiesis and causing anaemia.

Key Words: Croton zambesicus, haemotological indices, anaemia, rats.

Introduction

Croton zambesicus Muell Arg. (Euphorbiaceae) syn C. amabilis Muell. Arg.; syn. C. gratissimus Burch), is an ornamental tree grown in villages and towns of Nigeria. It is a Guineo - Congolese species widely spread in tropical Africa. The leaf decoction is used in Benin as anti hypertensive antimicrobial (urinary infections)(Adjanohoun et al, 2002). The Ibibios in Uruan Area of Akwa Ibom State use the leaf traditionally as malaria remedy. Block et al (2002) reported that ent - trachyloban -3β – ol, a trachylobane diterpene, isolated from dichloromethane extract of the leaves has cytotoxic activity on Hela cells. The alkaloidal fractions of the leaf have been reported to possess a weak activity against Klebsiella pneumoniae, Pseudomonas aeruginosa, Bacillis megaterium, Bacillus subtilis, Eschericia coli, Proteus mirabilis, Staphyloccocus aureus, Aspergillus niger, Microsporum species and Penicillium species (Abo et al 1999). The alkaloidal fractions of the stem have also been reported to be active against all micro- organisms mentioned above (Abo et al 1999). The essential oil found in the leaves contain P – cymene, linalool and beta – caryophyllene (Menut et al, 1995). The constituent of the essential oil also found in the flowering tops are pinene, limonene, linalool, methol, carvone, thymol, alpha humulene and ceisnerolidol (Mekkawi, 1985). The study was aimed at investigating the effect of repeated administration of ethanolic leaf extract of Croton zambesicus on haematological indices namely packed cell volume, haemoglobin concentration, red blood cell count, mean haemoglobin concentration, white

blood cell count and subsequently to evaluate whether its ethnopharmacological uses particularly as malaria remedy may have possible side effects such as anaemia, which is common with the use of most chemotherapeutic agents.

Materials and Methods

Preparation of Plants Extract

The leaves of *Croton Zambesicus* Muell. Arg. (Euphorbiaceae) were collected August, 2003, from a compound in Uyo, Akwa Ibom State, and authenticated by Dr Uduak Eshiet, a taxonomist in the department of Botany, University of Uyo, Uyo Nigeria. Hebarium specimen was deposited at faculty of pharmacy hebarium University of Uyo, Uyo with voucher no. FPUU209.

The fresh leaves (2kg) of the plant were dried on a laboratory table for 8 days and reduced to powder. The powder 100g was macerated in ethanol (300ml) for 72 hours. The liquid extract obtained was concentrated in vacuo at 40°C. The yield was 3.81%. The extract was stored in a refrigerator at 4°C until used for experiments reported in this study.

Animals

The animals used in the study were young adult, male and females, (150- 180g) albino wistar rats obtained from University of Uyo animal house, Uyo, Nigeria. The animals were used after an acclimatization period of 10days to room temperature and relative humidity of $28\pm5^{\circ}\text{C}$ and 50% respectively. The were housed in standard cages and

maintained on standard animal pellets (Pfizer livestock feeds, Aba Nigeria) and water ad libitum.

Experimental Design

The rats were weighed and randomly assigned on the basis of weight into 5 groups of 5 animal each. Animals in Groups A, B, C and D were orally administered with 400, 300, 200 and 100 mg/kg of the extract respectively, while animals in group E were orally given normal saline 5 ml/kg and served as control. Administration of the extract continued for 21 days between 0.8.00 and 0.9:00 hour each day. Twenty four hours after the last administration, the animals were anaesthetized with chloroform vapour and dissected. Whole blood was obtained by cardiac puncture from each rat and collected into anticoagalant – treated (EDTA 0. 77M) sterile bottles. This was used for haematological studies. haemoglobin (Hb) was determined spectrophotometrically by the cyanomethaemoglobin method (Jain, 1986), packed cell volume (PCV) was determined by method of Jain, (1986). Red Blood Cell (RBC) count was estimated by haemocytometer method of Arowolo (1982). Blood was diluted in 1,200 Dacie's fluid which keeps and preserves the integrity of the RBC. For White Blood Cell (WBC) counts the dilution factor was 1:20 using 2-3% solution of acetic acid to which gentian violet was added.

The calculations for red cell indices were made as described by Jain (1986) Mean Corpuscular Volume (MCV) = PCV x 10 / RBC (fl)

Mean Corpuscular Haemoglobin (MCH) = Hb x 10 / RBC (pg)

Mean corpuscular Haemoglibin concentration = Hbc x 100 / PCV

Statistical Analysis

All data obtained were statistically analysed using students' t-test. The data were expressed as average mean \pm standard error and values of P <0.05 were considered significant.

Results

The results of the effect of sub chronic administration of ethanolic leaf extract of Croton zambesicus on haematological indices of rats is presented in Table 1. Treatment of rats for 21days with the extract resulted in a dose- dependent reduction of PCV. The reduction was significant (P < 0.5) in group A with a value of 50.7 + 5.9% (mean +S.D) compared to control value of 56.7+4.8%. The values for group B, C and D were $52.5 \pm 7.6,53.2 \pm 4.9$ and $55.3 \pm 2.5\%$ respectively (Table 1). The blood haemoglobin concentrations (g/dl) were reduced in a dose – dependent fashion with significant (P< 0.05) decrease recorded in groups A and B relative to control. The values were 12.3 ± 0.8 , 13.2 ± 1.0 , 14.6 \pm 0.3, 15.5 \pm 0.8 and 16.0 \pm 0g/dl for groups A, B, C, D, and E respectively A non – significant (P< 0.5) reduction in RBC counts of rats treated with the various doses of the extract was observed. The counts were 8.84 ± 1.1 , 9.7 ± 0.5 , 9.7 ± 0.6 , 10.27 ± 2.0 millions per ml of blood respectively for groups A, B, C and D (Table 1), while the control group had a count of 10.6 ± 0.63 millions per ml of blood. There was a dose - dependent decrease in the value of MCHC calculated. A significant (P<0.05) decreased was recorded from rats in groups A with a value of $24.3 \pm 2.4\%$ compared to control group with 28. 22 \pm 1.0%. The values were 25.14 \pm 1.2, 27.4 \pm 0.9 and $28.02 \pm 3.1\%$ for groups B, C and D respectively. The calculated values of MCH showed a decrease compared to control in a dose - dependent fashion, although the differences were not significant. For groups A, B, C, and D, MCH values of 13. 91± 5.5, 13.6 ± 2.0 , 15.05 ± 4.6 and $15.06\pm3.0\%$ were recorded respectively, while that of control group was 15. 09 $\pm 1.0\%$. Elevation of MCV values was observed. A significant (P<0.05) elevation was recorded in group A with a value of $57.35 \pm 0.6 \mu \text{m}^3$ compared to control value of $53.49 \pm 0.4 \mu \text{m}^3$. MCV values were $54.12 \pm 0.4 \mu \text{m}^3$ 1.1, 54.84+0.7, and 53.74+1.0µm³ for groups B, C, and D respectively. The white blood cell counts (thousands / μ l) for groups A, B, C, and D were 7.7 \pm 3.3, 9.0±0.8, 13.8± 0.6 and 14.0± 4.9 respectively and the control had a counts of 14.7 + 6.0/ μ 1 of blood. The count was observed to reduce with increased doses.

Table 1: Effect of Ethanolic leaf extract of Croton zambesicus on haematological indices of rats.

Haamatologica	RBC	PCV	Hb	MCHC	МСН	MCV	TOTAL WBC
1 Parameters	$(x 10^6 \mu l)$	(%)	(g/dl)	(%)	(pg)0	(μm^3)	$(x 10^3 \mu l)$
Treatment							
GROUPS A (4000mg/ Kg)	8.84 ±1.1	50.7 <u>+</u> 5.9	12.3 ± 2.4	24.3 ± 0.8	13.9 ± 5.5	57.35 <u>+</u> 0.6	7.7 ± 3.3
GROUPS B (300mg/kg)	9.7 <u>+</u> 0.5	52.5 ± 7.6	13.2 <u>+</u> 7.6	25.14 ±1.2	13.6 ± 2.0	54.12 <u>+</u> 1.1	9.0 <u>+</u> 10.8
GROUPS C (200mg / kg)	9.7 <u>+</u> 0.6	53.2 ± 4.9	14.6 <u>+</u> 0.3	27.4 <u>+</u> 0.9	15.05 <u>+</u> 4.6	54.84 <u>+</u> 0.7	13.8 ± 0.6
GROUPS D (100mg / kg)	10.29 <u>+</u> 2.0	55.3 ± 2.5	15.5 ± 0.8	28.02 <u>+</u> 3.1	15.0.6 <u>+</u> 3.0	53. 74 <u>+</u> 1.0	14.0 ± 4.9
GROUP E (Control)	10.6 ± 0.6	56.7 ± 4.8	16.0 ± 0.0	28.22 <u>+</u> 1.0	15.09 ± 1.0	53.49 <u>+</u> 0.4	14.7 ± 6.0

Discussion

Administration of ethanolic leaf extract of Croton Zambesicus to rats for 21 days produced a dose dependent reduction in PCV, Hb, RBC, MCHC, MCH and WBC. Elevation of MCV values of the treated rats was also recorded. The dose dependent reduction in PCV, Hb and RBC may have resulted from hemolytic activity of the extract leading to the observed reduction. It could also be a result of suppressive action of the extract on erythropoiesis. These reductions result in anaemic condition. Keele et al (1983) reported that normocytic, hypochronic anaemia results when there is a reduction in MCHC and MCH, while MCV as normal. A similar case was observed in this study though the MCV values are slightly elevated. Total white blood cells were observed to have reduced following the administration of the extract to rats. The decrease observed may have resulted from suppression of leucocytosis by the extract and also from the suppression of their production in the bone marrow. Further studies to confirm this as well as evaluate its mechanism of action are suggested.

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References

Adjanohaun, E. J., Adjakidje, V. and de Souza, S. (1989) Contribution to Ethnobotainical and Floristic Studies in Benin Republic Vol. 1. Agency for Cultural and Technical Cooperation. P. 245.

Abo, K. A. Ogunleye, Ashidi, J. S (1999) Antimicrobial Potential of Spondias mombin, Croton zambesicus, and Zygotritonia crocea. Phytotherapy Research. 13 (6) 494 – 497.

Block, S. Stevigny, C; De Pauw – Gillet, M. C; de Hoffman, E. Llabres, G. Adjakidje, V. and Quetin – Lectercq, J. (2002) ent – Trachyloban - 3

ol, a New Cytotoxic Diterpene from *Croton zambesicus* . *Planta Medica*. 68. 647 - 648.

Jain, N. C. (1986) *Veterinary Haematology* (Jain N C Ed) Lea and Ferbiger, Philadelphia.

Keele, C. A., Neil, and Joels, N. (1983) *Sampson Wright's Applied Physiology*. 13th ed, Oxford University Press.

Mekkawi, A C, (1985) The Essential Oil of *Croton zambesicus Fitoterapia* 56 (3): 181 – 183.

Menut, C; Lamaty, G; Bessiere J. M: Suleiman, A. M; Fendro, L; Maidou, E and Denamgania, J. (1985). Aromatic Plants of Tropical *Central* Africa. *Journal of Essential Oil Research*. 7 (4): 419 – 422.

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