Dietary strategies to treat hyperhomocysteinaemia based on the biochemistry of homocysteine: a review

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Abstract

Hyperhomocysteinaemia is implicated in various diseases, including cardiovascular disease and cancer. Several conditions influence the concentration of homocysteine (Hcy), including demographic, genetic and lifestyle factors. With regard to the latter, dietary components may be manipulated as Hcy can be remethylated to methionine by folate, or metabolised by other one-carbon nutrients, such as betaine and its precursor, choline. This metabolic interplay enables the nutritionist or dietitian to be able to lower Hcy concentrations cost-effectively by tailoring an individual's diet, or by food enrichment and fortification strategies. Evidence supports the safety and benefits of Hcy reduction by simple dietary intervention. B vitamins, and betaine and choline intake lower Hcy, whereas methionine and certain beverages (coffee, tea and alcohol) increase it. Therefore, dietary determinants of Hcy raise the prospect of a simple, inexpensive and safe means of treating and/or preventing diseases contingent on this sulphur-containing protein.

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Introduction

Elevated concentrations of homocysteine (Hcy), the basis of hyperhomocysteinaemia (HHcy), are involved in a myriad of diseases, including cardiovascular disease (CVD),^{1,2} and various other conditions, such as impaired bone health,³ inflammatory bowel disease⁴ and cancer.⁵ The underlying causes of HHcy arise from the interplay of genetic, biological and environmental factors, which can roughly be divided into two groups: those that are unchangeable, such as genetics, gender and advancing age; and modifiable factors, including smoking, diet and physical activity. In addition, acquired conditions, such as diseases (renal failure, rheumatoid arthritis, malignancies, psoriasis and infection with the human immunodeficiency virus) and certain drugs (methotrexate, nitrous oxide, theophylline and thiazides) can also lead to elevated Hcy concentrations.

Since Hcy can be lowered by diet, the nutritionist and dietitian can play an important role in the prevention and therapeutic treatment of individuals at increased risk of diseases that are contingent on Hcy. The metabolism of Hcy and dietary determinants explored in this review equip these professionals with the knowledge to treat HHcy in ways that are simple, inexpensive and safe.

An overview of the biochemistry and metabolism of homocysteine

Hcy is a sulphur-containing amino acid, i.e. a thiol (–SH), with the chemical formula: $\rm HSCH_2CH_2CH_2CH_2CH_6$ Hcy is not present in food, but is formed by the body as it metabolises methionine, an essential amino acid.⁷

The structures of methionine and Hcy are identical, except for a conversion that results in removing a one-carbon methyl group $(-CH_3)$ from the former.⁸ While methionine is chemically stable, the free thiol of Hcy renders it highly reactive in cells and within the circulation.⁷ Seventy per cent to eighty per cent of Hcy is bound to the thiol groups of plasma proteins in human plasma.⁷ Only 1-4% circulates as free Hcy in its reduced form, whereas the remainder auto-oxidises to form homocystine dimers, or combines with other thiols, such as cysteine (Cys) and glutathione to form mixed disulphides.^{6,9} It is not yet clear which complexes are formed by the reduced Hcy molecules within cells, but it seems probable, with the reactivity of the thiol group of Hcy, that similar complexes will form.⁷

The metabolism of B vitamins, methyl groups and Hcy is inextricably linked (Figure 1). The clearance of Hcy from the human circulation depends on folate, vitamin B_2 , vitamin B_2 and vitamin B_{12} .

In cells there are approximately 28 enzymes that can be identified as methyltransferases. Methyltransferases are involved in the donation

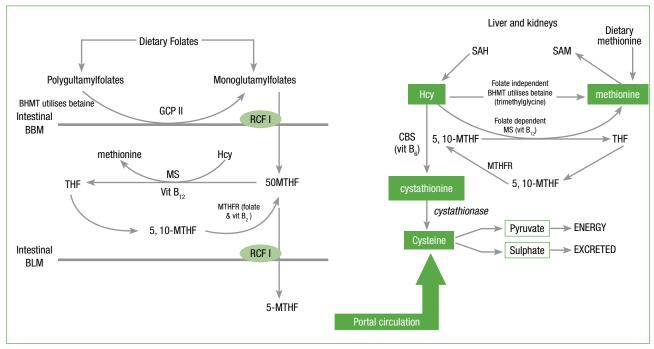


Figure 1: Homocysteine and folate metabolism¹⁰

5-MTHF: 5-methyltetrahydrofolate, 5,10-MTHF: 5,10-methylenetetrahydrofolate, BBM: brush border membrane, BHMT: betaine-homocysteine methyltransferase, BLM: basolateral membrane, CBS: cystathionine β-synthase, CSE: cystathionine γ-lyase, GCP II: glutamate carboxypeptidase II, Hcy: homocysteine, MS: methionine synthase, MTHFR: methylenetetrahydrofolate reductase, RFC I: reduced folate carrier 1, SAH: S-adenosylhomocysteine, SAM: S-adenosylmethionine, THF: tetrahydrofolate

(or transfer) of a methyl group from the activated form of methionine, namely, S-adenosylmethionine (SAM), to the synthesis of thymine, choline, creatine, epinephrine, the protein 3-methylhistidine or for deoxyribonucleic acid methylation.⁷ Methionine adenosyl-transferase converts methionine to SAM by a reaction that includes the addition of a methyl group and the purine base, adenine (from adenosine triphosphate or diphosphate). When SAM donates a methyl group, it is converted to S-adenosylhomocysteine (SAH). All SAM-dependent methyltransferase reactions result in the production of SAH, which can be catabolised immediately in vivo by SAH hydrolase to produce the nucleotide adenosine and Hcy.^{7,11} While the accumulation of SAH might in itself impair cellular function by inhibiting methyltransferase enzymes, thereby preventing the repair of aged or damaged cells,¹² Hcy, with its reactive free thiol group, could also be toxic to cells in its own right.⁷

Hcy can follow three possible pathways within cellular metabolism. First, Hcy may be remethylated back to methionine by either folate-dependent or folate-independent mechanisms.¹¹ During folate-dependent remethylation, methionine synthase (MS) uses a methyl group from 5-methyltetrahydrofolate (5-MTHF), while methylcobalamin (the biologically active form of vitamin B_{12}) acts as the coenzyme. The methyl group for this reaction is produced by the enzyme 5,10-MTHFR. MTHFR, in turn, uses flavin adenine dinucleotide (the biologically active form of vitamin B_2) as a cofactor.¹³ In an alternative folate-independent remethylation route, the enzyme betaine-homocysteine methyltransferase (BHMT) catalyses the remethylation of Hcy using betaine (trimethylglycine, which is a generator of methionine and SAM), a methyl group donor derived from choline oxidation, to convert Hcy to methionine and dimethylglycine. BHMT catalyses the synthesis of methionine from

betaine and Hcy, using a zinc ion to activate Hcy.¹⁴ Its expression is induced in mice on a methionine-deficient diet.¹⁵ SAM seems to function as a switch between the methionine cycle and the transsulphuration pathway. When SAM concentrations become limited, SAH and Hcy concentrations increase, and there is an accompanying reduction in the methylation cycle.¹⁶ High SAM concentrations seem to limit Hcy remethylation by inhibiting MTHF and BHMT. Transsulphuration seems to be enhanced by the stimulatory effect of SAM on cystathionine by cystathionine β -synthase (CBS) activity.¹⁶ This interplay is necessary for Hcy homeostasis and to prevent diseases contingent on HHcy.

Second, Hcy can be catabolised further to Cys by trans-sulphuration using pyridoxal 5'-phosphate (PLP) (the biologically active form of vitamin B₆) as a co-factor, beginning with the irreversible conversion to CBS. The enzyme, cystathionine γ -lyase (CSE or cystathionase), then uses CSE to form Cys, which is required for the synthesis of many compounds, including glutathionine, or can be further converted to pyruvate, which can be used for energy and sulphate, which is excreted through the urine.⁷ While the trans-sulphuration pathway contributes to the maintenance of normal postprandial Hcy concentrations, the remethylation pathway maintains normal fasting Hcy concentrations.¹⁷

Third, excess Hcy can be exported from the cell into the circulation, thereby regulating the intracellular Hcy concentrations.^{7,16} Earlier studies have indicated that the normal reference range for total plasma Hcy (i.e. the sum of free and protein-bound Hcy, Hcy and Hcy-Cys mixed disulphide) is 5-15 μ mol/l (to convert Hcy concentration to μ g/ml, multiply by 0.1352), mild to moderate HHcy between 16 μ mol/l and 100 μ mol/l, and severe HHcy > 100 μ mol/l.^{18,19} In

2007, Castañon, Lauricella, Kordich and Quintana¹ calculated that Hcy concentrations of > 12 µmol/l significantly increased the risk of developing venous thrombosis. Since the definition of elevated circulating Hcy concentrations should ideally be based on a health outcome, they proposed the use of 12 µmol/l as a cut-off value for HHcy, with clinical significance relating to CVD. The American Heart Association Nutrition Committee has recommended that basal Hcy should be 10 µmol/l as a therapeutic goal for subjects at increased risk of CVD.¹⁹ Cut-off values for diseases associated with Hcy other than CVD should be established in future studies.

It is important to note that standardisation of the methods (chromatography, capillary electrophoresis and immunoassays) used to quantify Hcy is still needed, since between-method and inter-laboratory variations in total Hcy quantification are not yet satisfactory.²⁰⁻²² According to Ubbink,²² fluorescence polarisation immunoassay may become the choice method for routine diagnostic laboratories.

Dietary determinants of homocysteine concentrations

The previously referred to description of the metabolism of Hcy shows that nutrition plays a major role in Hcy concentrations.

B vitamin intake

Insufficient B vitamin intake increases Hcy concentrations, and induces an imbalance of SAM and SAH,²³ whereas supplementation with B vitamins and their synthetic counterparts seems to reduce Hcy concentrations.²⁴ Hcy has been proposed to be a highly sensitive indicator of folate and vitamin B₁₂ status.²⁵

Folate status

Folate is a micronutrient found in green leafy vegetables and in some animal products, such as egg yolk. It is an important co-factor and methyl donor in converting Hcy to methionine. Reduced folate status inhibits the methylation cycle by failing to keep it supplied with methyl groups, resulting in reduced Hcy remethylation.²⁶ Folate deficiency, due to problems of malabsorption or inadequate dietary intake (malnutrition or alcoholism), is considered to be the most important nutritional cause of elevated Hcy.²⁷

Although everyone should consume sufficient folate, research highlights the necessity of adequate folate intake in drinkers of alcohol, as well as individuals who harbour the cytosine to thymine substitution at nucleotide position 677 (C677T) in the MTHFR gene, i.e. 677 TT genotype.²⁸⁻³⁰ Chiuve et al²⁸ reported that the adverse effects of moderate, but not excessive, alcohol use on Hcy concentrations may be overcome through adequate folate intake. MTHFR 677 TT homozygous individuals may require more folate for thermolabile MTHFR to function adequately and to ensure normal Hcy concentrations, than individuals who harbour the wild type alleles.²⁹ The adverse effects of a low intake of the methyl-related nutrients, including folate, with a high intake of alcohol, are additive in relation to cancer risk.³⁰

A high intake of natural folate from food reduces plasma Hcy concentrations,³¹ but folic acid (a synthetic chemically stable form of folate used in supplements and fortified foods) has been reported to be more effective than dietary folate in this regard, probably owing to its high bioavailability.³² Fasting Hcy concentrations decreased modestly after mandatory folic acid fortification in America in 1998.³³ Supplementation with folic acid alone, and in combination with other B vitamins, diminishes Hcy concentration.³⁴ It was concluded in a meta-analysis of 25 randomised controlled trials³⁴ that \geq 0.8 mg folic acid per day is required to achieve the maximal reduction in Hcy concentration, and that 0.2 mg and 0.4 mg were associated with a 60% and 90%, reduction of the maximal effect, respectively. Vitamin B₁₂ (0.4 mg/day) produced a further reduction of 7% [95% confidence interval (CI): 4-9%] in Hcy concentrations, but vitamin B₆ had no significant effect.

Evidence suggests that folic acid supplementation has a beneficial effect on the vascular endothelium by reducing Hcy, and through other mechanisms, for instance, by reducing oxidative stress.³⁵ In addition, acute administration of folic acid can restore impaired endothelial function induced by acute HHcy.³⁶ Folic acid was reported to have antioxidant properties and direct scavenging effects in vitro,³⁷ and may directly improve nitric oxide production by enhancing the enzymatic activity of nitric oxide synthase.³⁸ Hcy leads to increased cell proliferation and cell death through apoptosis and necrosis in vitro in smooth muscle cells,³⁹ but adding folic acid to the culture media. This is probably because of the increased remethylation of Hcy to methionine, and the reduced effects of Hcy on proliferation, apoptosis and necrosis.³⁹

Mager et al⁴⁰ reported that long-term folate-based vitamin therapy was independently associated with lower all-cause mortality in HHcy patients with CVD, and with reduced Hcy concentrations. By contrast, Albert et al⁴¹ reported that long-term supplementation with folic acid, and vitamins B_6 and B_{12} , did not reduce total CVD events in high-risk women, despite lowering Hcy significantly.

Riboflavin status

Riboflavin, or 7,8-dimethyl-10-ribityl-isoalloxazine (vitamin B_2), is present in a wide variety of foods. It is an essential precursor for the biosynthesis of the biologically active flavin adenine mononucleotide (FMN) and flavin adenine dinucleotide (FAD). FMN and FAD participate in a range of redoxication reactions, some of which are key to the function of aerobic cells.⁴² FAD is an essential co-factor for the folate-dependent enzyme, MTHFR, which metabolises folate to the form used in Hcy methylation, and for an enzyme that activates the vitamin B_6 precursor, pyridoxal, to the biologically active form, PLP.⁷ Thus, in theory, inadequate intake of riboflavin might give rise to increased plasma Hcy.⁷ In addition, riboflavin deficiency may exert some of its effects on Hcy by reducing the metabolism of other B vitamins, notably those of folate and vitamin B_6 .⁴²

Riboflavin intake emerged as a factor that influences Hcy in the Framingham Offspring cohort.⁴³ By contrast, Verhoef and

De Groot²⁷ reported that riboflavin has little influence on fasting Hcy concentrations. It may be relevant in individuals with the MTHFR 677 TT genotype, as riboflavin status was reported to be a modulator of Hcy in healthy adults, especially in those homozygous for the MTHFR C677T mutation.⁴⁴ Moat et al⁴⁵ confirmed a folate-riboflavin interaction in determining Hcy unrelated to the MTHFR genotype.

Vitamin B_e status

Reduced vitamin B_6 causes an accumulation of Hcy because CBS and CSE, which catalyse the trans-sulphuration of Hcy, are vitamin B_6 dependent.⁴⁶ The metabolism of vitamin B_6 is flavin dependent and impaired synthesis of PLP in the presence of a riboflavin deficiency has been demonstrated in studies on humans and animals.⁴⁷ McKinley et al⁴⁸ reported that low-dose vitamin B_6 supplementation effectively lowers Hcy in healthy elderly persons who are both folate and riboflavin replete.

Vitamin B₆ supplementation has little influence on fasting Hcy concentrations, although it may improve Hcy catabolism in elderly individuals.²⁷ It significantly reduces the post-methionine load increase in Hcy, and also reduces cystathionine concentrations, probably owing to enhanced CSE activity.⁴⁹

Vitamin B₁₂ status

Folate and cobalamin (as methylcobalamin) are involved as substrate and coenzyme, respectively, in the remethylation pathway of Hcy to methionine. Plasma Hcy increases considerably when there is an intracellular deficiency of folate or cobalamin, and is regarded as a sensitive marker of suboptimal vitamin concentrations.²⁵

Vitamin B₁₂ deficiencies are most commonly because of problems of malabsorption, e.g. alcoholism⁴⁷ or inadequate dietary intake, especially in individuals who follow a strict vegetarian diet since vitamin B₁₂ is found only in animal-source foods.⁵⁰ A reduced vitamin B₁₂ status prevents the proper functioning of the methylation cycle by directly reducing the activity of MS, one of the enzymes needed for the methylation cycle to turn.⁷ The enzyme, MS, is dependent on 5-MTHF as a methyl donor, but also on vitamin B₁₂ as methylcobalamin.⁷ A low vitamin B₁₂ status may alter Hcy by reducing its remethylation cycle towards methionine resynthesis in the same manner that low folate status alters Hcy metabolism.²⁶

Certain rare and drastic genetic mutations, such as the one described by Mudd et al in 1969,⁵¹ can lead to impaired vitamin B_{12} activation. This results in reduced MS activity, with a rise in urinary and plasma Hcy, as well as a reduction in the activity of methylmalonyl-CoA mutase (the other vitamin B_{12} -dependent enzyme), causing an accumulation of methylmalonic acid.⁵¹

Dietary lipotropics

Methionine, betaine and choline belong to a group of compounds called lipotropics.⁵² Lipotropics are compounds that have an affinity for lipids, and thus prevent or correct the excess accumulation of fat in the liver by promoting the transport of fatty acids from the liver to the tissues, or by accelerating the utilisation of fat in the liver itself.

Methionine

Methionine is an essential amino acid that is naturally found in dietary proteins. A typical Western diet contains 1.6-2.8 g of methionine per day,⁵³ which is more than that required for protein biosynthesis. Thus, the excess must be catabolised.⁷ The transamination of methionine only occurs at non-physiologically high concentrations, and instead, methionine catabolism takes place mainly in the liver, through its sequential conversion to SAM and SAH, and then to Hcy, followed by the trans-sulphuration pathway.⁷

For this reason, methionine loading tests (MLT) are often used to induce "stress" on Hcy metabolism, and could reveal any defect in Hcy metabolism (methionine intolerance) and cause a transient acute increase in Hcy concentrations. MLT permit the screening of 40-55% of persons⁵⁴ who may have clinically relevant HHcy which fasting Hcy determination alone may fail to identify. Ubbink et al⁵⁵ reported that the rise in Hcy after methionine loading is lower in black Africans than in Caucasians, which points towards a more effective Hcy metabolism. This seems to correlate with the lower prevalence of CVD seen in these subjects. Caucasians with an elevated risk of CVD showed higher Hcy concentrations after oral MLT.⁵⁶ It has been suggested that methionine handling capacity is more dependent on (genetically determined) enzyme activities, but fasting (basal) Hcy concentrations are more strongly influenced by environmental factors.⁵⁷

Oral MLT is associated with a small, but significant, enhancement of thrombin generation,⁵⁸ and is associated with impaired flow-mediated, endothelium-dependent vasodilatation.⁵⁹ Even low-dose methionine and animal protein intake was reported to increase Hcy and lead to the rapid onset of endothelial dysfunction,⁶⁰ suggesting that even diet-related increments in Hcy may contribute to the development and progression of atherosclerosis. It is unknown whether or not a diet that is rich in animal protein containing large amounts of methionine might activate platelets, increase thrombin production or induce endothelial dysfunction.

Betaine and choline intake

Betaine or trimethylglycine, and its precursor, choline, are major sources of methyl groups (one-carbon nutrients) in the diet,⁶¹ and thus can act as methyl donors in Hcy remethylation. Betaine generates methionine and SAM, then converts Hcy to methionine, using the enzyme, BHMT, independent of folate or vitamin B_{12} .⁷ Betaine can be found in food and it is estimated that a normal diet contains 0.5-2 g/day or can be synthesised endogenously from choline, also found in food.⁶² Choline is derived from the diet, as well as from *de novo* synthesis.⁶³

The consumption of betaine and choline can lower fasting Hcy concentrations to the same extent as folic acid, particularly in the setting of a high intake of methionine.²⁷ High doses of betaine (6 g/day and higher) can be used as Hcy-lowering therapy in individuals with HHcy because of inborn errors in their Hcy metabolism. Betaine has been reported to lower plasma Hcy in vitamin B_6 -resistant patients.⁶² In addition, betaine supplementation seems to lower fasting Hcy dose

dependently, up to 20% for a dose of 6 g/day, in healthy volunteers with normal Hcy concentrations. Betaine also reduces the increase in Hcy after MLT by up to 50%, whereas folic acid has no effect.⁶² Betaine could also lower Hcy independent of folate remethylation in the presence of ethanol.⁶⁴ In this regard, Chiuve et al⁶⁵ reported that total choline and betaine intake was inversely associated with Hcy in women, and that the strongest dose response was observed in women with a low methyl diet (high alcohol and inadequate folate intake). Therefore, Chuive et al⁶⁵ concluded that the remethylation of Hcy may be more dependent on the betaine pathway when methyl sources in the diet are low. Betaine and choline depletion in animals plays a role in the pathogenesis of homocystinuria owing to deficiencies of the MTHFR enzyme.⁶⁶

Thus, betaine and choline can be important food components that attenuate Hcy rises after meals. If Hcy plays a causal role in the development of CVD, a diet that is rich in betaine or choline might benefit cardiovascular health through its Hcy-lowering effects. However, betaine and choline may adversely affect serum lipid concentrations, which can increase the risk of CVD. It remains to be established whether or not the potential beneficial health effects of betaine and choline outweigh the possible adverse effects on serum lipids.

Coffee and tea consumption

The consumption of tea and coffee increases Hcy concentrations by up to 20%.²⁷ Habitual coffee drinking is positively associated with Hcy concentration in most, 43,67-70 but not all, 71-73 observational studies. Intervention trials indicated that the increasing effect of coffee consumption on Hcy concentrations may be causal.74-76 The vitamin B₆ concentration was markedly lower in a randomised crossover trial in the period when coffee was consumed, rather than when no coffee was drunk, but since vitamin B_e does not seem to influence fasting Hcy concentrations greatly, the significance of this finding is uncertain.⁷⁵ The constituent caffeine, which is a methyl xanthine, might be the culprit,^{43,75} because methyl xanthines are known to act as vitamin B₆ antagonists that may inhibit the conversion of Hcy to Cys.⁷⁵ The polyphenol, chlorogenic acid, which is present in coffee in the same amount as caffeine, may partly contribute to the increase in Hcy concentration. When polyphenols are metabolised, methyl groups from methionine are necessary, which results in a higher production of Hcv.⁷⁷ Both caffeine and chlorogenic acid are also present in tea, although in smaller doses, which explains the absence of a clear association between Hcy and tea consumption.77 The consumption of one litre of strong coffee daily may affect diet composition and other lifestyle factors. Therefore, it could influence Hcy concentrations indirectly,⁷⁸ but this has not yet been explored. It is also possible that the Hcy response to coffee may be modulated by the genetic factors mentioned earlier.78 Since the habit of coffee drinking is widespread, consequences at population level may not be negligible. However, the effect of coffee consumption on Hcy concentration is modest and much less than the changes associated with a variation in B vitamin status.

Alcohol consumption

Light to moderate alcohol consumption is associated with reduced mortality from CVD (the French paradox).⁷⁹ However, intermittent bouts of excessive consumption (binge drinking) and chronic high levels of alcohol consumption result in a sequelae of health problems, such as increased CVD (for example, cardiomyopathy and arrhythmia) morbidity, neurological disorders, certain cancers, chronic pancreatitis and liver cirrhosis.⁸⁰ Lifelong abstainers appear to be at a slightly higher risk than light or moderate consumers who are able to control their drinking.⁸¹ Therefore, total abstinence is unnecessary when the consumer can enjoy alcohol sensibly, but abstainers are not advised to start drinking in order to gain any claimed health benefits.⁸²

In a meta-analysis, Bagnardi et al⁸³ reported that regular heavy drinkers and heavy irregular binge drinkers showed significantly different pooled relative risks of 0.75 (95% CI: 0.64-0.89) and 1.10 (95% CI: 1.03-1.17), respectively, as opposed to abstainers, for developing CVD. Therefore, the pattern of drinking, as well as the amount consumed, plays a pivotal role in CVD development.

An elevated Hcy concentration is one of the myriad negative consequences of chronic alcoholism.84 The Hcy concentration seems to be twice as high, and plasma B vitamins lower, in chronic alcoholics, than in healthy controls. This is probably because of a combination of malnourishment, the direct effects of heavy alcohol intake on folate status and vitamin B_e (acetaldehyde dislodges vitamin B_e from its protective binding protein so that it is destroyed, and alcohol interferes with thiamine, folate and vitamin B₁₂ absorption), the decreased hepatic uptake and retention, and the increased urinary excretion of folate.85 A major difficulty in drawing conclusions about the effect of alcohol consumption on Hcy lies in distinguishing between the direct effects of alcohol and alcohol-induced malnutrition. An association between alcohol and Hcy has not been detected in some studies,^{86,87} but it has been demonstrated in several that moderate consumption inversely relates to Hcy concentration, compared with abstaining.^{88,89} However, moderate alcohol consumption in social drinkers is associated with increased Hcy,⁹⁰ and wine consumption, in particular, relates to Hcy in a J-shaped manner, i.e. Hcy is higher when alcohol consumption is high, lower when alcohol consumption is low or moderate, and tends to be slightly increased in individuals who do not consume any alcohol.68 Thus, the literature on the cardioprotective effects of moderate alcohol consumption, compared with non-drinking, in relation to Hcy, remains contentious.

It may be important to distinguish between the effects that can be attributed directly to alcohol, and those that may result from other constituents of alcoholic drinks. Different types of alcoholic beverages seem to influence Hcy in different ways. The results from studies to determine the effects of different alcoholic beverages are inconsistent.^{90,91} Beer consumption might be responsible for the inverse or absence of an association with alcohol consumption and Hcy concentration,^{43,68,88,91} but some studies report that it is positively associated with Hcy.⁹⁰ Researchers have ascribed the beneficial

Table I: Summary of the effects of dietary determinants on homocysteine concentrations, as a guide for dietitians

B vitamins	Intervention effect on homocysteine	Advice to patient
Riboflavin	Ļ	Meet DRI through the diet or supplementation
Vitamin B ₆	\downarrow	
Vitamin B ₁₂	\downarrow	
Folate or folic acid	\downarrow	
Dietary lipotropics		
Methionine	î	Do not exceed the DRI
Betaine and choline	Ļ	Meet the DRI through the diet, while monitoring the lipid profile
Beverages		
Coffee and tea	î	Avoid drinking unfiltered coffee, and do not consume coffee and tea excessively
Alcohol intake	ſ	Use alcohol sensibly, if at all

DRI: dietary reference intakes, $\downarrow : \ , \uparrow :$

 $\ensuremath{\overset{\bullet}{\leftarrow}}$ Limited data are available. Research is needed to make quantitative recommendations with regard to intake

effect of beer drinking on Hcy to its folate, riboflavin and vitamin B₆ content, all of which are important for enzymatic Hcy conversion. Wine consumption seems to elevate Hcy concentrations.^{90,91} It was revealed that Hcy in women showed a U-shaped curve with a minimum of 8.49 mmol/l at 10-20 g alcohol/day, whereas an inverse association was observed in men by Burger et al⁹² in a large cross-sectional study. Spirits, on the other hand, seem to elevate Hcy concentrations.^{90,91} In Africa, indigenous people brew traditional alcoholic beverages, such as sorghum, millet beers and *Mbamba*, the last of which is a concoction of water, bread, oats, pineapple juice, sugar and yeast. Sorghum beer has been reported to make positive contributions to dietary intake, particularly when the beer is brewed with a sorghum adjunct. Hence, the results for different types of alcoholic beverages are not clear.

The relationship between alcohol consumption and Hcy concentration is complex. Alcohol (ethanol) interferes with both the transmethylation and trans-sulphuration pathways of Hcy metabolism, either directly,⁹³ or through its metabolite, acetaldehyde,^{94,95} or through the indirect effects mediated by interactions with vitamin metabolism. Alcohol studies that have been conducted on humans are scarce, and when using chronic alcoholics, the results are often complicated by liver disease and/or dietary insufficiencies associated with alcohol consumption.

Acetaldehyde inhibits MS activity,⁹⁶ which could lead to HHcy, independent of vitamin status. It has been indicated in animal studies that alcohol consumption causes a compensatory increase in betaine, a Hcy methyltransferase that generates methionine from Hcy.⁸⁴ Although methionine levels seem to vary, methionine adenosyltransferase activity decreases in most, but not in all studies.^{84,97} The SAM to SAH ratio, an important determinant of methylation activity, has been reduced in most animal studies.⁹⁸ The alcohol intermediate, acetaldehyde, accelerates the intracellular degradation of PLP, reducing the PLP content of hepatic cells, despite an adequate dietary intake of vitamin B₆.⁹⁹ This inhibits the PLP-dependent enzymes, CBS and CSE, compromising Hcy transsulphuration. Nitric oxide production, secondary to augmented Hcy, inhibits MS activity, possibly by inactivating cobalamin,¹⁰⁰ which further disrupts MS activity, despite cobalamin supplementation, as well as directly inhibiting MS, owing to acetaldehyde.⁹⁶ It was reported in studies conducted on ethanol-fed rats that MS inhibition raises the "trapping" of folate as 5-MTHF.^{93,93}

Folate intake, as discussed previously, seems to be inversely associated with Hcy concentration, but this relation seems to be modified by the MTHFR C677T genotype and alcohol intake. Women with the thermolabile variant of MTHFR 677 or moderate alcohol intake demonstrated a significantly higher Hcy concentration at a low intake of folate, but adequate folate intake minimised these differences.²⁸ In addition, the elevation of Hcy in women who consumed low folate and drank moderate amounts of alcohol was greater in the presence of the variant, MTHFR 677 T allele.²⁸

Summary and conclusion

Since numerous clinical conditions are known to be associated with a high Hcy count, a reduction in Hcy concentration may be clinically relevant to prevent disease. However, it remains to be determined whether or not lowering Hcy would prevent clinical disease outcomes, such as stroke events. Randomised controlled trials that incorporate the dietary strategies reviewed here should clarify whether or not the nature of the observed association between Hcy and these diseases is indeed causal. Settling this dispute of causality is necessary since various modifiable factors have been identified that could be manipulated as a strategy to prevent and treat diseases contingent on Hcy if they were found to be causative. As summarised in Table I, Hcy can be lowered by the adequate intake of folate, and vitamins B₂, B₆ and B₁₂ (either through a diet containing lots of fruit, vegetables and some animal-derived foods, or the fortification of food with B vitamins), as well as betaine and choline, and the proscription of heavy irregular (binge) alcohol drinking. Dietary changes can modify Hcy concentrations in ways that are relevant to public health. Therefore, dietitians are able to manipulate Hcy, thereby raising the prospect of disease prevention.

References

- Castañon MM, Lauricella AM, Kordich L, Quintana I. Plasma homocysteine cutoff values for venous thrombosis. Clin Chem Lab Med. 2007;45(2):232-236.
- Humphrey LL, Fu R, Rogers K, et al. Homocysteine level and coronary heart disease incidence: a systematic review and meta-analysis. Mayo Clin Proc. 2008;83(11):1203-1212.
- Yang J, Hu X, Zhang Q, et al. Homocysteine level and risk of fracture: a meta-analysis and systematic review. Bone. 2012;51(3):376-382.
- Zintzaras E. Genetic variants of homocysteine/folate metabolism pathway and risk of inflammatory bowel disease: a synopsis and meta-analysis of genetic association studies. Biomarkers. 2010;15(1):69-79.
- Kohaar I, Kumar J, Thakur N, et al. Homocysteine levels are associated with cervical cancer independent of methylene tetrahydrofolate reductase gene (MTHFR) polymorphisms in Indian population. Biomarkers. 2010;15(1):61-68.

- Jacobsen DW. Practical chemistry of homocysteine and other thiols. In: Carmel R, Jacobsen DW, editors. Homocysteine in health and disease. Cambridge: Cambridge University Press, 2001; p. 9-20.
- Scott JM. Genetic variation of homocysteine metabolism and atherosclerosis. In: Bachmann C, Koletzko B, editors. Genetic expression and nutrition. Nestlé Nutrition Workshop Series, Pediatric Program, Vol 50 Nestec Ltd. Philadelphia: Vevey/Lippincott Williams and Wilkins, 2003; p. 1-24.
- Weir DG, Scott JM. Homocysteine as a risk factor for cardiovascular and related disease: nutritional implications. Nutr Res Rev. 1998;11(2):311-338.
- Mansoor MA, Svardal AM, Ueland PM. Determination of the in vivo redox status of cysteine, cysteinylglycine, homocysteine, and glutathione in human plasma. Anal Biochem. 1992;200(2):218-229.
- Devlin AM, Clarke R, Birks J, et al. Interactions among polymorphisms in folate-metabolizing genes and serum total homocysteine concentrations in a healthy elderly population. Am J Clin Nutr. 2006;83(3):708-713.
- Williams KT, Schalinske KL. New insights into the regulation of methyl group and homocysteine metabolism. J Nutr. 2007;137(2):311-314.
- Coppola A, Davi G, De Stefano V, et al. Homocysteine, coagulation, platelet function, and thrombosis. Semin Thromb Hemost. 2000;26(3):243-254.
- Guenther BD, Sheppard CA, Tran P, et al. The structure and properties of methylenetetrahydrofolate reductase from Escherichia coli suggest how folate ameliorates human hyperhomocysteinemia. Nat Struct Biol. 1999;6(4):359-365.
- Evans JC, Huddler DP, Jiracek J, et al. Betaine-homocysteine methyltransferase: zinc in a distorted barrel. Structure. 2002;10(9):1159-1171.
- Park El, Garrow TA. Interaction between dietary methionine and methyl donor intake on rat liver betaine-homocysteine methyltransferase gene expression and organization of the human gene. J Biol Chem. 1999;274(12):7816-7824.
- Finkelstein JD. Regulation of homocysteine metabolism. In: Carmel R, Jacobsen DW, editors. Homocysteine in health and disease. Cambridge: Cambridge University Press, 2001; p. 92-99.
- Rosenblatt DS. Inherited disorders of folate transport and metabolism. In: Scriver CR, Beaudet AI, Sly WS, Valle D, editors. The metabolic basis of inherited disease. New York: McGraw-Hill, 1989; p. 2049-2064.
- Eikelboom JW, Lonn E, Genest J Jr, et al. Homocyst(e)ine and cardiovascular disease: a critical review of the epidemiologic evidence. Ann Intern Med. 1999;131(5):363-375.
- Malinow MR, Bostom AG, Krauss RM. Homocyst(e)ine, diet, and cardiovascular diseases: a statement for healthcare professionals from the Nutrition Committee, American Heart Association. Circulation. 1999;99(1):178-182.
- Hanson NQ, Eckfeldt JH, Schwichtenberg K, et al. Interlaboratory variation of plasma total homocysteine measurements: results of three successive homocysteine proficiency testing surveys. Clin Chem. 2002;48(9):1539-1545.
- Ubbink JB, Delport R, Riezler R, Vermaak WJ. Comparison of three different plasma homocysteine assays with gas chromatography-mass spectrometry. Clin Chem. 1999;45(5):670-675.
- Ubbink JB. Assay methods for the measurement of total homocyst(e)ine in plasma. Semin Thromb Hemost. 2000;26(3):233-241.
- Fuso A, Nicolia V, Cavallaro RA, et al. B-vitamin deprivation induces hyperhomocysteinemia and brain S-adenosylhomocysteine, depletes brain S-adenosylmethionine, and enhances PS1 and BACE expression and amyloid-beta deposition in mice. Mol Cell Neurosci. 2008;37(4):731-746.
- Clarke R, Armitage J. Vitamin supplements and cardiovascular risk: review of the randomized trials of homocysteine-lowering vitamin supplements. Semin Thromb Hemost. 2000;26(3):341-348.
- Bates CJ, Mansoor MA, van der Pols J, et al. Plasma total homocysteine in a representative sample of 972 British men and women aged 65 and over. Eur J Clin Nutr. 1997;51(10):691-697.
- Beccia M, Mele MC, Ferrari M, et al. Young stroke and basal plasma and post-methionine load homocysteine and cysteine levels 1 year after the acute event: do plasma folates make the difference? Eur J Neurol. 2004;11(4):269-275.
- Verhoef P, de Groot LC. Dietary determinants of plasma homocysteine concentrations. Semin Vasc Med. 2005;5(2):110-123.
- Chiuve SE, Giovannucci EL, Hankinson SE, et al. Alcohol intake and methylenetetrahydrofolate reductase polymorphism modify the relation of folate intake to plasma homocysteine. Am J Clin Nutr. 2005;82(1):155-162.
- Jacques PF, Bostom AG, Williams RR, et al. Relation between folate status, a common mutation in methylenetetrahydrofolate reductase, and plasma homocysteine concentrations. Circulation. 1996;93(1):7-9.
- Bailey LB. Folate, methyl-related nutrients, alcohol, and the MTHFR 677C--> T polymorphism affect cancer risk: intake recommendations. J Nutr. 2003;133(11 Suppl 1):3748S-3753S.
- Verhaar MC, Stroes E, Rabelink TJ. Folates and cardiovascular disease. Arterioscler Thromb Vasc Biol. 2002;22(1):6-13.

- Winkels RM, Brouwer IA, Siebelink E, et al. Bioavailability of food folates is 80% of that of folic acid. Am J Clin Nutr. 2007;85(2):465-473.
- Anderson JL, Jensen KR, Carlquist JF, et al. Effect of folic acid fortification of food on homocysteine-related mortality. Am J Med. 2004;116(3):158-164.
- Homocysteine lowering trialists' collaboration. Dose-dependent effects of folic acid on blood concentrations of homocysteine: a meta-analysis of the randomized trials. Am J Clin Nutr. 2005;82(4):806-812.
- Wilmink HW, Stroes ES, Erkelens WD, et al. Influence of folic acid on postprandial endothelial dysfunction. Arterioscler Thromb Vasc Biol. 2000;20(1):185-188.
- Usui M, Matsuoka H, Miyazaki H, et al. Endothelial dysfunction by acute hyperhomocyst(e) inaemia: restoration by folic acid. Clin Sci (Lond). 1999;96(3):235-239.
- Verhaar MC, Wever RM, Kastelein JJ, et al. 5-methyltetrahydrofolate, the active form of folic acid, restores endothelial function in familial hypercholesterolemia. Circulation. 1998;97(3):237-241.
- 38 Stroes ES, van Faassen EE, Yo M, et al. Folic acid reverts dysfunction of endothelial nitric oxide synthase. Circ Res. 2000;86(11):1129-1134.
- Buemi M, Marino D, Di Pasquale G, et al. Effects of homocysteine on proliferation, necrosis and apoptosis of vascular smooth muscle cells in culture and influence of folic acid. Thromb Res. 2001;104(3):207-213.
- 40 Mager A, Orvin K, Koren-Morag N, et al. Impact of homocysteine-lowering vitamin therapy on long-term outcome of patients with coronary artery disease. Am J Cardiol. 2009;104(6):745-749.
- Albert CM, Cook NR, Gaziano JM, et al. Effect of folic acid and B vitamins on risk of cardiovascular events and total mortality among women at high risk for cardiovascular disease: a randomized trial. JAMA. 2008;299(17):2027-2036.
- 42. Powers HJ. Riboflavin (vitamin B-2) and health. Am J Clin Nutr. 2003;77(6):1352-1360.
- Jacques PF, Bostom AG, Wilson PW, et al. Determinants of plasma total homocysteine concentration in the Framingham Offspring cohort. Am J Clin Nutr. 2001;73(3):613-621.
- Hustad S, Ueland PM, Vollset SE, et al. Riboflavin as a determinant of plasma total homocysteine: effect modification by the methylenetetrahydrofolate reductase C677T polymorphism. Clin Chem. 2000;46(8 Pt 1):1065-1071.
- Moat SJ, Ashfield-Watt PA, Powers HJ, et al. Effect of riboflavin status on the homocysteinelowering effect of folate in relation to the MTHFR (C677T) genotype. Clin Chem. 2003;49(2):295-302.
- 46. Finkelstein JD. Methionine metabolism in mammals. J Nutr Biochem. 1990;1(5):228-237.
- Lakshmi AV, Bamji MS. Regulation of blood pyridoxal phosphate in riboflavin deficiency in man. Nutr Metab. 1976;20(4):228-233.
- McKinley MC, McNulty H, McPartlin J, et al. Low-dose vitamin B-6 effectively lowers fasting plasma homocysteine in healthy elderly persons who are folate and riboflavin replete. Am J Clin Nutr. 2001;73(4):759-764.
- Bleie Ø, Refsum H, Ueland PM, et al. Changes in basal and postmethionine load concentrations of total homocysteine and cystathionine after B vitamin intervention. Am J Clin Nutr. 2004;80(3):641-648.
- Obersby D, Chappell DC, Dunnett A, Tsiami AA. Plasma total homocysteine status of vegetarians compared with omnivores: a systematic review and meta-analysis. Br J Nutr. 2013;109(5):785-794.
- Mudd SH, Levy HL, Abeles RH, Jennedy JP Jr. A derangement in B 12 metabolism leading to homocystinemia, cystathioninemia and methylmalonic aciduria. Biochem Biophys Res Commun. 1969;35(1):121-126.
- Snyder F, Cornatzer WE, Simonson GE. Comparative lipotropic and lipide phosphorylating effects of choline, betaine and inositol. Proc Soc Exp Biol Med. 1957;96(3):670-672.
- Shoob HD, Sargent RG, Thompson SJ, et al. Dietary methionine is involved in the etiology of neural tube defect-affected pregnancies in humans. J Nutr. 2001;131(10):2653-2658.
- Bostom AG, Jacques PF, Nadeau MR, et al. Post-methionine load hyperhomocysteinemia in persons with normal fasting total plasma homocysteine: initial results from the NHLBI Family Heart Study. Atherosclerosis. 1995;116(1):147-151.
- Ubbink JB, Vermaak WJ, Delport R, et al. Effective homocysteine metabolism may protect South African blacks against coronary heart disease. Am J Clin Nutr. 1995;62(4):802-808.
- Simporè J, Pignatelli S, Meli C, et al. Nutritional and racial determinants of the increase in plasma homocysteine levels after methionine loading. Curr Ther Res. 2002:63(7):459-474.
- Den Heijer M, Graafsma S, Lee SY, et al. Homocysteine levels--before and after methionine loading: in 51 Dutch families. Eur J Hum Genet. 2005;13(6):753-762.
- Domagala TB, Undas A, Sydor WJ, Szczeklik A. Thrombin formation in platelet-rich plasma after oral methionine loading: preliminary report. Thromb Res. 2002;105(6):503-506.
- Bellamy MF, McDowell IF, Ramsey MW, et al. Hyperhomocysteinemia after an oral methionine load acutely impairs endothelial function in healthy adults. Circulation. 1998;98(18):1848-1852.

- Chambers JC, McGregor A, Jean-Marie J, et al. Demonstration of rapid onset vascular endothelial dysfunction after hyperhomocysteinemia: an effect reversible with vitamin C therapy. Circulation. 1999;99(9):1156-1160.
- Cho E, Zeisel SH, Jacques P, et al. Dietary choline and betaine assessed by food-frequency questionnaire in relation to plasma total homocysteine concentration in the Framingham Offspring Study. Am J Clin Nutr. 2006;83(4):905-911.
- Olthof MR, Verhoef P. Effects of betaine intake on plasma homocysteine concentrations and consequences for health. Curr Drug Metab. 2005;6(1):15-22.
- Zeisel SH. Gene response elements, genetic polymorphisms and epigenetics influence the human dietary requirement for choline. IUBMB Life. 2007;59 (6):380-387.
- BarakAJ, Beckenhauer HC, Mailliard ME, et al. Betaine lowers elevated s-adenosylhomocysteine levels in hepatocytes from ethanol-fed rats. J Nutr. 2003;133(9):2845-2848.
- Chiuve SE, Giovannucci EL, Hankinson SE, et al. The association between betaine and choline intakes and the plasma concentrations of homocysteine in women. Am J Clin Nutr. 2007;86(4):1073-1081.
- Schwahn BC, Laryea MD, Chen Z, et al. Betaine rescue of an animal model with methylenetetrahydrofolate reductase deficiency. Biochem J. 2004;382(Pt 3):831-840.
- De Bree A, Verschuren WM, et al. Lifestyle factors and plasma homocysteine concentrations in a general population sample. Am J Epidemiol. 2001;154(2):150-154.
- Husemoen LL, Thomsen TF, Fenger M, Jørgensen T. Effect of lifestyle factors on plasma total homocysteine concentrations in relation to MTHFR(C677T) genotype. Inter99 (7). Eur J Clin Nutr. 2004;58(8):1142-1150.
- Nygård O, Refsum H, Ueland PM, et al. Coffee consumption and plasma total homocysteine: the Hordaland Homocysteine Study. Am J Clin Nutr. 1997;65(1):136-143.
- Stolzenberg-Solomon RZ, Miller ER 3rd, Maguire MG, et al. Association of dietary protein intake and coffee consumption with serum homocysteine concentrations in an older population. Am J Clin Nutr. 1999;69(3):467-475.
- Nieto FJ, Comstock GW, Chambless LE, Malinow RM. Coffee consumption and plasma homocyst(e)ine: results from the Atherosclerosis Risk in Communities Study. Am J Clin Nutr. 1997;66(6):1475-1477.
- Rasmussen LB, Ovesen L, Bülow I, et al. Folate intake, lifestyle factors, and homocysteine concentrations in younger and older women. Am J Clin Nutr. 2000;72(5):1156-1163.
- Saw SM, Yuan JM, Ong CN, et al. Genetic, dietary, and other lifestyle determinants of plasma homocysteine concentrations in middle-aged and older Chinese men and women in Singapore. Am J Clin Nutr. 2001;73(2):232-239.
- Christensen B, Mosdol A, Retterstol L, et al. Abstention from filtered coffee reduces the concentrations of plasma homocysteine and serum cholesterol: a randomized controlled trial. Am J Clin Nutr. 2001;74(3):302-307.
- Grubben MJ, Boers GH, Blom HJ, et al. Unfiltered coffee increases plasma homocysteine concentrations in healthy volunteers: a randomized trial. Am J Clin Nutr. 2000;71(2):480-484.
- Urgert R, van Vliet T, Zock PL, Katan MB. Heavy coffee consumption and plasma homocysteine: a randomized controlled trial in healthy volunteers. Am J Clin Nutr. 2000;72(5):1107-1110.
- Olthof MR, Hollman PC, Zock PL, Katan MB. Consumption of high doses of chlorogenic acid, present in coffee, or of black tea increases plasma total homocysteine concentrations in humans. Am J Clin Nutr. 2001;73(3):532-538.
- Vollset SE, Nygârd O, Refsum H, Ueland PM. Coffee and homocysteine. Am J Clin Nutr. 2000;71(2):403-404.
- Renaud S, de Lorgeril M. Wine, alcohol, platelets, and the French paradox for coronary heart disease. Lancet. 1992;339(8808):1523-1526.
- Corrao G, Bagnardi V, Zambon A, La Vecchia C. A meta-analysis of alcohol consumption and the risk of 15 diseases. Prev Med. 2004;38(5):613-619.

- Yuan JM, Ross RK, Gao YT, et al. Follow up study of moderate alcohol intake and mortality among middle aged men in Shanghai, China. BMJ. 1997;314(7073):18-23.
- Van Heerden IV, Parry CDH. If you drink alcohol, drink sensibly. South Afr J Clin Nutr. 2001;14(3):S71-S77.
- Bagnardi V, Zatonski W, Scotti L, et al. Does drinking pattern modify the effect of alcohol on the risk of coronary heart disease? Evidence from a meta-analysis. J Epidemiol Community Health. 2008;62(7):615-619.
- Carmel R, James SJ. Alcohol abuse: an important cause of severe hyperhomocysteinemia. Nutr Rev. 2002;60 (7 Pt 1):215-221.
- Cravo ML, Glória LM, Selhub J, et al. Hyperhomocysteinemia in chronic alcoholism: correlation with folate, vitamin B-12, and vitamin B-6 status. Am J Clin Nutr. 1996;63 (2):220-224.
- Gudnason V, Stansbie D, Scott J, et al. C677T (thermolabile alanine/valine) polymorphism in methylenetetrahydrofolate reductase (MTHFR): its frequency and impact on plasma homocysteine concentration in different European populations. EARS group. Atherosclerosis. 1998;136(2):347-354.
- Lussier-Cacan S, Xhignesse M, Piolot A, et al. Plasma total homocysteine in healthy subjects: sex-specific relation with biological traits. Am J Clin Nutr. 1996;64(4):587-593.
- Mayer O Jr, Simon J, Rosolová H. A population study of the influence of beer consumption on folate and homocysteine concentrations. Eur J Clin Nutr. 2001;55(7):605-609.
- Vollset SE, Hygard O, Dvale G, et al. The Hordaland Homocysteine Study: Lifestyle and total plasma homocysteine in Western Norway. In: Graham I, Refsum H, Rosenberg IH, Ueland PM, editors. Homocysteine metabolism: from basic science to clinical medicine. Massachusetts: Kluwer Academic Publishers, 1997; p. 177-182.
- Bleich S, Bleich K, Kropp S, et al. Moderate alcohol consumption in social drinkers raises plasma homocysteine levels: a contradiction to the "French paradox"? Alcohol Alcohol. 2001;36(3):189-192.
- Van der Gaag MS, Ubbink JB, Sillanaukee P, et al. Effect of consumption of red wine, spirits, and beer on serum homocysteine. Lancet. 2000;355(9214):1522.
- Burger M, Mensink G, Brönstrup A, et al. Alcohol consumption and its relation to cardiovascular risk factors in Germany. Eur J Clin Nutr. 2004;58(4):605-614.
- Barak AJ, Beckenhauer HC, Hidiroglou N, et al. The relationship of ethanol feeding to the methyl folate trap. Alcohol. 1993;10(6):495-497.
- Hidiroglou N, Camilo ME, Beckenhauer HC, et al. Effect of chronic alcohol ingestion on hepatic folate distribution in the rat. Biochem Pharmacol. 1994;47(9):1561-1566.
- Shaw S, Jayatilleke E, Herbert V, Colman N. Cleavage of folates during ethanol metabolism. Role of acetaldehyde/xanthine oxidase-generated superoxide. Biochem J. 1989;257(1):277-280.
- Kenyon SH, Nicolaou A, Gibbons WA. The effect of ethanol and its metabolites upon methionine synthase activity in vitro. Alcohol. 1998;15(4):305-309.
- Finkelstein JD, Cello JP, Kyle WE. Ethanol-induced changes in methionine metabolism in rat liver. Biochem Biophys Res Commun. 1974;61(2):525-531.
- Halsted CH, Villanueva J, Chandler CJ, et al. Ethanol feeding of micropigs alters methionine metabolism and increases hepatocellular apoptosis and proliferation. Hepatology. 1996;23(3):497-505.
- Lumeng L, Ryan MP, Li TK. Validation of the diagnostic value of plasma pyridoxal 5'-phosphate measurements in vitamin B6 nutrition of the rat. J Nutr. 1978;108(4):545-553.
- 100. Danishpajooh IO, Gudi T, Chen Y, et al. Nitric oxide inhibits methionine synthase activity in vivo and disrupts carbon flow through the folate pathway. J Biol Chem. 2001;276(29):27296-27303.