

NIGERIAN VETERINARY JOURNAL

ISSN 0331-3026

Nig. Vet. J., June 2017

Vol 38 (2): 104-116. ORIGINAL ARTICLE

Prevalence of Bovine Brucellosis and Analysis of Risk Factors in Resident Cattle Herds of Kanke Local Government Area, Plateau State, Nigeria

Agada, C. A.¹*; Goden, C. P.² and Ogugua, J. O.³

¹Department of Veterinary Public Health and Preventive Medicine, College of Veterinary Medicine, Federal University of Agriculture, Makurdi, Nigeria; ²Veterinary Section, Department of Agriculture, Kanke Local Government Council, Plateau State; ⁴Department of Veterinary Public Health and Preventive Medicine, Faculty of Veterinary Medicine, Universit3 of Ibadan, Ibadan, Nigeria. *Corresponding author: Email: caysla@gmail.com; Tel No:+2348036506966

SUMMARY

Brucellosis an important zoonotic disease is endemic in Nigeria resulting to huge economic losses in livestock and loss of man hour in infected people. Information about the prevalence and risk factors for the disease in resident cattle herds in the North Central Zone of Nigeria is however lacking. A cross-sectional study was conducted to determine the prevalence of bovine brucellosis and the risk factors associated with the disease in Kanke Local Government Area (LGA) of Plateau State. A total of 479 resident cattle sera from 39 herds in the four districts of the LGA were examined for antibodies against *Brucella* species using Rose Bengal plate test (RBPT) and competitive Enzyme-Linked Immunosorbent Assay (cELISA). Risk factors responsible for the occurrence of the disease in the herds were investigated using pre-tested structured questionnaire. The strength of association between risk factors and seropositivity to brucellosis was measured using logistic regression analysis. Out of the 479 sera examined, 1.0% (5/479) and 3.6% (18/479) were positive for B. abortus antibodies using RBPT and cELISA respectively. The herd prevalences were 10.3% (4/39) and 38.5% (15/39) with the RBPT and cELISA, respectively. There was a significant association between seroprevalence of brucellosis and herd size (OR: 4.3, 95% CI: 1.0.-18.3; P=0.05) as well as a number of milking cows (OR: 4.7, 95% CI: 1.2.-18.9; P=0.03). The study found brucellosis to be prevalent in resident cattle herds in the study area and milk from cows in these herds are likely to transmit the disease to humans.

Key words: Brucellosis, Resident herds, Cattle, Kanke LGA.

INTRODUCTION

Brucellosis is a chronic disease of animals caused by Gram negative, facultative nonmotile, intracellular bacteria of the genus *Brucella* (OIE, 2009). It is a contagious systemic disease primarily of ruminants, characterized by inflammation of the genital organs and foetal membranes, abortion, sterility, and formation of localized lesions in the lymphatic system and joints (CDC, 2011). The disease has a worldwide distribution but has been eradicated from the livestock populations of most European countries, Japan, Canada and the United States of America (USA) (Radostits et al 2000; (WHO, 2001). Brucellosis is. however, prevalent in parts of Asia (Chahota et al., 2003); South America (Dias et al., 2009); and Africa (Ogugua et al., 2015). In cattle, the disease is transmitted by contact with infected uterine discharges and maternal transfer either by suckling or invivo (Corbel, 2006). Humans are infected by inhalation, contact of abraded skin with infected materials as well as consumption of unpasteurised milk originating from infected animals (CFSPH, 2009; WHO, 2004). It is, therefore an occupational disease to veterinarians, abattoir workers, herdsmen, hides and skin factory workers as well as laboratory personnel (Falade, 2002; Traxler et al., 2013). Infection in cattle may be lifelong and in naïve cattle population, abortion storm (abortion rates varying from 30 to 70%) may occur (CFSPH, 2009; Godfroid et al., 2004; Pappas et al., 2005). abortion. After the first subsequent pregnancies are usually delivered normal but Brucella is still shed in the milk and uterine discharges of such animals (CFSPH, 2007). Since the reproductive performance of these carrier animals seems unaffected, they are retained in herds especially in developing countries like Nigeria despite the presence of pathognomonic clinical signs in some cases, making effective control programmes extremely difficult (Mai et al., 2012). In humans, it causes undulating fever, and when left untreated could result in complications such as meningitis. epididymo-orchitis, arthritis (Safirullah et al., 2014) and death due to cardiac involvement in about 5% of the cases (Chadda et al., 2004; Esuruoso et al., 2005). In cattle herds, brucellosis results in huge economic losses due to decreased calving percentage, culling for infertility, decreased milk production, abortion, stillbirth or birth to weak calves; as well as loss of man hours in infected people (McDermolt *et al.* 2002; Ocholi *et al.* 2004; Adamu, 2009). The presence of brucellosis in cattle herds portends a major public health problem especially to individuals with regular contact with cattle as well as the members of the general public who consume unpasteurised milk and milk products of cattle origin in Nigeria.

Brucellosis remains a problem in Nigeria due to lack of official policy for the control of the disease (Ibironke et al., 2008), uncontrolled movement of slaughter cattle within and from neighbouring countries (Ogundipe, 2001; Cadmus et al., 2008), nomadism (Mai et al., 2012) and poor knowledge and practices concerning the diseases among farmers and other risk groups (Adesokan et al., 2013). In Nigeria, varying prevalence rates have been recorded in different parts of the country: the prevalence of 7.8% and 1.9% was recorded from Oyo and Lagos (Ogugua et al., 2015); 20.0% prevalence was recorded in slaughter cattle in Zamfara State (Lawal et al., 2012), a within herd prevalence of 32.2% was recorded in a prison cattle farm in Sokoto State (Junaidu et al., 2008); in three states of Adamawa, Kano, Kaduna prevalence of 29.2%, 26.7% and 23.3%, respectively was recorded (Mai et al, 2012); 14.1% prevalence was recorded in Obudu, Cross River State (Nanven et al., 2013). In Plateau State, the prevalence of 37.3%, 2.5% and 3.7% was recorded in Bassa, Riyom and Jos South Local Governments Areas (LGAs), respectively (Nanven et al., 2013). In Nigeria, the most popularly consumed

In Nigeria, the most popularly consumed animal products are those of cattle origin (Alimi, 2013; Rauf, 2012). The livestock production system in Nigeria includes the nomadic, semi nomadic and intensive system. Although with time, population increase has resulted in corresponding increase in demand for livestock products, cattle production in Nigeria is concentrated in the hands of the nomadic Fulani herdsmen (Ibironke et al., 2008). However, in Plateau State, many farmers are involved in agro-pastoralist farming system whereby cattle are raised in small herds in the backvard of the farmers where grasses are cut and given to the animals or the animals are taken to the nearby communal grazing lands. The animals are therefore resident in the communities and not involved in long distance movement in search of feed and water. In Nigeria, where no control policy is employed to control brucellosis, grazing of cattle herds in communal lands result to exchange of diseases like brucellosis between different herds (Bertu et al., 2010; Hesterberg et al., 2008). In Plateau State, past studies on brucellosis were focused on pastoral herds (Nanven et al., 2013), cattle (Bertu. 2014) and settlements small ruminants (Bertu et al., 2010). Therefore, information regarding the prevalence and risk factors for brucellosis in resident herds in the state is scarce. This study, therefore used the RBPT, cELISA and questionnaire to determine the prevalence of bovine brucellosis and the associated risk factors in the resident herds of Kanke LGA of Plateau State, Nigeria.

MATERIALS AND METHODS Study area

Kanke LGA is in the central zone of Plateau State and located between latitude 80⁰ 24 North and Longitude 80⁰ 32 and 100⁰38 East. The LGA shares boundary with Bauchi State in the North, Pankshin LGA in the West, Kanam LGA in the East and Langtang North LGA in the South. The LGA has an area of 7,808. 85 km² and population of 268,000 people (NPC, 2006). The majority of the inhabitants are farmers while among others are civil servants, businessmen, artisans etc. Most of the farmers are crop farmers some of whom keep a few herds of cattle that make up the resident herds. These resident herds have grass cut in the fields and brought home for them as well as graze within the vicinity of the homesteads in private or communal grazing lands. There are pastoral herds in the area which are not indigenous but are reared by nomadic Fulani herdsmen that settle briefly and eventually move on in search of feed and water. The pastoral herds were not included in this work. The LGA has four districts namely; Amper, Kabwir, Ampang and Garam in order of decreased livestock population. Out of the four, Amper is the only district that harbours cattle market due to a suitable grazing topography.

Study population and design and animal sampling

A cross-sectional survey was conducted between January and June 2015 among resident cattle aged over six months. With the statistical formula $n = \frac{1.96^2 \cdot P \cdot (1-P)}{d^2}$, the sample size of 33 herds was calculated using the prevalence of 9.6% earlier recorded from cattle herds screened in northern Plateau State (Nanven et al., 2013). A non-response rate of 10% was added giving a total sample size of 37 although, a total of 39 herds were screened in the study. An interviewer administered questionnaire was issued to each herd owner. Also information about sex, age, breed of individual animal were collected along with the sample. About 5ml of blood collected from the jugular vein of each cattle after proper restraint using sterile needle and syringes was dispensed into centrifuge tubes and labelled accordingly. These tubes were placed in a slanting position to enhance serum separation, kept in a flask containing ice pack and transported to the laboratory at the Department of Public Health and Preventive Medicine College of Veterinary Medicine University of Agriculture, Makurdi. The blood samples were centrifuged at 3000rpm for 5 minutes, the sera decanted into serum vials and stored at -20° c until assav.

Serological tests

The serum samples were tested for *Brucella* antibodies by RBPT and cELISA.

Rose Bengal plate test (RBPT)

The serum samples were tested for *Brucella* antibodies by RBT as described by OIE (2009). The RBPT antigen consisting of standardized *B. abortus* antigen from the Animal and Plant Health Agency (APHA), Surrey KT15 3NB, U.K. was used to carry out the test. Briefly, equal volumes (30 μ l) of antigen and test serum were mixed thoroughly on a plate using a stick applicator and the plate was rocked for 4 minutes. The appearance or absence of agglutination (rough or smooth clumps with rim edges) was scored positive (+) and negative (-), respectively.

Competitive enzyme-linked immunosorbent assay (cELISA)

The cELISA kit was sourced from the APHA. The kit contained cELISA plate and reagents. The plate was coated with the lipopolysaccharide (LPS) of *B. melitensis* M16. The reagents included control sera, diluting buffer, conjugate, washing solution, chromogen and stopping solution. The reagents were reconstituted as directed by the manufacturers. The test was performed according to the manufacturer's instructions. Positive samples had a clear appearance

whereas negative samples appeared orange in colour. The optical density (OD) was measured at 450nm using a microplate ELISA reader. A positive/negative cut-off was calculated as 60% (as instructed by manufacturers) of the mean of the OD of the conjugate control wells. Samples in wells with OD equal to or less than the cut-off point were scored positive, while those above were negative.

Data analysis

Data analysis was performed using Stata Version 12. Group differences were tested for by using chi-square statistics for categorical variables. A multivariable adjusted logistic regression was carried out using all the variables that were statistically significant at the 10% level with the main outcome measure (RBPT) in bivariate analysis. All tests were two-tailed and statistical significance was set at p<0.05.

RESULTS

The results of the study show the individual prevalence of brucellosis to be 1.0% (5/497) and 3.8% (18/497) as well as the herd prevalence of 10.26% (4/39) and 38.46% (15/39) with the RBPT and cELISA, respectively (Tables I and III). While the brucellosis prevalence was not found to be associated with district, breed, sex and age on individual basis, it was found to be

Variable	Characteristi	Seropositive animals based on RBT				Odds	95%CI	p-
	с	Positive	%	Negative	%	ratio		value
		N=5	1.0	N=492	99.0			
District	*Others	2	1.1	177	98.9	1		
	Amper	3	0.9	315	99.1	0.84	0.14-5.09	0.84
Breed	**Others	1	5.6	17	94.4	1		
	Bunaji	4	0.8	475	99.2	0.14	0.02-1.35	0.09
Sex	Male	2	0.7	274	99.3	1		
	Female	3	1.4	218	98.6	1.89	0.31-11.38	0.26
Age	1-2 years	1	0.6	180	99.4	1		
	3-8 years	4	1.3	312	98.7	2.31	0.26-20.8	0.25

TABLE I: Factors associated with the individual level prevalence of brucellosis among resident cattle screened in Kanke LGA of Plateau State as measured by RBT

*Others include Ampang, Garam and Kabwir; **Others include Muturu and Sokoto Gudali

TABLE II; Results of logistic regression a	analysis of a	variable significan	t at 10% leve	el with main
outcome measure RBT in bivariate analys	sis			

		2				
	Category	Brucella infe	ction	OR	95%CI	P -value
Variables		Positive	Negative			
		n=5(1.0%)	N=492(99.0%)			
Breed	**Others	1(5.6)	17 (94.4)	1		
	Bunaji	4 (0.8)	475 (99.2)	7.0		
					0.74-65.88	0.09
breed	Bunaji	4 (0.8)	475 (99.2)	7.0	0.74-65.88	0.09

*Others include Ampang, Garam and Kabwir; **Others include Muturu and Sokoto Gudali

TABLE III: Factors associated with the individual level prevalence of brucellosis among re-	sident
cattle screened in Kanke LGA of Plateau State as measured by cELISA	

Variable	Characteristic	Seropositive animals based on cELISA			ELISA	Odds	95%CI	p-value
		positive	%	Negative	%	ratio		
		N=18	3.6	N=479	96.4			
District	*Others	5	2.8	174	97.2	1		
	Amper	13	4.1	305	95.9	1.5	0.52-4.23	0.32
Breed	**Others	1	5.6	17	94.4	1		
	Bunaji	17	3.5	462	96.5	0.6	0.1-4.98	0.49
Sex	Male	11	4.0	265	96.0	1		
	Female	7	3.2	214	96.8	0.79	0.30-2.07	0.32
Age	1-2 years	5	2.8	176	97.2	1		
	3-8 years	13	4.1	303	96.5	1.51	0.53-4.31	0.23

*Others include Ampang, Garam and Kabwir; **Others include Muturu and Sokoto Gudali

significantly associated with the number of milking cows (OR: 4.7, 95%CI: 1.2-18.9; P=0.03) and herd size (OR: 4.26, 95%CI: 1.0-18.3; P=0.05) in the herds. (Tables II and V).

DISCUSSION

The prevalence of brucellosis recorded in this study (1.0%) showed that brucellosis is prevalent in cattle in the study area. The prevalence could be attributed to the lack of official policy for the control of the disease Nigeria (Cadmus et al., in 2006), uncontrolled movement of livestock within and from neighbouring countries (Ogundipe, ignorance of the 2001), mode of transmission the disease among farmers (Adesokan et al, 2013), retaining of animals showing pathognomonic signs of the disease (Mai et al., 2012) and many other factors. However, the prevalence recorded is lower than the 26.3% in three northern states of Nigeria (Mai et al., 2012), 7.1% in Kaduna

State (Mbuk et al., 2011), 9.6% in Plateau State (Nanven et al., 2013), 42.1% in Obudu Cross River State (Nanven et al., 2013) and 8.4% in Cameroon (Bayemi et al., 2009). This low prevalence could be due to the fact that the cattle herds in the study area are relatively small in size and are therefore at low risk of exposure to the disease (Megersa et al., 2011). It could also be as a result of the fact that the herds in the area are resident cattle and are not involved in seasonal migration which is common with cattle herds in Nigeria (Mbuk et al., 2011). Brucellosis prevalence has been reported to be higher in pastoral than resident herds (Unger et al., 2003) due to increased exposure potential as a result of movement from one location to another; interacting and sharing grazing lands and watering points with other potentially infected cattle herds and other animals (Mai et al., 2012; Matope et al., 2011).

Variable	Characteris	Seropositi	ve anima	ls based on	RBT	Odds	95%CI	p-value
	tic	Positive	%	Negative	%	ratio		
		n=4	10.3	n=35	89.7			
Herd size	1and2	2	6.5	29	93.5	1		
	3	2	25.0	6	75.0	4.83	0.56-41.41	0.10
Number of	0, 1 and 2	2	6.3	30	93.7	1		
milking cows	3	2	28.6	5	71.4	6.00	0.68-52.89	0.07
Period of	1 and 2	1	5.6	17	94.4	1		
existence of								
herd	3	3	14.3	18	85.7	2.83	0.27-29.95	0.36
Originating	purchases	1	5.3	18	94.7	1		
herd	Inheritance	3	15.0	17	85.0	3.17	0.30-33.58	0.32
Abortion	No history	1	5.6	17	94.4	1		
	of abortion							
	History of	3	14.3	18	85.7	2.00	0.19-21.57	0.502
	abortion							
Retained	No history	1	7.7	12	92.3	1		
placenta	of retained							
	placenta							
	History of	3	11.5	23	88.5	1.57	0.15-16.71	0.59
	retained							
	placenta							
Knowledge	Good	3	9.4	29	90.6	1		
of brucellosis	Poor	1	14.3	6	85.7	1.61	0.14-18.26	0.56
in animals								
Attitudes of	Good	0	0.0	6	100			
farmers	Poor	4	12.1	29	87.9			
Practices of	Good	1	25.0	3	75.0	1		
farmers	Poor	3	8.6	32	91.4	0.28	0.02-3.62	0.36

TABLE IV: Prevalence and risk factors associated with the occurrence of brucellosis in resident cattle herds as measured by RBT in Kanke LGA

TABLE V: Results of logistic regression analysis of variables significant at 10% level with main outcome measure RBT in bivariate analysis

Variables	Category	Brucella infection		OR	95%CI	P value	
		Positive	Negative				
		n=4(10.3%)	n=35(89.7%)				
Herd size	Small	2 (6.5)	29 (93.5)	1			
	Large	2(25.0)	6(75.0)	4.3	1.0-18.3	0.05	
No of	Few	2 (6.3)	30 (93.7)				
milking	number			1			
cows	Large	2 (28.6)	5 (71.4)				
	number			4.7	1.2-18.9	0.03	

The study found the prevalence of brucellosis to be significantly associated with herd size (OR: 4.26, 95%CI: 1.0.-18.3; P=0.05). This agrees with other investigators (Jergefa *et al.*, 2009; Makita *et al.*, 2011; Megersa *et al.*,

2011; Unger *et al.*, 2003) that recorded the prevalence of brucellosis to be higher in large herds than small herds. This is in consonance with the epizootiological rule of "large herds, large incidence and small herds, low

incidence" as stated by Akakpo and Bornarel, (1987). Large herd sizes have been shown to increase the exposure to brucellosis especially after abortion or calving by *Brucella* infected animals because of the higher stocking density as compared to small sized herds (Megersa *et al.*, 2011).

Number of milking cows was found to be significantly associated with the prevalence of brucellosis in the study (OR: 4.7, 95%CI: 1.2.-18.9; P=0.03). The pregnancy period which precedes milking is noted to be associated with brucellosis (Swai and Schoonman, 2010). This is because Brucella species have tropism for the pregnant uterus ervthritol sugar because which is preferentially metabolised by the organism is produced in the placenta (Neta et al., 2008). The multiplication of the Brucella organism results in inflammation that leads to abortion which may not occur in subsequent pregnancies (Corbel, 2006). However, such animals may become latent carriers that could only be detected by serological during and tests after pregnancies (CFSPH, 2009). Retaining of such animals in the herds is common in Nigeria (Mai et al., 2012) and tests during the milking periods may detect infection in such animals. Also, some cows infected inutero may not be serologically positive until during and after pregnancy (Forbes and Steele, 1989). This indicates a risk of infection to the cattle owners who have regular contact and custom of consuming unpasteurised milk and milk products.

The prevalence recorded with the RBPT (1.0%) is lower than that recorded with the cELISA (3.6%). This is in contrast with the findings of other studies that recorded higher prevalence with the RBPT than the cELISA (Mohammed *et al.*, 2011; Mai *et al.*, 2012; Cadmus *et al.*, 2013). This can be explained by the fact that serological tests differ in their ability to detect a particular immunoglobulin (Beh, 1974). The immunoglobulin isotypes found in the blood in early or acute infections are the IgM and IgG1 (Ismail *et al.*, 2002),

which may not be seen in cases with insidious onset, in chronic, recurrent and relapse cases (Serra and Viñas, 2004) where IgG2, IgG3 and IgA are predominant (Henk et al., 2003; Diaz et al., 2011). While the RBPT is better suited for detecting acute cases (Chenais et al., 2012) (i.e. the IgM and IgG1), the cELISA is the test of choice in chronic cases (Smits et al., 2003). This may, therefore suggest that most of the cases in this study were of the chronic form of brucellosis which is supported by a previous report that many cases of brucellosis in endemic areas could be in chronic or relapse stage of the disease (Serra and Viñas, 2004). The IgG ELISA has been used to monitor chronic and relapse infections because of its better ability to detect IgG and IgA in sera (Smits et al., 2003). However, the discrepancy between the two tests could also be due to the presence of non-specific antibodies due to infection with antigenically related bacteria like Yersinia enterocolitica 0:9, Salmonella Urbana, Escherichia coli 0:157 and Francisela tularensis (Bowden et al., 1997; Chenais et al., 2012). Although, Yersinia *enterocolitica* 0:9, the most antigenically related bacteria to Brucella, has been noted by Shey-Njila et al. (2005) to belong to temperate regions, but it has been isolated in Nigeria (Okwori et al., 2009).

The use of cELISA to complement the RBPT has been noted as one of the best combinations of specificity and sensitivity especially in areas where vaccination is practiced (Marín et al., 1999). In Nigeria however, vaccination is not generally practiced since there is no official policy on control of brucellosis (Aworh et al., 2013; Ibironke et al., 2008). The RBPT however, has been described as being superior to the cELISA and therefore cannot be confirmed with an inferior test like the cELISA (Ducrotoy et al., 2014; Megersa et al., 2011). The RBPT is therefore recommended as the test of choice in endemic and resource-poor countries where vaccination is not generally practiced like Nigeria because of its simplicity, relatively low cost and high standard in testing for brucellosis (Mcgiven,

2013). Meanwhile, the ELISA kit used in this study was manufactured in the United Kingdom which is an area of low prevalence and the cut-off point (60%) was set as the Brucella infection rate applies to the UK. Since cut-off points are set as the points of highest accuracy (minimal false positive and negative) results, it cannot be extrapolated from areas devoid of brucellosis and with good hygienic conditions to areas where brucellosis is endemic (Greiner and Gardner, 2000). This is because in endemic areas cattle population sometimes acquire antibodies due to possible contact with the organism, but without having the disease (Corbel, 2006). In Adamawa region of Cameroon for instance, 50% was suggested to be a better cut-off point using the cELISA in the cattle (Bronsvoort et al., 2009). The OIE, therefore, recommends that cut-off points for ELISA be validated under local settings (OIE, 2009).

Although the study found no association between prevalence of brucellosis and sex, higher seropositivity was recorded among the females than males. This result is in agreement with other studies that found prevalence of brucellosis to be higher in cows than bulls (Dinka and Chala, 2009; Junaidu et al., 2011) but contrasts some other studies that reported higher prevalence in males (Chimana et al., 2010; Cadmus et al., 2013). This finding may be due to the fact that unlike the bulls which are sooner sold for beef once they attain market weight, most cows are retained for a longer period for reproduction and milk production in Africa (Mangen et al., 2002). Such longer period of existence in endemic areas has been associated with greater chances of exposure to brucellosis (Kebede et al., 2008; Megersa et al., 2011). In addition, bulls have been reported to show limited immunological response to Brucella infection (Berhe et al., 2007).

Despite the findings of the study, some limitations were observed. Most of the cattle screened were of the Bunaji because it is the most common breed in the study area. Also, cattle screened in Amper District were more than that in the other three districts and this was due to the fact that more animals are reared in this district. These discrepancies in numbers might have introduced bias in the study. Also, the *Brucella* organisms responsible for the disease were not isolated to confirm the *Brucella* species responsible for brucellosis in the resident herds screened.

CONCLUSION

This study shows that brucellosis is prevalent in resident herds in the study area although at a low level. There is need to test individuals in regular contact with these animals for brucellosis. Food products derived from these animals should be properly cooked to protect the consumers from Brucella infection. In addition. individuals in regular contact with the infected animals should be mindful of personal protection especially when assisting in calving. Also, the herd owners and members of the public who consume unpasteurised milk in the area should be educated on the economic and public health importance of the disease. Finally, further studies should confirm brucellosis in the study area by isolation of the Brucella species responsible for the disease as well as validation of cut-off points for serological methods like cELISA in such local setting.

REFERENCES

- ADESOKAN, H. K., ALABI, P. I., STACK, J. A. and CADMUS, S. I. B. (2013). Knowledge and practices related to bovine brucellosis transmission amongst livestock workers in Yewa, south-western Nigeria. Journal of South African Veterinary Association, 84(1), Art N 121, 5 pages. doi:10.4102/jsava.v84i1.121.
- AKAKPO, J. A. and BORNAREL, P. (1987). Epidémiologie des en Afrique brucelloses animales tropicale : enquêtes clinique sérologique bactériologique. et Revue Scientifique et

Technique (International Office of Epizootics), 6(4), 981–1027.

- ALIMI, R. S. (2013). An analysis of meat demand in Akungba-Akoko, Nigeria. *Nigerian Journal of Behavioural Sciences*, 1, 96–104.
- AWORH, K., OKOLOCHA, M. Е., KWAGA, FASINA. F., J., LAZARUS, D., SULEMAN, I., ... NSUBUGA, P. (2013). Human seroprevalence brucellosis: and associated exposure factors among abattoir workers in Abuja, Nigeria -2011. Pan African Medical Journal, 8688, 1–9.
- BAYEMI, P. Н., WEBB, E. С., NSONGKA, M. V, UNGER, H. and NJAKOI, H. (2009). Prevalence of Brucella abortus antibodies in serum of Holstein cattle in Cameroon. Tropical Animal Health and Production, 41(2), 141-4. doi:10.1007/s11250-008-9184-8.
- BEH, K. J. (1974). Quantitative distribution of *Brucella* antibody amongst immunoglobulin classes in vaccinated and infected cattle. *Research in Veterinary Science.*, *17*(1), 1–4.
- BERHE, G., BELIHU, K. and ASFAW, Y. (2007). Seroepidemiological Investigation of Bovine Brucellosis in the Extensive Cattle Production System of Tigray Region of Ethiopia. *International Journal of Applied Research in Veterinary Medicine*, 5(2), 65–71.
- BERTU, W. J, AJOGI, I., BALE, J. O. O., KWAGA, J. K. P. and OCHOLI, R.
 A. (2010). Sero-epidemiology of brucellosis in small ruminants in Plateau State , Nigeria. *African Journal of Microbiology Research*, 4(19), 1935–1938.
- BERTU, W. J. (2014). Prevalence, Bacteriology and Molecular Epidemilogy of Brucellosis in Two Cattle Settlements in Plateau State.

PhD Thesis. Department of Veterinary Public Health and Preventive Medicine, Ahmadu Bello University, Zaria, Nigeria.

- BOWDEN, L. A., VERGER, J., GRAYON, M. and CLOECKAERT, A. (1997). Rapid Identification of Rough Brucella Isolates by a Latex Coagglutination Assay with the 25-Kilodalton Outer Membrane Protein Rough-Lipopolysaccharideand Specific Monoclonal Antibodies. Clinical and Diagnostic Laboratory Immunology, 4(5), 611–614.
- BRONSVOORT, B.M.D., KOTERWAS, B., LAND, F., HANDEL, I.G., TUCKER, J., MORGAN, K. L., TANYA, V.N., ABDOEL, T.H. and HENK L. SMITS, H.L. (2009). Comparison of flow assay for brucellosis antibodies with reference to cELISA test in West African Bos Indicus. PloS One, 4(4), e5221. Retrieved from doi:10.1371/journal.pone.0005221.
- CADMUS, S I B, ADESOKAN, H. K. and STACK, J. (2008). The use of the milk ring test and rose bengal test in brucellosis control and eradication in Nigeria. *Journal of South African Veterinary Association*, 79(3), 113– 115.
- CADMUS, S I B, IJAGBONE, I. F., OPUTA, H. E., ADESOKO, H. K. and STACK, J. A. (2006). Serological Survey of Brucellosis in Livestock Animals and Workers in Ibadan, Nigeria. *African Journal of Biomedical Research*, 9, 163–168.
- CADMUS, S., ALABI, P. I., ADESOKAN, H. K., DALE, E. J. and STACK, J.
 A. (2013). Serological investigation of bovine brucellosis in three cattle production systems in Yewa Division, south-western Nigeria. *South African Veterinary Association*, 84(1),

- CADMUS, S. I B, ALABI, P. I., ADESOKAN, H. K., DALE, E. J. and STACK, J. A. (2013). Serological investigation of bovine brucellosis in three cattle production systems in Yewa Division, southwestern Nigeria. Journal of South African Veterinary Association, 84(1), 1–6.
- CFSPH. (2007). Bovine Brucellosis: Brucella abortus. Center for Food Security and Public Health, College of Veterinary Medicine, Iowa State University, Ames, Iowa., pp. 1–6.
- CFSPH. (2009a). Bovine Brucellosis : Brucella abortus. Center for Food Security and Public Health, College of Veterinary Medicine, Iowa State University, Ames, Iowa., pp. 1–6.
- CFSPH. (2009b). Porcine and Rangiferine Brucellosis : Brucella suis. Center for Food Security and Public Health, College of Veterinary Medicine, Iowa State University, Ames, Iowa. (pp. 1–6).
- CHAHOTA, R., SHARMA, M. and KATOCH, R. C. (2003). Brucellosis outbreak in an organized dairy farm involving cows and in contact human beings, in Himachal Pradesh, India. *Veterinarski Arhiv*, 73(2), 95–102.
- CHENAIS, E., BAGGE, E., LAMBERTZ, S. T. and ARTURSSON, K. (2012). Yersinia enterocolitica serotype O:9 cultured from Swedish sheep showing serologically false - positive reactions for *Brucella* melitensis. *Infection Ecology & Epidemiology*, 2, 19027. doi:http://dx.doi.org/10.3402/iee.v2i

doi:http://dx.doi.org/10.3402/iee.v21 0.19027.

CHIMANA, H., MUMA, J. B., SAMUI, K. L., HANGOMBE, Β. М., MUNYEME, M., MATOPE, G., PHIRI, A.M., SKJERVE, E. and TRYLAND, M. (2010). А study comparative of the of brucellosis seroprevalence in commercial and small-scale mixed dairy-beef cattle enterprises of Lusaka province and Chibombo district, Zambia. *Tropical Animal Health and Production*, 42, 1541– 1545.

- CORBEL, M. J. (2006). Brucellosis in humans and animals (pp. 1–102). World Health Organization, 20 Avenue Appia, 1211 Geneva 27, Switzerland. Retrieved from http://www.who.int/csr/resources/pu blications/Brucellosis.pdf May, 2014.
- DIAS, R. A., GONÇALVES, V. S. P. and FIGUEIREDO, V. C. F. (2009). Epidemiological situation of bovine brucellosis in the State of São Paulo, Brazil. Arquivo Brasilero Medicina Veterinaria Zooteccnia, 61(1), 118– 125.
- DIAZ, R., CASANOVA, A., ARIZA, J. and MORIYO, I. (2011). The Rose Bengal Test in Human Brucellosis : A Neglected Test for the Diagnosis of a Neglected Disease. *PLoS Neglected Tropical Diseases*, 5(4), 1–7.
- DINKA, H. and CHALA, R. (2009). Seroprevalence Study of Bovine Brucellosis in Pastoral and Agro-Pastoral Areas of East Showa Zone, Oromia Regional State, Ethiopia. *American-Euroasian Journal of Agricultural and Environmental Sciences*, 6(5), 508–512.
- DUCROTOY, M. J., BERTU, W. J., OCHOLI, R. A., GUSI, A. M., BRYSSINCKX, W., WELBURN, S. and MORIYO, I. (2014). Brucellosis as an Emerging Threat in Developing Economies : Lessons from Nigeria. *PLos One*, 8(7), e3008.
- ESURUOSO, G. O., IJAGBONE, I. F. and OLUGASA, B. O. (2005). *Introductory epizootiology* (Second Edi., p. 181). Ibadan: VetAcademic

- Resources Publishers and Consultants, U.I.P.O. Box 14400, Ibadan, Oyo State, Nigeria.
- FALADE, S. (2002). A case of possible brucellosis relapse in a veterinarian. *Tropical Veterinarian*, 20(4), 226– 230.
- FORBES, L. B. and STEELE, T. B. (1989). An outbreak of *Brucella* abortus biovar 2 in Canadian cattle. *Canadian Veterinary Journal*, 30, 888 – 893.
- GREINER, M. and GARDNER, I. A. (2000). Epidemiologic issues in the validation of veterinary diagnostic tests. *Preventive Veterinary Medicine*, 45, 3–22.
- HESTERBERG, U. W., BAGNALL, R., PERRETT, K., BOSCH, B., HORNER, R. and GUMMOW, B. (2008). A serological prevalence survey of *Brucella* abortus in cattle of rural communities in the province of KwaZulu-natal, South Africa. *Journal of the South African Veterinary Association*, 79(1), 15–8.
- IBIRONKE, A. A., MCCRINDLE, C. M. E., FASINA, F. O. and GODFROID, J. (2008). Evaluation of problems and possible solutions linked to the surveillance and control of bovine brucellosis in sub-Saharan Africa, with special emphasis on Nigeria. *Veterinaria Itliana*, 44(3), 549–56.
- ISMAIL, T. F., SMITS, H., WASFY, M. O., MALONE, J. L., FADEEL, M. A. and MAHONEY, F. (2002). Evaluation of Dipstick Serologic Tests for Diagnosis of Brucellosis and Typhoid Fever in Egypt. *Journal of Clinical Mcrobiology*, 40(9), 3509–3511.
- JERGEFA, T., KELAY, B., BEKANA, M., TESHALE, S., GUSTAFSON, H. and KINDAHL, H. (2009). Epidemiological Study of Bovine Brucellosis in Three Agro-ecological

Areas of Central Oromiya, Ethiopia. *Revue Scientifique et Technique* (*International Office of Epizootics*), 28(3), 933–43.

- JUNAIDU, A. U., OBOEGBULEM, S. I. and SALIHU, M. D. (2008). Seroprevalence of Brucellosis in Prison Farm in Sokoto, Nigeria. *Asian Journal of Epidemiology*, 1(1), 24–28.
- JUNAIDU, A. U., OBOEGBULEM, S. I. and SALIHU, M. D. (2011). Serological survey of *Brucella* antibodies in breeding herds. *Journal* of Microbiology and Biotechnology Resesources, 1(1), 60–65.
- KEBEDE, T., EJETA, G. & AMENI, G. (2008). Seroprevalence of bovine brucellosis in smallholder farms in central Ethiopia (Wuchale-Jida district). *Revue de Médecine Vétérinaire*, 159(1), 3–9.
- LAWAL, N., EGWU, G. O., TAMBUWAL,
 F. M., JUNAIDU, A. U.,
 ABUBAKAR, M. B., MAGAJI, A.
 A., ... TAMBUWAL, Z. A. (2012).
 Prevalence of *Brucella* abortus antibodies in bovine serum from Gusau modern abattoir , Zamfara State , Nigeria. *Scientific Journal of Microbiology*, 1(4), 91–96.
- MAI, H. M., IRONS, P. C., KABIR, J. and THOMPSON, P. N. (2012). A large seroprevalence survey of brucellosis in cattle herds under diverse production systems in northern Nigeria. *Biomed Central Veterinary Research*, 8, 144.
- MAKITA, K., FÈVRE, E. M., WAISWA, C., EISLER, M. C., THRUSFIELD, M. and WELBURN, S. C. (2011). prevalence Herd of bovine brucellosis and analysis of risk factors in cattle in urban and periurban areas of the Kampala economic zone, Uganda. Biomed Central Veterinary Research, 7(1):

60.

- MANGEN, M. J., OTTE, J., PFEIFFER, D. and CHLONDA, P. (2002). Bovine brucellosis in Sub-Saharan Africa: Estimation of sero-prevalence and impact on meat and milk offtake potential.
- MARÍN, C. M., MORENO, E., MORIYÓN, I., DÍAZ, R. and BLASCO, J. M. (1999). Performance of competitive indirect enzyme-linked and immunosorbent assays, gel immunoprecipitation with native hapten polysaccharide, and standard serological tests in diagnosis of sheep brucellosis. Clinical and Diagnostic Laboratory Immunology, 6(2), 269–72.
- MATOPE, G., BHEBHE, E., MUMA, J. B., OLOYA, J., MADEKUROZWA, R. L., LUND, A. and SKJERVE, E. (2011). Seroprevalence of brucellosis and its associated risk factors in cattle from small holder dairy farms in Zimbabwe. *Tropical Animal Health and Production*, 43, 975–982.
- MBUK, E. U., AJOGI, I., BALE, J. O. O. and UMOH, J. U. (2011). Prevalence of *Brucella* antibodies in migratory Fulani cattle herds in Kaduna State, Nigeria. *Nigerian Veterinary Journal*, 32(1), 26–29.
- MCGIVEN, J. A. (2013). New developments in the immunodiagnosis of brucellosis in livestock and wildlife. *Revue Scientifique et Technique* (*International Office of Epizootics*), 32(1), 163–176.
- MEGERSA, B., BIFFA, D., NIGUSE, F., RUFAEL, T., ASMARE, K. and SKJERVE, E. (2011). Cattle brucellosis in traditional livestock husbandry practice in Southern and Eastern Ethiopia, and its zoonotic implication. Acta Veterinaria Scandinavica. 53(1), 24. doi:10.1186/1751-0147-53-24
- MOHAMMED, F. U., IBRAHIM, S., AJOGI, I. and OLANIYI, B. J. O.

(2011). Prevalence of Bovine Brucellosis and Risk Factors Assessment in Cattle Herds in Jigawa State. *ISRN Veterinary Science*, doi:10.5402/2011/132897

- NANVEN. M. A., BALE. J. O., KWANASHIE, C. N., AJOGI, I., NANVEN, M. B., NGBEDE, E. O. and MAURICE, N. A. (2013). Bacteriological and Serological Studies of Bovine Brucellosis on the Obudu Plateau, Cross River State, Nigeria. European Journal of Experimantal *Biology*, 3(6), 484–488.
- NANVEN, M. A., WUNGAK, S. Y., GANA, B. A., NANVEN, M. B., NGBEDE, E. O., IBRAHIM, M., ... GUGONG, V. T. (2013). Seroprevalence of bovine brucellosis in northern Plateau State, North Central Nigeria. Asian Pacific Journal of Tropical Disease, 3(5), 337–340.
- NETA, A. V. C., STYNEN, A. P. R., PAIXÃO, T. A., MIRANDA, K. L., SILVA, F. L., ROUX, C. M., ... SANTOS, R. L. (2008). Modulation of the bovine trophoblastic innate immune response by *Brucella* abortus. *Infection and Immunity*, *76*(5), 1897–907.
- OGUGUA, A. J., AKINSEYE, O. V., AYOOLA, M. C., STACK, J. and CADMUS, S. I. B. (2015). Risk factors associated with brucellosis among slaughtered cattle: Epidemiological insight from two metropolitan abattoirs in Southwestern Nigeria. *Asian Pacific Journal of Tropical Disease*, 5(9), 930– 936.
- OGUNDIPE, G. A. T. (2001). The Roles of Veterinary Quarantine Services in Monitoring the Movements of Animals and Disease Prevention in Nigeria. *Nigerian Veterinary Journal*, 23(1), 1– 15.
- OIE. (2009). Bovine brucellosis: Terrestrial Manual, Chapter 2.4.3. Version adopted by the World Assembly of

Delegates of the OIE in May, (pp. 1–35).

- OKWORI, A., MARTINEZ, Ρ., FREDRIKSSON-AHOMAA, М., AGINA, S. and KORKEALA, H. (2009).Pathogenic Yersinia enterocolitica 2/O:9 and Yersinia pseudotuberculosis 1/0:1strains isolated from human and non-human sources in the Plateau State of Nigeria. Food Microbiology, 26, 872–5.
- RAUF, M. O. (2012). Analysis of household demand for meat, in Southwest, Nigeria. *Global Journal of Science Frontier Research Agriculture and Biology*, 12(1 and 20).
- SAFIRULLAH, ANWAR, K., ABDUR-RAZIQ, SHAHID, M., RAZA, S., KHAN, N., and AMIN, Y. (2014). Epidemiological Study of Brucellosis in Equines of District Peshawar Khyber Pakhtunkhwa Pakistan. *International Journal of Current Microbiology and Applied Sciences*, 3(2), 795–800.
- SERRA, J. and VIÑAS, M. (2004). Laboratory diagnosis of brucellosis in a rural endemic area in northeastern Spain. International Microbiology : the Official Journal of the Spanish Society for Microbiology, 7(1), 53–8. Retrieved from

http://www.ncbi.nlm.nih.gov/pubmed/1 5179607

SHEY-NJILA, O., DAOUDA, E., NYA, E., ZOLI, P. A., WALRAVENS, K., GODFROID, J. and GEERTS, S. (2005). Serological Survey of Bovine Brucellosis in Cameroon. *Revue* D'Elevage et de Medecine Veterinaire Des Pays Tropicaux, 58(3), 139–143.

- SMITS, H. L., ABDOEL, T. H., SOLERA,
 J., CLAVIJO, E. and DIAZ, R. (2003).
 Immunochromatographic BrucellaSpecific Immunoglobulin M and G
 Lateral Flow Assays for Rapid
 Serodiagnosis of Human Brucellosis.
 Clinical and Diagnostic Laboratory
 Immunology, 10(6), 1141–1146.
- SWAI, E. S. and SCHOONMAN, L. (2010). The Use of Rose Bengal Plate Test to Asses Cattle Exposure to Brucella Traditional Infection in and Smallholder Dairy Production Systems Tanga Region of Tanzania. of Veterinary Medicine International, 2010. doi:10.4061/2010/837950
- TRAXLER, R. M., LEHMAN, M. W., BOSSERMAN, E. A., GUERRA, M. A. and SMITH, L. (2013). A Literature Review of Laboratory-Acquired Brucellosis. *Journal of Clinical Mcrobiology*, *10*, 135–148.
- UNGER, F., MUNSTERMANN, S., GOUMOU, A., APIA, C. N., KONTE, M., and MICHAELA, H. (2003). Risk associated with bovine brucellosis in selected study herds and market places in four countries of West Africa. Animal Health Working Paper 2. ITC (International Trypanotolerance Centre), Banjul, The Gambia. 37 pp.
- WHO. (2004). Laboratory Biosafety Manual. Third edition World Health Organization.