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# Abstract

The growth performance and the carcass and physical and chemical characteristics of the meat of 24 Landrace X Large White pigs were compared when reared under a free-range or a conventional housing system. The free-range pigs had lower feed intakes and slower growth rates than the conventionally housed pigs. The free-range pigs also had a lower  $P_2$  fat depth and therefore yielded a carcass with a higher percentage lean meat. Housing systems had no effect on the weight distribution of the commercial cuts (as a percentage of cold carcass weight). The meat from the free-range pigs was slightly more reddish in colour, but apart from that housing systems had no effect on the water-holding capacity (WHC) of the meat, its initial pH (pH<sub>45</sub>) or its final pH (pH<sub>24</sub>). The meat from the free-range pigs had the same shear force (WBS) values as those of the conventionally housed pigs. Housing systems had an influence on the fatty acid composition. Stearic acid (C18:0) was significantly lower in the meat of the free-range pigs than that of the conventionally housed pigs while linoleic acid (C18:2n-6) and polyunsaturated fatty acid (PUFA) concentrations were significantly higher. However, the moisture, fat, protein and ash contents as well as the mineral composition in the meat were unaffected by housing systems. It could be concluded that pigs raised in a conventional housing system produced meat with similar quality characteristics to that of pigs raised in a free-range housing system.

Keywords: Free-range, housing systems, growth performance, carcass yield, physical quality, chemical composition

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#### Introduction

The 'ethical' management of animals, in particular that of animals produced in intensive systems, is becoming a more important issue throughout the entire animal industry. In the past few years there has been commercial interest in animal products originating from animal systems that could be considered 'natural' or 'traditional'. Some consumers do not consider modern intensive animal production systems synonymous with product quality (Barton Gade, 2002). There is also an increasing perception amongst consumers that there is widespread use of synthetic chemicals, both in medication and as growth promoters, in pig feed components (Barton Gade, 2002).

Intensive animal production systems, such as those practised in pig and poultry production, are being questioned by animal welfare groups, as these animals are bred, housed and fattened to slaughter in total confinement, an environment perceived to be unnatural (Sather *et al.*, 1997). The restriction of the animals from behaving naturally may lead to behavioural problems and aggression (Barton Gade, 2002). Warriss *et al.* (1983) and Beattie *et al.* (2000) suggested that pigs housed in a barren environment might be more susceptible to stress, which may affect the quality of pork. However, the management of conventional systems is more efficient in terms of controlling diseases and treating individual animals (Barton Gade, 2002). On the other hand, the advantage of free-range systems is that these animals have more freedom to express natural behaviour patterns. As such, these animals are less likely to behave aggressively and may be less susceptible to stress which may result in a better pork quality (Beattie *et al.*, 2000). However, the control in the environment in free-range systems is much more difficult with regard to the control and treatment of animals for diseases (Barton Gade, 2002). Therefore, there are some welfare and management advantages and disadvantages in both free-range and conventional housing systems which may positively or negatively influence the quality of pork.

Consumers, especially those from Europe, have an underlying desire to purchase pork products with some social attributes or with certain social assurances such as protection of the environment, ethical rearing

of the animals, development of the small farmer, etc. (Windhorst, 2001). A segment of consumers is willing to pay more for pork products with social assurances, even if improvements in sensory features are not expected or directly included in the presentation of the pork product (Gentry *et al.*, 2001). There are, however, some consumers who associate free-range products with a better taste and nutritional value than conventionally produced products (Nilzén *et al.*, 2001). This obviously opens niche markets for naturally produced pork.

The effect of housing systems on pork quality are inconclusive, though some studies showed benefits in housing outdoors (Beattie *et al.*, 2000), while others showed benefits from pigs housed indoors (Enfält *et al.*, 1997; Sather *et al.*, 1997). Enfält *et al.* (1997) and Sather *et al.* (1997) concluded that the meat derived from the outdoor housed pigs had a lower ultimate pH, higher drip loss (lower water-holding capacity) and higher shear force values than pigs housed indoors. These researchers also recorded higher percentages of lean meat from pigs housed outdoors, and consequently less tender meat. On the other hand, Beattie *et al.* (2000) reported that pigs from an enriched environment, e.g. access to field plants and outdoor housing, produced pork with greater tenderness (lower shear force values) than pigs raised in a barren environment. Warriss *et al.* (1983) and Van der Wal *et al.* (1993), however, found no difference in water-holding capacity or ultimate pH from indoor or outdoor housed pigs.

Studies comparing indoor and outdoor housing systems have thus produced inconsistent results and it appears that both positive and negative influences of free-range housing systems on pork quality might be obtained. The aim of this study was to compare the production potential and carcass yield, as well as the physical and chemical meat composition of pigs raised either in a conventional or in a free-range housing system. Such a study has not been performed under South African conditions, specifically in the Western Cape area.

### **Materials and Methods**

Twenty-four Landrace X Large White pigs (12 gilts and 12 boars) were divided equally (pertaining to sex) into two subgroups: a free-range (housed outdoors, FR) and conventionally housed (housed indoors, Conv) pigs. The pigs  $(27.65 \pm 3.757 \text{ kg live weight})$  were randomly selected out of six litters of similar aged weaners that had been housed together in a weaning facility. They were obtained from a commercial producer in the Western Cape and raised at the Elsenburg Agricultural Research Centre from September to November 2000 (spring, early summer period mean temperatures ranging between 14 and 28 °C). The freerange pigs were housed as a group in an 1800 m<sup>2</sup> pen. This pen had a group of conifers that provided shade as well as an A-frame hut (1.5 m x 1.5 m) built on the ground with a straw roof. The A-frame hut was filled with straw that was replaced as required. The other group of pigs was housed indoors in a 100 m<sup>2</sup> pen. This pen formed part of a group of pens that were inside a pig grower's shed. This shed was enclosed with walls and had a roof – manually opening or closing windows controlled airflow. The floor of the pen was concrete with a third of the pen being concrete slats above the faeces and urine drainage channel. Both groups received the same commercial diet ad libitum (13.8 MJ DE/kg, 170 g protein/kg; 10 g lysine/kg feed) from the same type of self-feeder and had free access to water (nipple supply). Every 14 days each individual pig was weighed and the back fat thickness (P2 fat measurement) was taken ultrasonically at the last rib, 45 mm from the spine. Feed intake was measured for each group of pigs. Both groups were slaughtered after 81 days in the trial. Prior to being slaughtered, feed was withheld for 10 h, but water was supplied until loading and transportation. The pigs were weighed again after the 10 h feed withdrawal period to record empty live weight. No electrical prodders were used to handle the pigs at any stage. Loading and transportation were done under conditions of minimal stress. This involved early morning transportation to avoid high temperatures, no overcrowding and no mixing of unfamiliar groups. Two vehicles were used in tandem. Transportation by road to the slaughter facility took approximately 15 min. The pigs were kept in lairage for approximately 1 h prior to slaughtering on the same day, the one group being slaughtered immediately after the other. The pigs were slaughtered according to standard commercial procedures. The latter consisted of electrical stunning (250 V AC, ear to ear for 3-5 s) and sticking within 30 s. Thereafter the pigs were eviscerated and inspected by the appropriate government health official. The P2 fat measurement was taken on each carcass with an intrascope between the 2<sup>nd</sup> and 3<sup>rd</sup> last rib, 45 mm from the carcass midline. This measurement was used to calculate the percentage lean meat yield of each carcass using the formula: lean% =  $74.4367 - 0.4023X_1$ , where  $X_1$  = fat-thickness in mm (Government Notice No. R 1748, 1992).

Each carcass was weighed warm and after chilling (2 °C for 24 h). Forty-five minutes post mortem, the initial pH was taken ( $pH_{45}$ ) and after the 24 h chilling period, the ultimate pH ( $pH_{24}$ ) was measured. The

pH measurements were taken on the *M. longissimus lumborum* (MLL) between the  $2^{nd}$  and  $3^{rd}$  lumbar vertebrae counting from the caudal end. The pH was measured with a penetrating glass electrode on a handheld Crison pH/mV-506 meter. The pH meter consisted of an automatic temperature compensator to ensure the adjustment of the pH for temperature. The pH meter was rinsed with distilled water after every reading and re-calibrated after every fourth reading.

After a chilling period of 24 h, the head was removed between the junction of the axis and atlas bones by means of a sharp knife. Thereafter, the removal of the primal cuts (shoulder, belly, back and legs) from the carcasses was done on a stationary band saw. The shoulder was removed by cutting caudally through the  $5^{th}$  and  $6^{th}$  thoracic vertebrae, with the front trotter removed by cutting through the metacarpal region (at the joint of the carcasses by cutting caudally at a line perpendicular to the spinal column, between the last lumbar and first sacral vertebrae. Sawing along the natural midline between the legs to split the two legs. The trotter was removed from the carcass. Sawing along the spinal column split the remaining carcass. Thereafter, the belly was removed from the back by cutting in a line parallel to the spinal column, *ca*. 18 cm from the spinal column (Fisher *et al.*, 2000).

For the physical and chemical analyses, the MLL (from the 1<sup>st</sup> to the last lumbar vertebrae) was removed from one of the backs, and trimmed of all visible subcutaneous fat. For the determination of the drip loss, 1.5 cm thick meat samples, cut perpendicular to the longitudinal axis of the muscle on the caudal side of the removed MLL, were weighed. Drip loss was determined according to the procedure described by Honikel (1998) with the samples weighed individually (approximately 110 g) and placed under atmospheric pressure in a net enclosed in a polythene bag in such a manner that the exudate did not come into contact with the sample, but collected in the bag. After a storage period of 24 h at 4 °C, the samples were dried and weighed again and drip loss was expressed as a percentage of the initial weight (Honikel, 1998). For cooking loss determination, samples were freshly cut (in the position next to that removed for drip loss determination) and weighed (initial weight). Individual slices of approximately 110 g (in thin-walled plastic bags) were placed in a water-bath at 75 °C. After one hour the samples were removed from the water-bath and cooled in cold water. The meat was removed from the bag, blotted dry, weighed and cooking loss was expressed as a percentage of the initial sample weight (Honikel, 1998). The cooking loss samples were also used to determine shear force values. Toughness was measured as the maximum force (N/1.27 cm diameter) required to shear a 1.27 cm diameter cylindrical core (five cores per MLL sample per pig) of cooked meat perpendicular to the grain, at a crosshead speed of 200 mm/min. The shear force measurements were generated with a Warner-Bratzler shear attachment, fitted to an Instron Universal Testing Machine. A higher reading indicated greater shear force and therefore tougher meat.

Prior to using the MLL sample for cooking loss determination, the sample was allowed to bloom for 20 mins, whereafter three readings per sample (on the cut surface perpendicular to the longitudinal axis of the muscle) were taken for colour determination. Colour was evaluated according to the method described by Honikel (1998) using a Colorgard System 2000 colorimeter (Pacific Scientific, Silver Spring, MD, USA) to determine L\*, a\* and b\* values with L\* indicating lightness, a\* the red-green range and b\* the blue-yellow range. These values were also used to calculate the chroma value and hue angle according to the following equations: chroma =  $\sqrt{(a^{*2}+b^{*2})}$ , and the hue angle (°) = tan<sup>-1</sup> (b\*/a\*).

Proximate chemical analyses were conducted on the fresh MLL samples. The frozen muscle samples were cut into smaller portions, minced three times through a 2 mm sieve to ensure homogeneity, and chemically analysed. Total concentrations of moisture, protein and ash were determined according to standard AOAC methods (AOAC, 1995). The protein (N x 6.25) content was determined by the block digestion method (AOAC, 1997) and ashing was done at 500 °C for a period of 5 h. The moisture content was analysed by drying a 2.5 g sample at 100 °C for a period of 24 h. The total fat content was determined by extracting the fat with a 2:1 mixture of chloroform: methanol (Lee *et al.*, 1996).

After the extraction of the lipids, the fatty acid methyl esters (FAME) were prepared according to the procedures published by Morrison & Smith (1964). The FAME were analysed with a gas-liquid chromatograph (Varian Model 3300), equipped with flame ionisation detection and two 30 m fused silica megabore DB-225 columns of 0.53 mm internal diameter (J&W Scientific, Folsom, CA). Gas flow rates were hydrogen 25 ml/min and nitrogen (carrier gas) 5-8 ml/min. The temperature programme was linear at 4 °C/min with initial and final temperatures of 160 °C and 220 °C (held for 10 min), respectively. The injector

temperature was 240 °C and the detector temperature 250 °C. The FAME was identified by comparison with the retention times of a standard FAME mixture (Nu-Chek-Prep Inc., Elysian, Minnesota).

A wet ashing method was used to prepare the samples for mineral analyses according to Method IIA of Watson (1994) and involves the boiling of the samples in concentrated nitric acid and perchloric acid. The concentrations of calcium (Ca), copper (Cu), iron (Fe), potassium (K), magnesium (Mg), sodium (Na), phosphorus (P), lead (Pb) and zinc (Zn) of the digestates were determined by direct current plasma emission spectrometry (Pinta, 1982).

The experiment consisted of a completely randomised design with two treatments (housing systems and sex). Analysis of variance (ANOVA) was performed on all the variables using the SAS Version 8.12 (SAS, 1990). The Shapiro-Wilk test was performed to test for non-normality (Shapiro & Wilk, 1965). In some cases deviations from normality were the cause of one or two outliers, which were removed before the final analysis. Where there was still significant evidence of non-normality, this could be ascribed to kurtosis rather than skewness. Student's t-least significant difference (LSD) was calculated at a 5% significance level to compare treatment comparison of means.

## **Results and Discussion**

For all the parameters tested there was no housing system x sex interaction. Sex had no influence on any of the parameters and the data were therefore pooled to test for the main effect of housing systems. As a group the free-range pigs consumed 1330 kg of feed (1368.31 g/pig/day), while the group of conventionally housed pigs consumed 1655 kg (1702.27 g/pig/day). This gave a calculated feed conversion ratio (FCR) of 2.17 and 2.35 kg feed per kg live weight gain, respectively. When the pigs were loaded onto the truck, it was noted that the free-range housed pigs had mosquito bites and fleas. It was also observed that the free-range housed pigs who had little contact with humans, seemed more stressed when they were loaded for transport to the abattoir than the conventionally reared ones. Similar findings were recorded by Hemsworth *et al.* (2002), who noted that free-range pigs showed a stronger fear response towards humans during the loading activity than conventionally reared ones. In contrast, the results of Warriss *et al.* (1983) showed that although free-range pigs were more easily startled than conventionally housed pigs, the latter were much more difficult to load into trucks than free-range housed pigs. Barton Gade & Blaabjerg (1989) also found that free-range pigs were calmer and handled easier in the abattoir than conventionally reared pigs.

The growth and carcass characteristics of the pigs from the free-range and conventionally housed systems can be seen in Table 1. Although there was no significant difference between the initial live weight and initial  $P_2$  measurements of the two housing systems, the conventionally housed pigs had a significantly higher weight gain (P = 0.035) as well as a significantly higher increase in subcutaneous fat (P<sub>2</sub>) (P = 0.01).

The average daily gain (ADG) of the free-range pigs was significantly lower (0.639  $\pm$  0.0605 kg) than that of the conventionally housed pigs (0.725  $\pm$  0.1255 kg). This agrees with the findings of Enfält *et al.* (1997) and Sather *et al.* (1997), that conventionally housed pigs grow faster compared to free-range pigs. There was a tendency for the conventionally housed pigs to have both a higher warm (P = 0.067) and cold (P = 0.057) dressed weight, but neither warm nor cold dressing percentage differed significantly between the two housing systems. Warriss *et al.* (1983) also found that the housing systems had no significant effect on dressing percentage. However, Sather *et al.* (1997) found a higher dressing percentage in conventionally housed pigs. The two housing systems did not influence the amount of moisture lost during the cooling regimen in the present investigation.

The carcasses of the conventionally housed pigs had higher (P = 0.051)  $P_2$  fat values and, therefore, lower ( $P \le 0.05$ ) calculated lean meat percentages (69%) than the free-range pigs (70%). Warriss *et al.* (1983), Enfält *et al.* (1997) and Sather *et al.* (1997) noted that free-range pigs had lower percentages of back fat thickness (lower  $P_2$  fat values) compared to indoor housed pigs. However, both the effects can be attributed to the higher body weight (Hoffman *et al.*, 1991) of the conventionally housed pigs compared to the free-range pigs.

Although there was a tendency for the conventionally housed pigs to have higher cold carcass weights, when the commercial cuts were expressed as a percentage of cold carcass weight, there were no significant differences (P > 0.05) between the two housing systems. These commercial yields are similar to those noted by Fisher *et al.* (2000) for South African commercial hybrids.

Housing systems			
	Conv (n=12)	FR (n=12)	Р
Initial body weight (kg)	$27.67 \pm 3.601$	$27.63 \pm 4.068$	0.983
Final body weight (kg)	$86.38 \pm 11.670$	$78.67 \pm 6.783$	0.064
Weight gain (kg) (over 81-day period)	58.71 ± 10.163	$51.60 \pm 5.600$	0.035
Average daily gain (kg/day)	$0.725 \pm 0.1255$	$0.639 \pm 0.0605$	0.049
Initial P <sub>2</sub> (mm)	$5.75 \pm 1.357$	$4.75 \pm 1.485$	0.099
Final P <sub>2</sub> (mm)	$14.08 \pm 1.621$	$11.25 \pm 1.815$	0.001
P <sub>2</sub> gain (mm)	$8.33 \pm 1.557$	$6.50 \pm 1.784$	0.010
Warm carcass weight (kg)	$67.17 \pm 9.833$	$60.92 \pm 4.963$	0.067
Cold carcass weight (kg)	$66.00 \pm 9.553$	$59.68 \pm 4.795$	0.057
Weight loss (kg)	$1.17\pm0.458$	$1.23 \pm 0.339$	0.690
<sup>a</sup> Dressing percentage (%)	$77.70 \pm 1.591$	$77.49 \pm 1.986$	0.780
<sup>b</sup> Dressing percentage (%)	$76.36 \pm 1.475$	$75.92 \pm 1.945$	0.548
<sup>c</sup> P <sub>2</sub> carcass (mm)	$12.83 \pm 2.209$	$10.75 \pm 2.701$	0.051
Calculated lean meat (%)	$69.27\pm0.889$	$70.11 \pm 1.087$	0.051
<sup>d</sup> Shoulder weight (%)	$13.65 \pm 1.811$	$13.70\pm0.745$	0.927
<sup>d</sup> Back weight (%)	$7.40\pm0.400$	$7.30\pm0.374$	0.688
<sup>d</sup> Leg weight (%)	$15.60 \pm 0.646$	$15.37 \pm 0.552$	0.389
<sup>d</sup> Belly weight (%)	$6.03 \pm 0.532$	$5.75\pm0.392$	0.157

Table 1 The growth and carcass characteristics of pigs (ca. 28-85 kg) raised either in a free-range of
in a conventional housing system (Mean $\pm$ s.d.)

<sup>a</sup> Warm carcass weight as a percentage of live weight

<sup>b</sup> Cold carcass weight as a percentage of live weight <sup>c</sup>Fat depth measured between the 2<sup>nd</sup>-3<sup>rd</sup> last rib, 45 mm in from the midline

<sup>d</sup>As a percentage of cold carcass weight

Conv = Conventionally housed pigs

FR = Free-range housed pigs

The physical meat quality attributes (water-holding capacity, shear force values, colour measurements and pH values) of the pigs from the two housing systems are depicted in Table 2. Neither cooking loss nor drip loss differed significantly (P > 0.05) between the free-range and conventionally housed pigs. Van der Wal et al. (1993) also found no differences in water-holding capacity between regular and free-range housed pigs. There was also no difference (P = 0.735) in the shear force (WBS) values of the two housing systems, which indicates that there were no differences in the physical tenderness of the meat. However, Enfält et al. (1997) found outdoor housed pigs to have a higher drip loss and higher shear force values than those of indoor housed pigs. The a\* values of the free-range pigs were significantly higher (P =0.043), indicating that the meat was more reddish in colour (also shown by the hue and chroma values) than that of the conventionally housed pigs. None of the other colour parameters differed statistically. In their study Warriss et al. (1983) reported that housing pigs outdoors resulted in a darker meat colour than it did in a conventionally housed system. Pigs are born with a predominance of Type 1 (darker red) fibres and as they develop there is a shift to Type IIA and Type IIB fibres (Gentry et al., 2001). These researchers noted that pigs in an outdoor system tend to have more of Type I muscle fibres at processing than pigs bred and housed indoors. The muscle pH values ( $pH_{45}$  and  $pH_{24}$ ) were not influenced by the two housing systems. However, Sather et al. (1997) found that initial muscle pH of free-range housed pigs tended to be lower than

that of conventionally housed pigs. Results of Barton Gade & Blaabjerg (1989) and Enfält *et al.* (1997) showed that free-range pigs had lower ultimate pH measurements compared to the indoor housed pigs. Therefore, Barton Gade (1987) suggested that free-range pigs had a tendency to produce higher levels of pale, soft and exudative (PSE) meat than conventionally housed pigs. These researchers reasoned that free-range pigs had higher levels of muscle glycogen than their pen-housed counterparts, which resulted in lower pH readings.

	Housing systems		
	Conv (n=12)	FR (n=12)	Р
Cooking loss (%)	22.59 ± 3.026	$23.97 \pm 4.036$	0.354
Drip loss (%)	4.11 ± 1.223	$3.91\pm0.891$	0.656
Shear force (N/1.27 cm diameter)	$117.20 \pm 12.045$	115.48 ± 12.465	0.735
Colour L*	$55.32 \pm 3.078$	53.98 ± 1.912	0.213
a*	$2.81\pm0.666$	$3.46\pm0.820$	0.043
b*	$10.10\pm0.690$	$10.47\pm0.672$	0.196
Hue (°)	$74.46 \pm 3.760$	$71.76 \pm 4.122$	0.108
Chroma	$10.48\pm0.959$	$11.03 \pm 1.060$	0.200
pH <sub>45</sub>	6.16±0.418	$6.22 \pm 0.391$	0.724
pH <sub>24</sub>	5.53 ± 0.138	$5.52 \pm 0.146$	0.966

**Table 2** Water-holding capacity, shear force values, colour measurements and pH values of the *M. longissimus lumborum* (MLL) from the free-range and conventionally housed pigs (Mean  $\pm$  s.d.)

 $L^* = Lightness; a^* = redness; b^* = yellowness$ 

Conv = Conventionally housed pigs

FR = Free-range pigs

Comparisons of the means for the proximate chemical composition of the MLL derived from the free-range and conventionally housed pigs are presented in Table 3. No significant differences (P > 0.05) were found with regard to moisture, fat, protein and ash content of the MLL between the two housing systems. There was a tendency for the meat from the free-range pigs to have a lower fat content (intramuscular fat, IMF) (1.52%) compared to the meat from the conventionally housed pigs (1.82%). This trend correlates with the thicker back fat (P<sub>2</sub> measurement, Table 2) of the conventionally housed pigs and thus corresponds to the development of the various different depots due to the growth of the pigs. This tendency is consistent with the results of Warriss *et al.* (1983), who found that pigs housed outdoors had significantly thinner back fat than the pigs housed indoors. These researchers also stated that the difference in back fat possibly illustrated the use of a corresponding amount of energy for the maintenance of body temperature and motor activity. Therefore, this tendency was not present to the same degree in the pigs housed indoors. Results of Enfält *et al.* (1997) also showed that the meat from pigs housed outdoors had less IMF compared to that of indoor housed pigs.

	Housing systems		
	Conv (n=12)	FR (n=12)	Р
Moisture	$74.54 \pm 0.913$	$74.79 \pm 1.312$	0.592
Lipid	$1.82\pm0.491$	$1.52 \pm 0.619$	0.206
Protein	22.31 ± 1.194	$22.83\pm0.710$	0.212
Ash	$1.20\pm0.141$	$1.29\pm0.203$	0.226

**Table 3** Comparison of the means for the proximate chemical composition of the *M. longissimus lumborum* (MLL) between the free-range and conventionally housed pigs (g/100 g meat sample) (Mean  $\pm$  s.d.)

Conv = Conventionally housed pigs

FR = Free-range pigs

Comparison of the means for the fatty acid composition of the MLL derived from the free-range and conventionally housed pigs is presented in Table 4. Palmitic acid (C16:0) was the most abundant SFA and oleic acid (C18:1n-9) the mono-unsaturated fatty acids (MUFA) and linoleic acid (C18:2n-6) the polyunsaturated fatty acids (PUFA) for the muscle of the pigs housed in both systems.

Stearic acid (C18:0), one of the major SFA differed significantly ( $P \le 0.05$ ) between the two housing The free-range pigs had a lower (P  $\leq$  0.05) C18:0 concentration compared to that of the systems. conventionally housed pigs. However, stearic acid (C18:0) is considered as a neutral fatty acid that has no effect on blood cholesterol (Mahan & Escott-Stump, 2000). Aside from C18:0, the concentrations of only some of the PUFA differed significantly ( $P \le 0.05$ ) between the two housing systems. Linoleic acid (C18:2n-6) was higher (P  $\leq$  0.05) in the MLL of the free-range pigs (70.4% of the total PUFA) compared to the MLL of the conventionally housed pigs (65% of the total PUFA). The higher C18:2n-6 concentration of the freerange pigs contributed to the higher (P  $\leq$  0.05) total PUFA concentration of the free-range pigs (24.65%) compared to that of the conventionally housed pigs (18.43%). This finding is consistent with the results of Nilzén et al. (2001). The higher level of this fatty acid in the FR pigs resulted in these animals having a higher calculated sum of n-6 fatty acids (21.4%) compared to the Conv pigs (15.56%). This also resulted in the FR having the higher n6:n3 ratio (6.54 vs. 5.44). The British Health authority recommends a value closer to 4 (Wiseman, 1997), to amongst others, help prevent coronary heart disease by reducing plasma lipids (Kinsella et al., 1990). In this respect the Conv housed pigs have the more desirable fatty acid profile. Table 4 also shows that the ratios of the total PUFA to SFA of the MLL differed significantly, with a higher ( $P \le$ 0.05) higher PUFA: SFA ratio for the free-range pigs (0.64) compared to that of the conventionally housed pigs (0.46). Both these ratios lie within the dietary guidelines of the British Committee on Medical Aspects of Food and Nutrition Policy (COMA), who recommend a PUFA: SFA ratio of >0.45 and <1.0 (as cited by Corino et al., 2002). However, it must be born in mind that a higher PUFA concentration could also result in a softer, oilier fat that could also be more prone to oxidation and rancidity that has negative consequences for human health.

Housing systems had no effect on the mineral composition of the MLL derived from the free-range and the conventionally housed pigs (Table 5). There was a tendency for the MLL derived from the free-range pigs to have a slightly higher concentration of Cu, Fe, Mg, Na, P and Zn compared to that of the conventionally housed pigs. Dworschák *et al.* (1995) found similar results with regard to the Zn and Cu concentration, with higher levels of Zn and Cu ( $\mu$ g/g) in the meat from the loin, neck-end and liver of pigs kept in natural farming conditions compared to that of pigs kept in large-scale farming conditions.

Housing systems			
	Conv (n=12)	FR (n=12)	Р
C14:0	$1.32 \pm 0.611$	$1.45 \pm 0.877$	0.663
C16:0	$23.69 \pm 5.867$	$23.80 \pm 2.443$	0.955
C18:0	$17.32 \pm 4.492$	$13.57 \pm 1.342$	0.014
C20:0	$0.02\pm0.038$	$0.02\pm0.040$	0.983
C22:0	$0.03\pm0.053$	$0.02\pm0.063$	0.728
C24:0	$0.02\pm0.056$	$0.02\pm0.039$	0.899
C16:1n-7	$3.33 \pm 2.293$	$2.59 \pm 1.152$	0.340
C18:1n-9	$35.38\pm5.039$	$33.47 \pm 4.108$	0.325
C20:1n-9	$0.43 \pm 0.215$	$0.15 \pm 0.143$	0.001
C24:1n-9	$0.05 \pm 0.056$	$0.28 \pm 0.328$	0.019
C18:2n-6	$11.98 \pm 4.114$	$17.35 \pm 4.312$	0.005
C18:3n-6	$0.09\pm0.132$	$0.21\pm0.142$	0.057
C18:3n-3	$1.11\pm0.847$	$1.22\pm0.697$	0.722
C20:2n-6	$1.50 \pm 1.367$	$1.74 \pm 1.335$	0.668
C20:3n-6	$0.39 \pm 0.284$	$0.43\pm0.329$	0.771
C20:4n-6	$0.84 \pm 0.710$	$0.70 \pm 1.018$	0.683
C20:3n-3	$0.07\pm0.130$	$0.01\pm0.022$	0.158
C20:5n-3	$0.80 \pm 1.045$	$1.46 \pm 1.707$	0.262
C22:2n-6	$0.04\pm0.068$	$0.06 \pm 0.114$	0.524
C22:4n-6	$0.72 \pm 1.012$	$0.91 \pm 1.078$	0.663
C22:5n-3	$0.49 \pm 0.468$	$0.29\pm0.231$	0.196
C22:6n-3	$0.39 \pm 0.622$	$0.29 \pm 0.411$	0.646
SFA	$42.39 \pm 7.469$	$38.88 \pm 2.950$	0.159
MUFA	$39.19 \pm 5.763$	$36.48 \pm 4.057$	0.204
PUFA	$18.43 \pm 3.331$	$24.65 \pm 2.984$	0.0001
PUFA:SFA	$0.46 \pm 0.148$	$0.64 \pm 0.094$	0.002

**Table 4** Comparison of the means for the fatty acid composition of the *M*. *longissimus dorsi* (MLL) between the free-range and conventionally housed pigs (% by weight of total fatty acids) (Mean  $\pm$  s.d)

SFA = Saturated fatty acids

MUFA = Mono-unsaturated fatty acids

PUFA = Polyunsaturated fatty acids

PUFA:SFA = Ratio of polyunsaturated to saturated fatty acids

Conv = Conventionally housed pigs

FR = Free-range pigs

### Conclusion

From the present investigation it can be concluded that the free-range housed pigs had a lower feed intake and slower growth rate than conventionally housed pigs. If pigs are therefore to be housed in a free-range production system, they would have to be retained for a longer period to reach their targeted slaughter weight. An aspect that warrants further research is what the effect of the lice and mosquito bites noted on the free-range pigs would have on meat quality and consumer acceptability, especially when the meat is sold as fresh with the skin intact.

Although the free-range pigs had a lower  $P_2$  fat depth and therefore leaner meat, these differences could be attributed to their lighter slaughter body weight and not to the production system. Housing system had no effect on the weight distribution of the commercial cuts (as a percentage of cold carcass weight). The meat from the free-range pigs was slightly more reddish in colour, but apart from that, housing systems had no effect on the WHC of the meat, its initial pH (pH<sub>45</sub>) or its final pH (pH<sub>24</sub>). The meat derived from the free-range pigs was just as tender as that of the conventionally housed (indoors) pigs.

Housing systems			
	Conv (n=12)	FR (n=12)	Р
Ca	$4.88 \pm 4.046$	$2.58 \pm 0.867$	0.067
Cu	$2.42 \pm 2.558$	$2.55 \pm 4.142$	0.930
Fe	$0.16 \pm 0.118$	$0.50\pm0.802$	0.231
К	tr <sup>a</sup>	tr <sup>a</sup>	-
Mg	$25.89 \pm 2.106$	$26.89 \pm 2.325$	0.281
Na	$23.09 \pm 1.956$	$23.96 \pm 2.366$	0.334
Р	$617.91 \pm 64.210$	$643.60 \pm 46.962$	0.275
Pb	$0.23\pm0.037$	$0.23\pm0.029$	0.939
Zn	$262.35 \pm 27.268$	$272.56 \pm 81.624$	0.685

**Table 5** Comparison of the means  $(\pm \text{ s.d.})$  for the mineral composition of the *M. longissimus luborum* (MLL) between the free-range and conventionally housed pigs (mg/100 g meat sample)

 $^{a}$ tr = trace = <0.001 mg/100 g meat sample

Conv = Conventionally housed pigs

FR = Free-range pigs

The moisture, fat, protein and ash content as well as the mineral composition did not differ significantly between the free-range and conventionally housed pigs. However, housing systems did influence the fatty acid composition of the fresh meat. Stearic acid (C18:0) was significantly lower in the meat of the free-range pigs than in the conventionally housed pigs. Higher linoleic acid (C18:2n-6) and PUFA concentrations were found in the free-range pigs compared to the conventionally housed pigs. The ratio of n-6:n-3 fatty acids was also such that the Conv housed pigs had the more desirable profile.

The decision whether one should raise pigs in free-range or in conventional systems in South Africa (Western Cape Province) should depend on the market. If the market is willing to purchase meat at a higher price for the assurance that the pigs were raised "under natural conditions", then the free-range production system could be used as a viable alternative. However, it can be concluded that pigs raised in a conventional housing system produced meat with similar quality characteristics to the meat produced from pigs raised in a free-range housing system. Further investigations are required to see what the effect of the housing systems on the commercial processing and intrinsic quality attributes of the processed pork would be.

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