ISSN 1118 – 1931

INVESTIGATING THE FEASIBILITY OF UTILIZING *PENNISETUM PURPUREUM* LEAVES WASTE AS A SUSTAINABLE DYE: EXTRACTION, CHARACTERIZATION AND APPLICATION ON TEXTILE

231

Clark, P. D.^{1,2*}, Otutu, J.O.², Asiagwu, K. A.², Ndukwe, G. I.³ and Idibie, C. A.¹

¹Department of Chemical Sciences, Edwin Clark University, Kiagbodo, Nigeria
 ²Department of Chemistry, Delta State University, Abraka, Nigeria
 ³Department of Chemistry, Rivers State University, Port Harcourt, Nigeria
 *Corresponding author: poroclark81@gmail.com, clarkporo@edwinclarkuniversity.edu.ng

Received: 27-11-2023 *Accepted:* 06-12-2023

https://dx.doi.org/10.4314/sa.v22i3.21 This is an Open Access article distributed under the terms of the Creative Commons Licenses [CC BY-NC-ND 4.0] http://creativecommons.org/licenses/by-nc-nd/4.0. Journal Homepage: http://www.scientia-african.uniportjournal.info Publisher: *Faculty of Science, University of Port Harcourt.*

ABSTRACT

This study investigated the potential of Pennisetum purpureum (Elephant grass) leaves waste as a source of natural dyes. The objective was to extract, characterize and apply the natural dyes on textile fabrics. Elephant grass was chosen due to its abundant availability as agricultural waste, making it an environmentally sustainable alternative to synthetic dyes. The extraction process involved maceration, followed by filtration to obtain the dye extract. The dye components were isolated using vacuum liquid chromatography and then characterized using analytical tools such as UV-Visible Spectrophotometry, High-Performance Liquid Chromatography (HPLC) and Fourier-Transform Infrared Spectroscopy (FTIR) to identify the presence of specific compounds responsible for the dyeing potential. The perspiration fastness, rubbing fastness, light fastness and wash fastness properties were assessed to evaluate the durability and suitability of the natural dyes. The UV-Visible spectrum, HPLC and FTIR analysis confirmed the presence of chromophores such as conjugated systems, and provided information about chemical components namely rutin, quercetin, senecionine, hyoscyamine and tannic acid present in the dye, as well as the types of bond present in the molecules including C-H, O-H, C=O and C-O groups, which are characteristics of natural dyes. The dyed textile fabrics demonstrated somewhat fair to good with a rating of 3-7 for perspiration fastness, light fastness, rubbing fastness and wash fastness, indicating the potential of the natural dye for practical applications. The findings can inspire further research and development in utilizing agricultural waste for natural dye production, promoting sustainable practices in textile manufacturing.

Keywords: Natural dye, Characterization, Maceration, Isolation, *Pennisetum purpureum*, Leaves, Waste, Textile

INTRODUCTION

In recent years, there has been growing interest in finding sustainable and ecofriendly alternatives in various industries, including the textile industry. The textile industry is notorious for its heavy reliance on synthetic dyes, which not only contribute to environmental pollution but also have detrimental effects on human health (Maleki and Barani, 2019). In response to these concerns, researchers have focused their efforts on exploring natural dye sources that are environmentally sustainable and offer viable alternatives to synthetic dyes (Maleki and Barani, 2019).

One such potential natural dye source is Pennisetum purpureum (Elephant grass) leaves waste. Pennisetum purpureum (Schumach), commonly referred to as elephant grass, is a tall and fast-growing perennial grass that belongs to the Poaceae family (Jack et al., 2020). It is widely cultivated in tropical and subtropical regions as a weed or forage crop (Obi et al., 2008). Due to its extensive cultivation, the leaves of Pennisetum purpureum (elephant grass) are abundantly available as agricultural waste. This waste material presents an opportunity to explore its potential for extracting natural dyes, thereby offering a sustainable and economically viable solution.

This study explored the potential of Pennisetum purpureum leaves (Elephant grass) waste as a source of natural dye and its application in dyeing textile fabrics. The colouring matters identified in natural dyes include several classes of compounds such as tannins, alkaloids, anthraquinones, naphthoquinones and carotenoids. Due to the relatively low exhaustion of natural dyes, mordants are usually employed to improve the colour strength and fastness, and to obtain multiple shades (UI-Islam et al., 2018; Adeel et al., 2018; Barani, 2018). By investigating characterization the extraction, and application of natural dyes derived from Pennisetum purpureum leaves waste, this research endeavours to contribute to the development of sustainable and eco-friendly dyeing practices.

MATERIALS AND METHODS

Chemicals and Reagents

Laboratory grade ferrous sulphate (FeSO₄) was used as mordant while a diluted solution

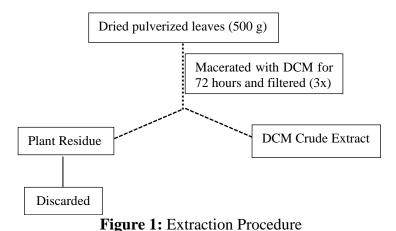
(2 g/L) of sodium carbonate (Na₂CO₃) was used to adjust the pH of the dye solution to 7. Reference detergent A soap (5 g/L) was used for the wash fastness test. *n*-Hexane, ethyl acetate, ethanol, hydrochloric acid (37% fuming HCl), acetonitrile, glacial acetic acid, Wagner's reagent (potassium iodide and iodine crystal) and sulfuric acid, all of which were of analytical grade and obtained from Merck (Darmstadt, Germany).

Plant Material

Pennisetum purpureum leaves were collected from Umuola-Egbelu, Abia State. The leaves were then identified and given a voucher number (No. 0615) from the Department of Forestry and Wood Technology at the Federal University of Technology, Akure. The leaves were washed with distilled water without squeezing to remove debris and dust particles, and then sun-dried for 3 weeks to retain the vibrant colours as the sun's energy aids in the natural preserving pigment. Once completely dried, the plant material was pulverized into a powder, using a manual blender (Porkert Manual Grinder No. 32) and stored at room temperature until further use.

Extraction of Crude Dyes

The dried powdered sample (500 g) of purpureum leaves were Pennisetum macerated with 2.0 L of dichloromethane (DCM) in an aspiratory bottle. This mixture was left at room temperature for 72 hours and stirred regularly. Afterward, the resulting extract was filtered into a conical flask using a funnel and filter paper to obtain the dichloromethane extract. The residue left was again subjected to a second extraction with fresh DCM according to the procedure described above to obtain the second extract of DCM, this procedure was repeated three (3) times in total to ensure thorough extraction of the leaves components (Figure 1).



Isolation and Purification of Crude Dyes

Vacuum liquid chromatography (VLC) was performed using the method described by Paranagama (2016) and Ndukwe *et al.* (2020) with slight modifications (Figure 2). To ensure optimal packing density, the VLC column was dry packed with thin layer chromatography (TLC) grade silica under vacuum. Subsequently, dichloromethane crude extract of *Pennisetum purpureum leaves* was prepared, along with silica gel mesh, and loaded onto the column. The elution process was carried out bv sequentially using 300 ml of suitable solvent mixtures (mobile phase), beginning with a low polarity solvent (100 % *n*-hexane); subsequently, the polarity was gradually increased by adjusting the solvent ratio (nhexane-DCM in ratios of 3:1, 2:2, 1:3), 100% DCM, DCM-ethyl acetate (3:1, 2:2, 1:3), 100 % ethyl acetate, ethyl acetate-EtOH (3:1, 2:2, 1:3) and 100 % EtOH) between each fraction collected. The column was pulled dry after each mobile phase to ensure proper separation.

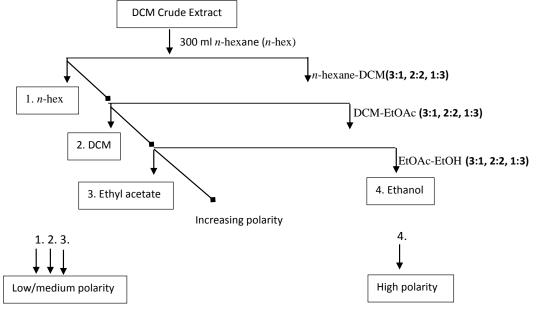


Figure 2: Framework of VLC model

Phytochemical Screening Tests

Phytochemical examination was conducted on the dye extract of *P. purpureum* leaves using standard procedures (Jack *et al.*, 2020; Clark and Omo-Udoyo, 2021) to detect the following bioactive compounds: alkaloids, flavonoids, glycosides, terpenoids, tannins and steroids.

Characterization of the Isolated Dye

Properties of a spectrum are examined and understood through a scientific process known as spectra characterization. The chemical composition and properties of dye from *Pennisetum purpureum* leaves were analyzed and interpreted by utilizing HPLC, FTIR and UV-vis scanning techniques. This process yields valuable information essential for evaluating their potential as a sustainable dye.

UV-Visible Analysis

A UV-VIS-NIR scanning spectrophotometer UV-3101PC (SHIMADZU) was used for all spectrophotometric measurements. All measurements were carried out using quartz cells 10-mm at room temperature (25 ± 2 °C) and changes in their absorption (400-800 nm) were noted.

Fourier Transform-Infrared Analysis

The software used for FTIR data collection was the Infrared Data Management (IRDM) system. The infrared spectrum was recorded at room temperature with a PerkinElmer Fourier Transform Infrared Spectrometer, Model spectra 100 series (Perkin-Elmer Corporation, Norwalk, CT, USA), equipped with a deuterated triglycine sulfate (DTGS) detector and controlled by a Perkin-Elmer PC. The instruments were maintained in constant humidity to minimize water vapor interference.

Drops from each standard were placed on the attenuated total reflection element and scanned. After each scan, the ATR diamond was rinsed three times with acetone and dried with soft tissue before adding the sample; Calibration spectrum was obtained from 64 scans at a resolution of 2 cm⁻¹ with strong apodization through 3500-1000 cm⁻¹ frequency region. The spectrum was rationed against the background air spectrum. All the scans were done in triplicate with the

spectrum recorded as absorbance and stored on a disk.

High Performance Liquid Chromatography Analysis

The HPLC analysis was carried out using AGILENT 1260 infinity HPLC system with a detector photo diode array (Agilent Technologies, Palo Alto, CA). The chromatographic separations were carried out using an XbridgeTM Shield RP₁₈ column (4.6 mm I.D. \times 150 mm, 3.5µm) (Waters, Milford with column oven temperature USA). maintained at 20 °C (Park et al., 2016). The mobile phase consisted of 0.1% acetic acid (Solvent A) and 100 % acetonitrile (Solvent B). The mobile phase flow rate was 1.0 ml/min with gradient elution. The percentage composition of Solvent B was maintained at 20 % for 3 min, gradually increased to 38% for 24 min. further increased to 90 % for 1 min and maintained at 90 % for 5 min, which was followed by equilibration to the initial composition for 6 min. the injection volume was 10 µL and UV absorbance was monitored at 365 nm.

Mordanting

In this experiment, the process of mordanting was conducted before dyeing, referred to as pre-mordanting. The aim of pre-mordanting was to enhance the adsorption of the dye and ensure a strong bond between the dye and the fabric. The commonly used mordant, such as iron (ferrous sulfate) was selected. Initially, the cotton fabric was immersed in warm water (approximately 46 °C) for 30 minutes to relax the fabrics, which would make the fabric more receptive to mordanting and dveing. Subsequently, the specific mordanting procedure was carried out based on the information found in the literature (Feng et al., 2007).

Dyeing Procedure

The dyeing procedures were performed in accordance with the general dyeing method (Baaka *et al.*, 2015). A fabric-to-dye ratio of

1:10 was chosen based on the weight of the fresh natural dyes extracted and the fabrics used in the experiment. The fabric was immersed in a dyebath composed of 0.25% aqueous solution of the dye. The dye liquor ratio of 1:40 was kept constant for all samples, and the pH value of the dyebath was optimized depending on the type of raw material. For P. pupureum, the pH values were adjusted by adding drops of sodium hydroxide or hydrochloric acid to achieve pH levels of 9-10 and 3-4, respectively. The temperature of the dyebath was gradually increased (about 1 °C) until it reached 100 °C and was kept at this temperature for about 60 minutes. Afterwards, the dyebath was allowed to cool to around 60 °C. The dyed fabric was then squeezed, thoroughly rinsed with water and air-dried.

Determination of Wash Fastness

The dyed specimens of wool and nylon fabrics with a dimension of 5 cm \times 4 cm were placed between two pieces of undyed white fabrics of the same dimension. Three pieces were stitched together around the edges to create a composite specimen. The composite specimen was agitated with ten steel balls in a 100 ml beaker, containing a solution made-up of 5 g/L soap and 2 g/L soda ash with a liquor ratio of 1:50 as stipulated by ISO 3 standards. The washing process was carried out at 60 \pm 2 °C for a duration of 30 minutes in a laundero-meter. The composite specimen was then rinsed, separated and dried. The change in colour of the test samples and the staining of the adjacent undyed white fabrics were evaluated using the grey scale. with references to the ISO 9001 2000 group.

Determination of the Light Fastness

Strips of the fabrics and the blue wool standards were cut and mounted on cardboard paper. Half portions of the specimens were covered to obstruct the light source from getting to that portion. The specimens were exposed to natural daylight in a south-facing direction at an angle of 45 °C, sloping from the horizontal, for a duration of 72 hours.

After 72 hours, the specimens were removed and the extent of their fading was assessed by comparing them to the blue wool standards.

Determination of Fastness to Dry and Wet Rubbing

The dyed samples' dry and wet rubbing fastness was tested using a Crock meter in accordance with ISO 105-X 12:2001 standards. The specimen was placed in the Crock meter and a piece of standard white cloth (starch free 96.100 cotton fabric of a long type) was used to rub against the coloured specimen. This rubbing process was carried out under controlled conditions of pressure and speed. For both the dry and wet tests, the rubbing fingers were covered with white cloth and moved back and forth for a total of 20 rubbing strokes. The colour transferred onto the white cloth was compared with a Grayscale for alteration of colour, consisting of grades 1-8.

Fastness to Perspiration Test

The fastness to perspiration test evaluates the ability of textile fabrics to resist colour fading or running when exposed to perspiration. This test was conducted using acidic and alkaline solutions; the acidic solution consists of sodium chloride (NaCl, 5 g/L), disodium hydrogen orthophosphate dehydrate (Na₂HPO₄ 2.5 g/L) and histidine monohydrochloride monohydrate. The pH of the solution was adjusted to 5.5 while the alkaline solution consists of C₆H₉O₂N₃.HCl.H₂O (0.5 g/l) and is adjusted to pH 8 using 0.1 N sodium hydroxide (NaOH). The liquor ratio for the test was 20:1.

RESULTS AND DISCUSSION

Phytochemicals of the Isolated Dye

The crude dye extracted from *P. pupureum* leaves using the maceration method were analyzed to identify specific compounds. Previous research conducted by Hayouni *et al.* (2007) suggested that the maceration method may be more effective for extracting secondary metabolites. The crude dichloromethane extract was separated into thirteen (13) fractions using vacuum liquid chromatography. Remarkably, one of the fractions exhibited the presence of metabolites such as tannins, flavonoids, steroids and alkaloids. However, cardiac glycoside and terpenoid were not observed (as indicated in Table 1). Interestingly, these findings align with a prior investigation by Adeove (2021), who also reported the absence of these compounds (cardiac glycoside and terpenoid) in the leaves of the same plant species. Therefore, the active fraction containing tannins, flavonoids and alkaloids are of relevance to the dye industry because of their potential applications. Tannins are known for their astringent properties and ability to form complexes with metal ions, making them useful in dyeing processes (Janani and Winifred, 2013). Flavonoids on the other hand are recognized for their therapeutic potential and possesses antimicrobial and antioxidant properties (Ndukwe et al., 2020). While their direct application in the dye industry may be limited, their antioxidant properties can be valuable for the preservation of natural dyes and pigments. Some alkaloids have been used as natural dyes in the past, but their application in modern dyeing processes is limited. However, they may still have niche applications in natural dyeing processes (Rather et al., 2017).

Table 1: Phytochemical groups present in the Isolated Dye

Phytochemical group	VLC Fraction 13 (Isolated Dye)		
Alkaloids	+		
Steroids	+		
Tannins	+		
Flavonoids	+		
Terpenoids	-		
Cardiac Glycosides	-		

KEY: + Present, - Absent

Chemical Characteristics and Composition of the Isolated Dye

The UV-visible wavelength of *P. purpureum* leaves dye is shown in the first column of Table 2. The wavelength at 401 nm is attributed to a $\pi \rightarrow \pi^*$ transition resulting from the presence of multiple conjugated bonds and Figure 3 represented a plot of the absorbance versus wavelength. The absorption peak observed in this study aligns with the research of Budidha et al. (2020). In their findings, they have associated the absorption peak of 401 nm with the 8th harmonic of O-H stretching vibrational bands. This information is valuable as it provides additional insight into the molecular structure and other pertinent details of the dye. The colour associated with this absorption peak falls within the violet region of the light spectrum, and its complementary colour was yellow.

FT-IR spectroscopy has been widely used for the analysis of natural dyes (Amir-Al Zumahi et al., 2020). The dye extracted using dichloromethane exhibits distinct bands within different segments of a spectrum: 3600-3200 cm⁻¹, 3100-2800 cm⁻¹, 1650-1600 cm⁻¹, 1480-1300 cm⁻¹ and 1300-900 cm⁻¹ as shown in Table 2. These bands signify the stretching vibrations of various functional groups, such as O-H, C-H, C-C, C=C, C=O, aromatics. and nitrile. Several studies conducted by different researchers (Ezeokonkwo et al., 2018 and Sofyan et al., 2018) have extensively discussed and identified the functional groups corresponding to the different specific segments of the FT-IR spectrum.

The spectrum for the *P. purpureum* leaf dye as represented in Figure 4 provides valuable insights into its chemical composition. This data was obtained within the frequency range of 3500 - 1000 cm⁻¹, enabling the analysis of the dye molecular structures and functional groups. The spectrum revealed several peaks at specific wavenumbers, including 3257.7 cm⁻¹, 2094.8 cm⁻¹, 1636.3 cm⁻¹ and 1390.3 cm⁻¹. These peaks indicated the presence of compounds responsible for the colouring properties of the dye. Notably, within the range of 1700 to 1100 cm⁻¹, the absorption bands exhibited similarities to the characteristic associated with values flavonoid-like dye compounds, as reported by and Parac-Osterman (2017). The Jemo characteristics of *P. purpureum* leaf dye can be observed through stretching vibrations at 3257.7 cm⁻¹, which indicates the presence of O-H in phenol, as suggested by Al-Sharairi et al. (2020). According to Kassim et al. (2011), the shape of the OH-stretching band provides preliminary information on the occurrence of a polymerization process. Condensed tannins, which have varying degrees of polymerization, display a broad range of peaks ranging from 3,700 to 3,000 cm⁻¹.

The overview of flavonoids detected in the isolated dye (Table 3 and Figure 5) revealed that Rutin had the highest percentage composition and affinity for the stationary phase, with values of 51.6% and 10.508 minutes, respectively. Isoquercetin followed with a value of 38. 8% and 8.536 minutes. On

the other hand, quercetin had the lowest retention time and percentage composition with a value of 7.095 and 9.63%. These findings align with the research conducted by Engida et al. (2013) who also identified rutin and quercetin as the main flavonoid colourants in their study. However, Table 3 and Figure 6 illustrate the presence of tannins in P. purpureum leaf dye. Tannic acid was the sole compound detected in the isolated dye with concentration, retention time and percentage composition of 16.5350 µg/ml 3.1099 minutes and 100%. The presence of tannic acid, a recognized natural mordant, is crucial for enhancing the colour fastness of dyes (Janani and Winifred, 2013). In addition, Table 3 and Figure 7 depict the alkaloids identified in the isolated dye; atropine (RT 3.757 minutes) was observed to be the most prominent alkaloid with percentage a composition of 64.6%, followed by hyoscyamine (RT 4.438 minutes) as the second most prevalent alkaloid with a value of 21.5%. Conversely, senecionine (RT 7.299 minutes) exhibited a comparatively low percentage composition (3.9%). However, the striking observation was that comparisons with the existing literature on P. purpureum leaves were not currently feasible since this study represents the first attempt to identify these compounds using HPLC.

Table 2 . Spectra data of the isolated dye		
UV-visible (nm)	Infrared (cm ⁻¹)	
$401.00 \ (\pi \to \pi^*)$	3257 (O-H of phenol)	
	2095 (C-H stretch in aromatic)	
	1610-1440 (C=C stretch of aromatics)	
	260-1000 (C-O stretch in phenols)	

Table 2: Spectra data of the isolated dye

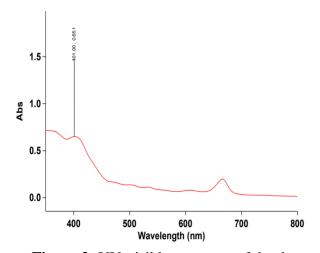
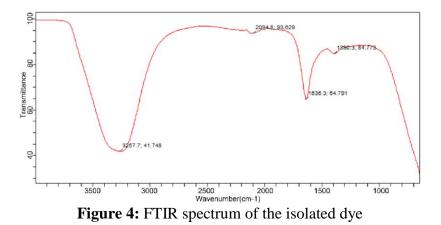


Figure 3: UV-visible spectrum of the dye



1 a D1	Table 5. Constituents of the dye isolated from 7. purpureum leaves					
Compound	Phytochemical Group	Concentration (µg/ml)	Percentage composition per Group			
Isoquercetin (1)	Flavonoids	44.096	38.8			
Rutin (2)	Flavonoids	58.641	51.6			
Quercetin acid (3)	Flavonoids	10.953	9.63			
Tannic acid (4)	Tannins	16.5350	100			
Senecionine (5)	Alkaloids	19. 147	13.9			
Hyoscyamine (6)	Alkaloids	29.631	21.5			
Atropine (7)	Alkaloids	89.095	64.6			

 Table 3: Constituents of the dye Isolated from P. purpureum leaves

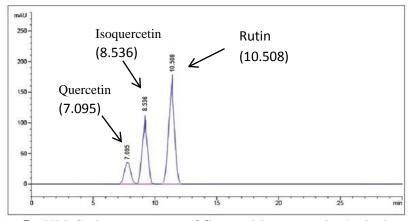


Figure 5: HPLC chromatogram of flavonoids present in the isolated dye

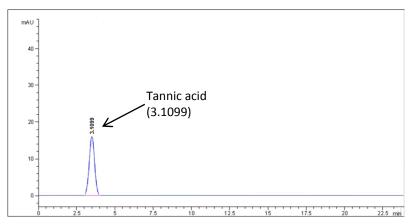


Figure 6: HPLC chromatogram of tannins present in the isolated dye

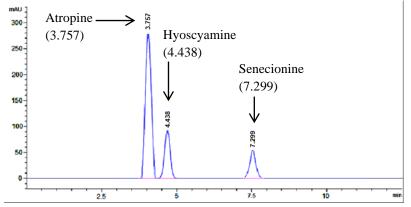
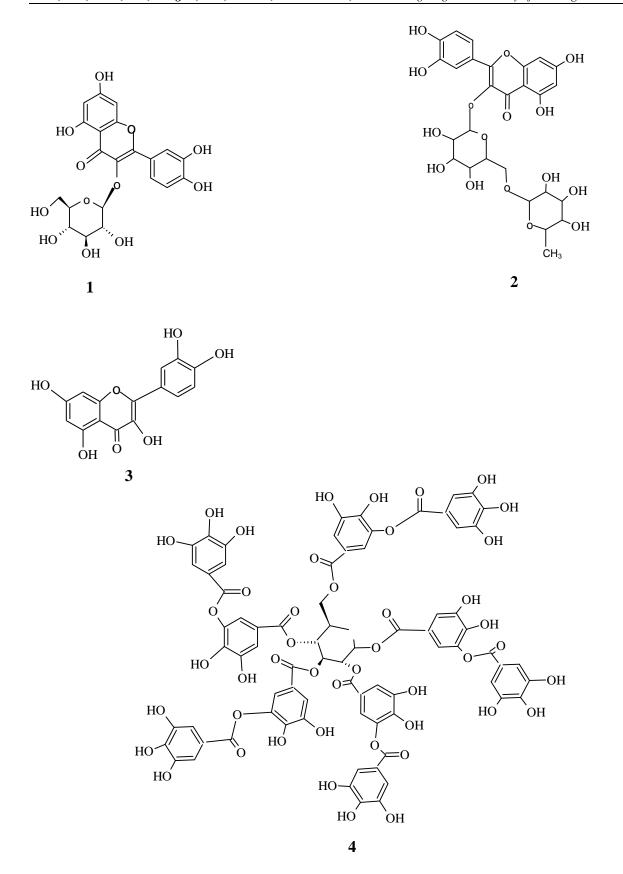
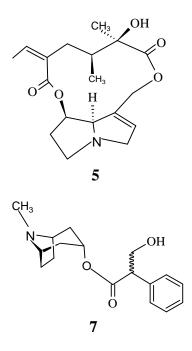
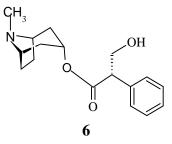


Figure 7: HPLC chromatogram of alkaloids present in the isolated dye



Clark, P.D., Otutu, J.O., Asiagwu, K.A., Ndukwe, G.I. and Idibie, C.A.: Investigating the Feasibility of Utilizing Pennisetum...





Colour Fastness Properties

This study reports on the fastness properties, including light fastness, perspiration fastness, wash fastness and rubbing fastness, of fabrics dyed with *P. purpureum* leaf dye.

Colour Fastness to Washing

Wash fastness of dye was influenced by the rate of diffusion of dye and the state of dye inside the fabric (Kanchana et al., 2013). The results presented in Table 4, Figures 8 and 9 demonstrate that the fabrics dyed with P. purpureum leaves dye exhibited somewhat fair to good colour fastness, ranging from DF 2% to DF 8%. However, when examining the specific cases of wool and nylon fabrics dyed with DF 4%, they received the lowest rating of 2-3/3. This rating indicates that there was a minimal colour change in the fabrics after Despite the deep shade of the washing. fabrics dyed with DF 4%, it did not receive a high rating on the scale. This could be attributed to the presence of smaller dyemetal complexes and weaker interaction between the dye and the fabric, making it easier for the dye to wash out or diffuse from the fabrics and ultimately leading to its poor rating on the scale.

Colour Fastness to Light

A Colour fastness test was conducted on the dyed fabrics to evaluate their resistance to daylight exposure. Overall, the samples exhibited good to excellent fastness to light, with ratings ranging from 5-7. However, DF 2 % nylon received a lower rating of 3 on the blue wool scale after 48 hours exposure to simulated sunlight (Table 5, Figures 8 and 9). This indicates that nylon and wool fabrics can be effectively dyed using P. purpureum leaves dye. The presence of flavonoids in P. purpureum leaves, as mentioned by Adeoye (2021), may be responsible for this successful Flavonoids are phenolic dyeing process. compounds that can form hydrogen bonds with carboxyl groups present in protein fibers such as wool (Agarwal and Patel, 2002). Additionally, Burkinshaw and Kumar (2009) suggested that the characteristics of mordants like ferrous sulfate play a more significant role in determining the fastness properties of natural dyes than the dyes themselves. The obtained dye and their properties are the result of the formation of wool mordant dye interactions (Figure 10).

Clark, P.D., Otutu, J.O., Asiagwu, K.A., Ndukwe, G.I. and Idibie, C.A.: Investigating the Feasibility of Utilizing Pennisetum...



Figure 8: Wool fabrics dyed with the isolated dye and tested for colour fastness



Figure 9: Nylon fabrics dyed with the isolated dye and tested for colour fastness

Table 4: Colour fastness to wash

Sample Code	Colour C	Change
	Wool	Nylon
DF 2 %	3	4
DF 4 %	2-3	3
DF 6 %	3	4
DF 8 %	4-5	3-4

Key: DF-Dye concentration, 1-very poor, 2-poor, 3-fair, 4-moderate, 5-good, 6-very good, 7-excellent, 8-outstanding

Table 5:	Colour fastness to light
1	a 1 a1

0

Sample	Colour C	Change
code	Wool	Nylon
DF 2 %	6	3
DF 4 %	5	5
DF 6 %	7	6
DF 8 %	5	6
II DED		a a a i

Key: DF-Dye concentration, 1-very poor, 2-poor, 3-fair, 4-moderate, 5-good, 6-very good, 7-excellent, 8-outstanding

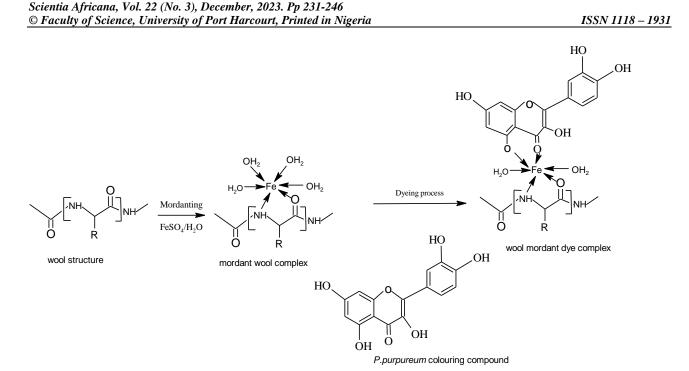


Figure 10: Schematic representation of wool-mordant-dye interaction

Colour Fastness to Perspiration

The perspiration fastness of nylon and wool fabrics dyed with the isolated dye was evaluated under acidic and alkali conditions, as shown in Table 6 and Figures 8 and 9. The fabrics (mordanted with ferrous sulfate) had moderate fastness to alkali perspiration at 6% and 8% dye concentrations for both nylon and wool. The fabrics also showed fair to moderate fastness to acidic perspiration at 2% and 4% dye concentrations for both nylon and wool, with very light staining on adjacent fabrics. These results indicate that the alkali extract of P. purpureum leaves dye can produce fabrics that are resistant to perspiration in different environments.

Colour Fastness to Rubbing

The rubbing fastness results for nylon and wool dyed with isolated dye from P. purpureum leaves extract at concentrations ranging from DF 2% to DF 8% revealed that the use of ferrous sulfate as a mordant resulted in a rating of 5 for dry rubbing and a rating of 4 for wet rubbing at DF 8 % concentration on nylon, which was better than its performance on wool (Table 7 and Figures 8 and 9). Comparatively, the results indicated that the dry rubbing performance for both wool and nylon was superior to their wet rubbing performance.

Table 6: Colour fastness to perspiration					
Sample code	Colour Change				
	Wool		Nylon		
	Acid	Alkaline	Acid	Alkaline	
DF 2 %	3	2-3	3	3	
DF 4 %	4	3-4	4	3-4	
DF 6 %	2-3	4	3-4	4	
DF 8 %	4	4	3	4	

Key: DF-Dye concentration, 1-very poor, 2-poor, 3-fair, 4-moderate, 5-good, 6-very good, 7-excellent, 8-outstanding

Clark, P.D., Otutu, J.O., Asiagwu, K.A., Ndukwe, G.I. and Idibie,	C.A.: Investigating the Feasibility of Utilizing Pennisetum
---	---

Table 7: Colour fastness to rubbing					
Sample Code	Wool		Nylon		
	Dry rubbing Wet rubbing		Dry rubbing	Wet rubbing	
DF 2 %	4	3	4	2-3	
DF 4 %	3	3	3-4	4	
DF 6 %	3	4	4	3-4	
DF 8 %	4	3-4	5	4	

Key: DF-Dye concentration, 1-very poor, 2-poor, 3-fair, 4-moderate, 5-good, 6-very good, 7-excellent, 8-outstanding

CONCLUSION

Extraction of natural dyes from Pennisetum purpureum (Elephant grass) leaves waste and characterization of the resulting dyes were effectively achieved. The application of this dye on textile fabrics, specifically nylon and promising wool. demonstrated results especially in terms of colour fastness to light. The fabric samples treated with the dye showed moderate to fair levels of fastness to perspiration in both alkali and acidic conditions. This study has shown that Pennisetum purpureum leaves waste can be used as a green alternative for textile dyeing.

Conflict of Interest

The authors declare that they have no conflict of interest regarding the publication of this manuscript.

REFERENCES

- Adeel, S., Hussaan, F., Rehman, N., Habib, M., Salman, S., Naz, N. and Amin, N. A. (2018). Microwave-assisted sustainable dyeing of wool fabric using cochinealbased carminic acid as natural colourant. *Journal of Natural Fibers*, 5:1-9.
- Adeoye, S. T. (2021). Proximate and phytochemical analysis of Azadirachta indica and Pennisetum purpureum leaves. *Mountain Top University*.
- Agarwal, B. J. and Patel, B. H. (2002). Studies on dyeing of wool with a natural dye using padding techniques. *Man Made Textiles in India, 45*(6):237-341.
- Al-Sharairi, N., Sandu, I. C. A. and Sandu, I. (2020). Recognition of natural silk fibers, dyes and metal threads of historical Romanian textile fragments using the multi-analytical techniques approach.

Textile Research Journal, 90:15-16.

- Amir-Al Zumahi, S. M., Arobi, N., Taha, H., Hossain, M., Kabir, H., Mati, R., Bashar, M. S., Ahmed, F., Hossain, M. and Rahman, M. M. (2020). Extraction, optical properties and aging studies of natural pigments of various flower plants. *Cell Press*, 2-13.
- Baaka, N., Ben, M. T., Hadder, W., Hammami, S. and Mhenni, M. F. (2015). Extraction of natural dye from waste wine industry: Optimization survey based on central composite design model. Fibers Polymers, 16(1):38-45.
- Barani, H. (2018). Modification of bentonite with different surfactants and substitute as a mordant in wool natural dyeing. *Chiang Mai Journal of Science*, 45:492-504.
- Budidha, K., Mamouei, M., Baishya, N., Qassem, M., Vadgama, P. and Kyriacou, P. A. (2020). Identification and quantitative determination of lactate using optical spectrosocopy-towards a non invasive tool for early recognition of sepsis. *National Center for Biotechnology Information*.
- Burkinshaw, S. M. and Kumar, N. (2009). The mordant dyeing of wool using tannic acid and FeSO₄, part 1: Initial findings. *Dyes and Pigments*, 53-60.
- Clark, P. D. and Omo-udoyo, E. (2020). Comparative assessment on antioxidant and phytochemical of *Trichilia monadelpha* (Thonn) J. J. De Wilde (Meliaceae) plant extracts. *Chemical Science International Journal*, *30*(10):24-33.
- Engida, A. M., Kasim, N. S., Tsigie, Y. A., Ismadji, S., Huynh, L. H., and Ju, Y. (2013). Extraction, identification and

quantitative HPLC analysis of flavonoids from Sarang semut (Myrmecodia pendan). Industrial Crops Production, 41:392-396.

- Ezeokonkwo, M. A., Okafor, S. N. and Godwin-Nwakwasi, E. U. (2018). Preliminary characterization of some natural dyes. *African Journal of Pure and Applied Chemistry*, 55-61.
- Feng, X. X., Zhang, L. L., Chen, J. Y. and Zhang, J. C. (2007). New insights into solar UV-protective properties of natural dye. *Journal of Cleaner Production*, 15:366-372.
- Hayouni, E. A., Abedrabba, M., Bouix, M. and Hamdi, M. (2007). The effects of solvents and extraction method on the phenolic contents and biological activities *in vitro* of *Tunisian Quercus coccifera* L and *Juniperus pheonica* L. fruits extracts. *Food Chemistry*, 105:1126-1134.
- Jack, I. R., Clark, P. D. and Ndukwe, G. I. (2020). Evaluation of phytochemical, antimicrobial and antioxidant capacities of *Pennisetum purpureum* (Schumach) extracts. *Chemical Science International Journal*, 29(4):1-14.
- Janani, L. and Winifred, D. (2013). Suitability of dyes from mulberry and coffee leaves on silk fabrics using ecofriendly mordants. *Interntional Journal of Scientific and Research Publications*, 3(11):1-4.
- Jemo, D. and Parac-Osterman, D. (2017). Identification of natural dyes on 18th century liturgical textiles from Dubrovnik. *Fibres and Textile in Eastern Europe*, 25:113-120.
- Kanchana, R., Fernandes, A., Bhat, B., Budkule, S., Dessai, S. and Mohan, R. (2013). Dyeing of textiles with natural dyes- an eco-friendly approach. *International Journal of ChemTech Research*, 5(5): 2103-2109.
- Kassim, M. J., Hussin, M. H., Achmad, A., Dahon, N. H., Suan, T. K. and Hamdan, H. S. (2011). Determination of total phenol, condensed tannin and flavonoid contents and antioxidant activity of

Uncaria gambir extracts. *Indonesian Journal of Pharmacy*, 22(1): 50-59.

- Maleki, H. and Barani, H. (2019). Extraction and antibacterial activity of Pulicaria gnaphalodes as a natural colouarant: Characterization and application on wool fibers. *Progress in Color Colorants and Coatings*, 12(3):145-154.
- Ndukwe, G. I., Clark, P. D. and Jack, I. R. (2020). In *vitro* antioxidant and antimicrobial potentials of three extracts of *Amaranthus hybridus* L. leaf and their phytochemicals. *European Chemical Bulletin*, 9(7):164-173.
- Ndukwe, G. I., Oluah, A. and Fekarurhobo, G. K. (2020). Isolation of an isoflavonoid and a terpenoid from the heartwood of Baphia nitida Lodd. (camwood). *Ovidius University Annals* of Chemistry, 31(1): 5-8.
- Obi, O. O., Omole, A. J., Ajasin, F. O. and Tewe, O. O. (2008). Nutritive potential of forages fed to growing grass-cutter (Tryonomys Swinderianns). *Livestock Research for Rural Development*, 5:284-301.
- Paranagama, P. A. (2016). Vacuum liquid chromatography and gel permeation chromatography in natural product research. *National Workshop on Seperation Technique in Natural Product Research*, 95-98.
- Park, J. S., Kim, I. S., Rehman, S. U., Na, C. and Yoo, H. H. (2016). HPLC determination of bioactive flavonoids in Hovenia dulcis fruit extracts. *Journal of Chromatographic Science*, 2:130-135.
- Rather, L. J., Shahid, U., Akhter, S., Hassan, Q.
 P. and Mohammad, F. (2017). Chemistry of plant dyes: Applications and environmental implications of dyeing processes. *Current Environmental Engineering*, 4(1): 2212-7178.
- Sofyan, N., Ridhova, A., Pramono, K. R. O., Yuwono, A. H. and Udhiarto, A. (2018). Visible light absorption and photosensitizing characteristics of natural dye extracted from Mangosteen pericarps using different solvents. *International*

Clark, P.D., Otutu, J.O., Asiagwu, K.A., Ndukwe, G.I. and Idibie, C.A.: Investigating the Feasibility of Utilizing Pennisetum...

Journal on		on	Ad	vanced	Science	
Engi	neer	ing Infor	mati	on Techno	logy.	
Ialam	C	Dathar	NЛ	Chabbin	ΝЛ	NI

Ul-Islam, S., Rather, M., Shabbir, M. N., Bukhari, M. A. and Khan, F. M. (2018). First application of mix metallic salt mordant combinations to develop newer shades on wool with Bixa orellana natural dye using reflectance spectroscopy. *Journal of Natural Fibers*, 15: 363-372.