Resistance pattern of enterobacteriaceae isolates from urinary tract infections to selected quinolones in Yaoundé

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Abstract

Introduction: It is estimated that 150 million urinary tract infections (UTIs) occur yearly worldwide, resulting in more than 6 billion dollar in direct healthcare cost. The etiology of UTIs is predictable, with Escherichia coli, an Enterobacteriaceae being the principal pathogen. Quinolones are usually the drug of choice. In this study, we report the resistance pattern of Enterobacteriaceae isolates from UTIs to quinolones among in-patients and out-patients at the Yaoundé Reference Hospital in Cameroon. Methods: A cross-sectional descriptive study was carried out for a ten-month period. Consecutive clean-catch mid-stream urine samples were collected from 207 in and out-patients. Identification was done using the Api 20E, and susceptibility testing using the Kirby Bauer’s disc diffusion method and the MIC was done using the E-test. Results: Out of the 207 isolates, 58(28.0%) were found to be resistant to all the quinolones used in the study. The resistances observed by species were in the order: Enterobacter 4(30.8%); Klebsiella 19(29.7%); Escherichia 25 (29.4%); Proteus 2(11.8%); Serratia 4(25.0%). Quinolone resistance for Escherichia was 42.9% for In-Patients (IP) and 16.3% for Out-Patient (OP) (P-value = 0.006); Klebsiella 35.9% for IP and 20% for OP; Proteus 11.1% for IP and 12.5% for OP; Serratia 18.2% for IP and 40% for OP; Enterobacter 22.2 for IP and 50% for OP. Conclusion: High resistance rates to quinolones were observed not only for in-patients but also for out-patients with urinary tract enterobacterial infections. These findings demonstrate the importance of antibiotics susceptibility testing in improving quinolones prescription practices in Cameroon.
Introduction

Urinary tract infections (UTIs) are common infections. It is estimated that 150 million urinary tract infections occur yearly worldwide, resulting in more than 6 billion dollar in direct healthcare cost [1]. Complicated UTIs include those in patients with stones or obstructive uropathies and in patients with catheter-related infections. These infections are often associated with nosocomial, antibiotic-resistant gram-negative and gram-positive bacteria [2]. The etiology of UTIs is predictable, with Escherichia coli an Enterobacteriaceae being the principal pathogen [3].

However, E. coli and other uropathogens are becoming increasingly resistant to commonly prescribed antimicrobials, resulting in decreased effectiveness of some standard regimes. Quinolones are the drug of choice for these infections [4]. Ciprofloxacin, ofloxacin, lomefloxacin (Maxaquin), enoxacin (Penetrex), levofloxacin, and gatifloxacin have higher renal clearance and greater renal concentrations; they are optimal choices for the treatment of complicated UTIs [2].

Quinolones were first introduced into use in 1962 in the form of nalidixic acid, which is a completely synthetic agent that in clinical concentrations has bactericidal effects on most members of the Enterobacteriaceae [5,6]. Just as with other antimicrobials, the extensive use or administration of these compounds has led to the development of resistance by the bacteria [7-10].

Little is known about the antimicrobial resistance patterns of community-acquired organisms that circulate in developing countries where antimicrobials are available without prior consultation with a physician most often leading to auto medication [11].

Several surveillance studies from different parts of the world have shown that resistance to the fluoroquinolones among Enterobacteriaceae has increased dramatically worldwide especially during the past five years [12].

In this study, we report the resistance pattern in Enterobacteriaceae isolates from urinary tract infections to quinolones among in and out-patients at the Yaoundé Reference Hospital in Cameroon.

Methods

Specimen

A total of 207 urine specimens were collected consecutively within a period of ten months. Specimens were collected from in and out-patients at the Yaoundé General Hospital. The collection was done by trained medical personnel avoiding contamination. Clean-catch mid-stream urine samples were collected from consenting patients. The specimens were immediately transported to the laboratory after collection and processed. All contaminated urine specimens and all patients or children whose guardians refused to fill the consent form were excluded from the study.

Isolation and identification

The specimens were inoculated onto Nutrient agar plates using a calibrated loop designed to deliver a known volume of 5µl. Inoculated plates were then incubated aerobically at 37°C for 24 hours. After 24 hours, discrete colonies were picked up and gram stained. Further sub-culturing for gram negative bacilli was done on Eosin Methylene Blue agar to obtain a pure culture and biochemical tests were carried out using the Api 20E identification kit in accordance with the manufacturer’s manual ((BioMérieux SA, Lyon, France).

Antimicrobial susceptibility testing

This was done using the Kirby Bauer Disc diffusion method with reference to the Clinical Laboratory Standard Institute (CLSI) performance guideline for antimicrobial susceptibility testing [13]. Quality was assured by testing the E. coli quality control strain, ATCC 25922, in every batch. All zones of inhibition determined were within the ranges prescribed by the CLSI. Seven quinolones were used; nalidixic acid (NA) and pipemidic acid (PI) of the first generation; ciprofloxacin (CIP), norfloxacin (NOR), and ofloxacin (OFX) of the second generation; sparofloxacin (SPX) of the third generation; and moxifloxacin (MXF) of the fourth generation.

Minimum inhibitory concentration (MIC)

The minimum inhibitory concentration was determined for all the isolates that were found to be resistant in the disc diffusion method using the e-test. Ciprofloxacin was used. The method was controlled
by parallel testing using quality control reference strain (ATCC 25922) as recommended by the CLSI

**Ethical considerations**

This study received approval from the ethics committee of the Faculty of Medicine and Biomedical Sciences of the University of Yaoundé 1. All the patients signed the consent form before specimen collection. All the samples were assigned codes after collection.

**Results**

From the 207 urine specimens collected, 118 (57%) isolates were from in-patients and 89 (43%) from out-patients. The prevalence of Enterobacteriaceae in the isolates was in the order: Escherichia 85 (41.1%); Klebsiella 64 (30.9%); Proteus 17 (8.2%); Serratia 16 (7.7%); Enterobacter 13 (6.3%); others (Citrobacter, Kluvyvera, and Morganella) 12 (5.9%)

Out of the 207 isolates, 58 (28.0%) were found to be resistant to all the quinolones. The resistance observed in each species were in the order: Enterobacter 4 (30.8%); Klebsiella 19 (29.7%); Escherichia 25 (29.4%); Proteus 2 (11.8%); Serratia 4 (25.0%); others 4 (33.3%) Figure 1. The resistance of each quinolone was NA 74 (35.4%); PI 84 (40.6%); CIP 60 (29.0%); OFX 63 (30.4%); NOR 63 (30.4%); SPX 62 (30.0%) and MXF 61 (29.5%) (Shown in Figure 1) Table 1.

The prevalence of all the Enterobacteriaceae in the in-patients (IP) was 118/207 (57.0%) and in out-patients (OP) was 89/207 (43%) (See Table 1). The prevalence of Escherichia was 42/118 (35.6%) for IP and 43/89 (48.3%) for OP, for Klebsiella 39/118 (33.1%) for IP and 28/89 (28%) for OP, for Proteus 9/118 (7.6%) for IP and 8/89 (9%) for OP, for Serratia 11/118 (9.3%) for IP and 5/89 (5.6%) for OP, for Enterobacter 9/118 (7.6%) for IP and 4/89 (4.5%) for OPTable 2.

The resistance of all the Enterobacteriaceae in the in-patients (IP) was 41/118 (34.75%) and in out-patients (OP) was 18/89 (20.23%) as shown on Table 2. The resistance in Escherichia was 18/42 (42.9%) for IP and 7/43 (16.3%) for OP, for Klebsiella 14/39 (35.9%) for IP and 5/28 (17.9%) for OP, for Proteus 1/9 (11.1%) for IP and 1/8 (12.5%) for OP, for Serratia 2/11 (18.2%) for IP and 2/5 (40%) for OP, for Enterobacter 2/9 (22.2%) for IP and 1/4 (25%) for OP.

**Discussion**

In this study we describe the resistance pattern of Enterobacteriaceae isolates from urinary tract infections to a selected number of quinolones among in and out-patients. The overall resistance of the Enterobacteriaceae to these quinolones was high. In African countries with high infectious disease burden, formal and informal health systems depend heavily on broad spectrum orally-administrable antibacterial. This has caused an increase in the resistance of these antibacterial over the years and is also affecting even the newer antibacterial like the fluoroquinolones [14].

Quinolones are widely used for the treatment of serious *E. coli* Urinary tract infections (UTIs) and may also be used to treat other infections caused by other members of the Enterobacteriacae family. Hence, these high levels of quinolone resistance may lead to treatment failure which is of significant concern. These high rates of fluoroquinolone resistance observed could also limit treatment options especially oral treatment [15,16]. The statistically significance difference in resistance between the in and out-patients could be due to hospitalized patient being infected by nosocomial bacteria.

The strength of this study is the broad population of interest that is both the in and out-patients reflecting the circulating resistance. There are however some limitations. Our study had a relatively small sample size. We also carried out susceptibility testing on only a limited numbers of quinolones using only two first generations, three second generation, one third generation and one fourth generation. This might not be representative of the whole class of quinolones which includes several quinolones in each generation.

The overall resistance of 28% to all selected quinolone was slightly higher than what was found in other studies carried out in Ghana, Greece, the Netherlands and Brazil [16-19]. The resistant rate of the isolate from inpatients was higher than that from the outpatients. This is similar to what was found in other studies [17]. In our study we found that the *Escherichia* species were the most resistant
among the inpatient whereas other studies carried out in Greece [17] found Klebsiella to be the most resistant. However, among the out-patient Klebsiella was classified as the most resistant just as found in other studies. There was a statistically significant difference between the overall resistance in inpatients and out-patients just as also shown by another study [17].

Many factors have contributed to this high resistance rates; excessive antibiotic prescription is related to a higher prevalence of antibiotic resistant bacteria, misuse of antibiotics by health professional, unskilled practitioners and laypersons (antibiotics can be purchased without a prescription). Poor drug quality, unhygienic conditions and inadequate surveillance also account for the spread of resistant bacteria [20-23].

Another area to be exploited for future research is the genetic diversity and the characterization of these quinolone resistant Enterobacteriaceae strains. This will enable us determine the prevalence of plasmid-mediated resistance which is a common phenomenon among quinolone resistant strains. Further work may also identify the prevalence of expanded spectrum beta-lactamases among these quinolone resistant Enterobacteriaceae strains.

Conclusion

High resistance rates to quinolones were observed not only for in-patients but also for out-patients with urinary tract enterobacterial infections. These findings demonstrate the importance of antibiotics susceptibility testing in improving quinolones prescription practices in Cameroon.

Competing interests

The authors declare no competing interest.

Authors’ contributions

Emilia Enjema Lyonga conceived the study and designed it together with Michel Toukam and Sinata Koulla-Shiro. Emilia Enjema Lyonga, Michel Toukam, and Celine Nkenfou, conducted the laboratory aspect of the study with contribution from Marie-Claire Okomo Assoumou, Martha Tonge Mesembe, Agnes Bedie Eyoh, George Mondinde Ikomey. The general supervision was carried out by Sinata Koulla-Shiro. Emilia Enjema Lyonga drafted the article with contribution from Martha Tonge Mesembe and Valantine Ngum Ndze. All the authors reviewed the article. All the authors read and agreed to the final manuscript.

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References


<table>
<thead>
<tr>
<th>Table 1: The prevalence of the enterobacteriaceae isolates in hospitalized and community patients</th>
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<td>---------</td>
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</tr>
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</tr>
<tr>
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<td>Enterobacter</td>
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### Table 2: The resistance of the enterobacteriaceae to quinolones

<table>
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<th>Out-patients (n=89)</th>
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<tr>
<td>Serratia</td>
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<td>2</td>
</tr>
<tr>
<td>Enterobacter</td>
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<td>22.2%</td>
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<tr>
<td>Total</td>
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<td>34.75%</td>
<td>18</td>
</tr>
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</table>

**Figure 1:** The resistance rate of the enterobacteriaceae isolates to the selected quinolones

NA= Nalidixic acid; PI= Pipemidic acid; CIP= Ciprofloxacin; NOR= Norfloxacin; OFX= Ofloxacin; MXF= Moxifloxacin; SPX= Sparfloxacin