

## Optimal composition of intravenous lipids

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### Abstract

The provision of energy from a lipid source is an essential component of any parenteral nutrition (PN) therapeutic regimen in the appropriate clinical setting. All available sources of intravenous lipid emulsions have a low osmolarity but they strongly differ in their immunologic effects and their effects on oxidative stress, liver injury and mitochondrial function. The  $\omega$ -9/ $\omega$ -6 lipid emulsion with its relative immunoneutrality and also the newer fish oil admixtures are lipid emulsions that can be used in most critically ill and non-critically ill patients. Despite extensive research and encouraging progress in the availability of such lipid emulsions, there is still need for a lipid emulsions that could be advantageous in patients with real hyperinflammation.

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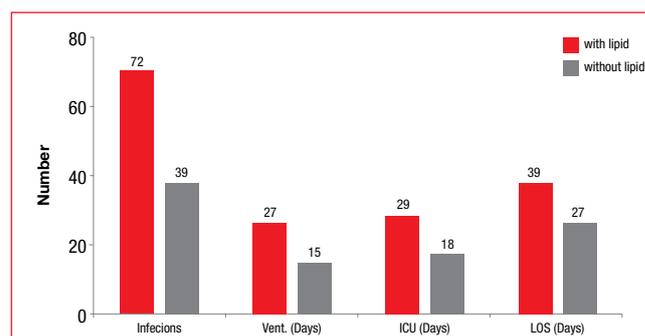
The composition of an intravenous (IV) lipid emulsion is of great importance in parenteral nutrition (PN) therapy, as most of its effects depend on the kind of fatty acids included and their respective ratio to each other. Today's lipid emulsions may include four classes of different fatty acids (FA), namely  $\omega$ -6 long chain polyunsaturated fatty acids,  $\omega$ -3 long chain polyunsaturated fatty acids,  $\omega$ -9 long chain monounsaturated fatty acids and medium chain saturated fatty acids. All these fatty acids present a source of energy with a low osmolarity but they strongly differ in their immunologic effects and their effects on oxidative stress, liver injury and mitochondrial function. For this review the effects of the different admixtures will be discussed following the time line of their landmark introduction into clinical practice.

The first lipid emulsion for parenteral application was a 100% soy bean oil-based emulsion introduced in 1961. There were two reasons for the development of this type of lipid emulsion. Firstly, soy bean oil contains the two essential  $\omega$ -6 fatty acids, linoleic acid and linolenic acids, and secondly, soy bean oil was readily available. Very soon after its introduction into clinical practice its immunologic properties came into discussion<sup>1</sup> and with its widespread use concerns were raised regarding its immune-compromising effects and the increased risk of infections.<sup>2,3</sup> Today, there is consistent evidence from human and animal *in vitro* and *ex vivo* studies showing that  $\omega$ -6 fatty acids or a soy bean oil-based emulsion inhibit lymphocyte proliferation,<sup>4,5</sup> decrease natural and lymphokine activated killer cell activity,<sup>6,7</sup> decrease chemotaxis and random migration of granulocytes<sup>8-10</sup> and negatively affect the reticuloendothelial system.<sup>11,12</sup>

Despite these impressive experimental results, some smaller trials in malnourished or paediatric patients<sup>13-15</sup> and one large clinical trial in bone marrow transplanted patients<sup>16</sup> found no significant difference between patients who received a parenteral  $\omega$ -6 emulsion and

those who did not. One smaller crossover study even reported an immunorestorative effect of the lipid emulsion.<sup>17</sup> In contrast, the study reported by Battistella<sup>18</sup> randomised 60 trauma patients with a severity score of 27 and an APACHE II score of 23 to 10 days of postoperative PN with our without an  $\omega$ -6 emulsion. Patients who received the lipid emulsion had a longer length of hospitalisation (39 vs 27 days), a longer length of stay in the intensive care unit (29 vs 18 days) and more days of mechanical ventilation (27 vs 15) (Figure 1) The group which received the lipid emulsion also had a significantly reduced natural and lymphokine activated killer cell activity and a higher number of infections (72 vs 39). Although this trial is somewhat biased by the fact that the group which did not receive the lipid emulsion also received 25% less energy, these negative results made the Canadian, the European and also the American guideline committee recommend that – at least – intensive care patients should not receive pure  $\omega$ -6 lipid emulsions as part of their PN regimens.

Figure 1: Battistella FD et al. Lipid emulsion in trauma victims (Figure was created using data from this publication)<sup>18</sup>



In order to reduce the amount of  $\omega$ -6 fatty acids in a lipid emulsion, medium chain triglycerides were introduced into clinical practice in 1984. These fatty acids with a chain length of 8 resp. 10 carbon atoms are derived from coconut oil. *In vitro* and *ex vivo* studies could show that a 50/50 admixture of  $\omega$ -6 FA and MCT ( $\omega$ -6/MCT) could prevent the inhibition of lymphocyte proliferation,<sup>19</sup> that natural killer cell activity was less reduced,<sup>20</sup> and that the expression of adhesion molecules<sup>21,22</sup> as well as the phagocytic capacity of the RES were increased.<sup>23,24</sup> On the other hand it was also shown that the phagocytic capacity for *C. albicans* was significantly reduced.<sup>25,26</sup>

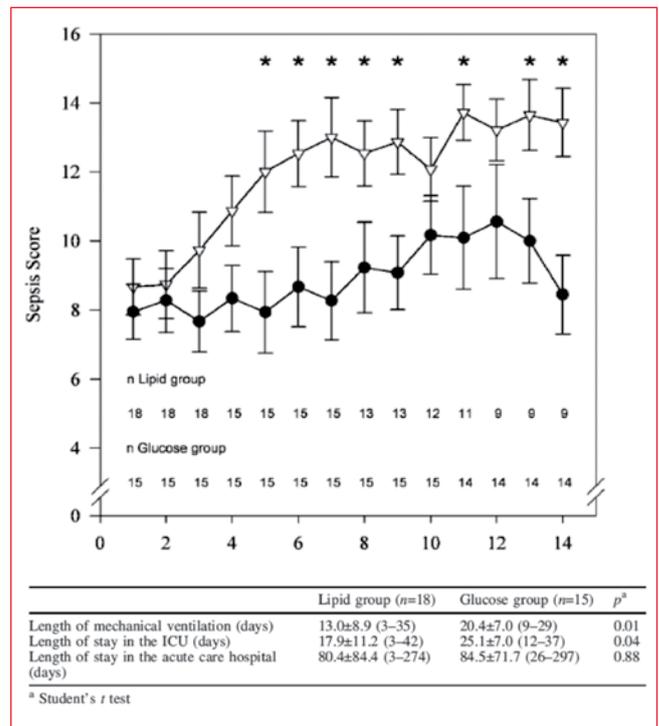
Seven smaller trials<sup>19,27-32</sup> in patients with ARDS, acquired immune deficiency syndrome, sepsis or undefined critically ill patients showed no significant difference in any important outcome parameter. One of two medium size trials (n = 72) showed less intra-abdominal infections and a trend towards reduced mortality in malnourished surgical patients<sup>33</sup> and the other one a significantly greater rise in retinol binding protein and nitrogen balance in septic patients.<sup>34</sup> However, as there are no harmful results,  $\omega$ -6/MCT lipid emulsions are still in use and may still be used in critically ill patients.

The next step in the development of intravenous lipid emulsions was a further reduction in the  $\omega$ -6 FA content to 20% and the completion with  $\omega$ -9 oleic acid from olive oil (OO) – which was introduced in 1996. The concern with this new emulsion was that the low content of  $\omega$ -6 FA might lead to a deficiency in essential FAs. This concern was partly addressed by a study in children reported by Munck<sup>35</sup> which documented that such a lipid emulsion could reduce the C20:3 content – a sign of essential acid deficiency – over 15 days as well as  $\omega$ -6 FA. There are also experimental studies that showed that  $\omega$ -9 FA or a lipid emulsion based on  $\omega$ -9 FA ( $\omega$ -9/ $\omega$ -6) did not inhibit lymphocyte proliferation,<sup>35,36</sup> did not reduce the expression of activation markers on granulocytes<sup>21,35,37</sup> and that it had almost no effect on the release of proinflammatory cytokines.<sup>38-41</sup> Thus, its effect on most immunologic parameters was almost neutral.

This  $\omega$ -9/ $\omega$ -6 emulsion was evaluated in six trials in preterm infants<sup>42-48</sup> and in three trials in paediatric patients.<sup>35,49-51</sup> All trials showed a good short and (some studies) long term tolerability and no signs of essential fatty acid deficiency. In adults, this  $\omega$ -9/ $\omega$ -6 emulsion was administered over three to six months in home parenteral nutrition.<sup>52-54</sup> Besides seven mostly smaller trials in various patients, this emulsion was also studied in one observational<sup>55</sup> and two controlled trials<sup>56,57</sup> in critically ill patients. The trial reported by Huschak<sup>57</sup> randomised 33 severe multiple trauma patients to either high dose  $\omega$ -9/ $\omega$ -6 (75% of non-protein energy) or normal dose  $\omega$ -6 FA (37% of non-protein energy). They found a significant shorter length of ventilation and a significantly shorter length of stay in the ICU in the high dose  $\omega$ -9/ $\omega$ -6 group (Figure 2). The trial reported by Garcia-de-Lorenzo<sup>56</sup> also found significantly less abnormalities in liver function in the  $\omega$ -9/ $\omega$ -6 group.

The next lipid emulsion – introduced in 1991 – was based on  $\omega$ -3 fatty acids derived from fish oil (FO). In many *in vitro* and *ex vivo* studies it could be shown that  $\omega$ -3 fatty acids had strong anti-inflammatory and immunosuppressive effects, already documented in various diseases such as rheumatoid arthritis, inflammatory bowel disease and asthma. Thus lipid emulsion inhibited lymphocyte proliferation,<sup>58-61</sup> decreased natural killer cell activity<sup>6,62</sup> as well as monocyte chemiluminescence, chemotaxis and adhesion to

Figure 2 : Huschak G et al.<sup>57</sup> Olive oil based nutrition in multiple trauma patients



endothelial cells<sup>63-65</sup> and it reduced the release of proinflammatory cytokines.<sup>66-68</sup>

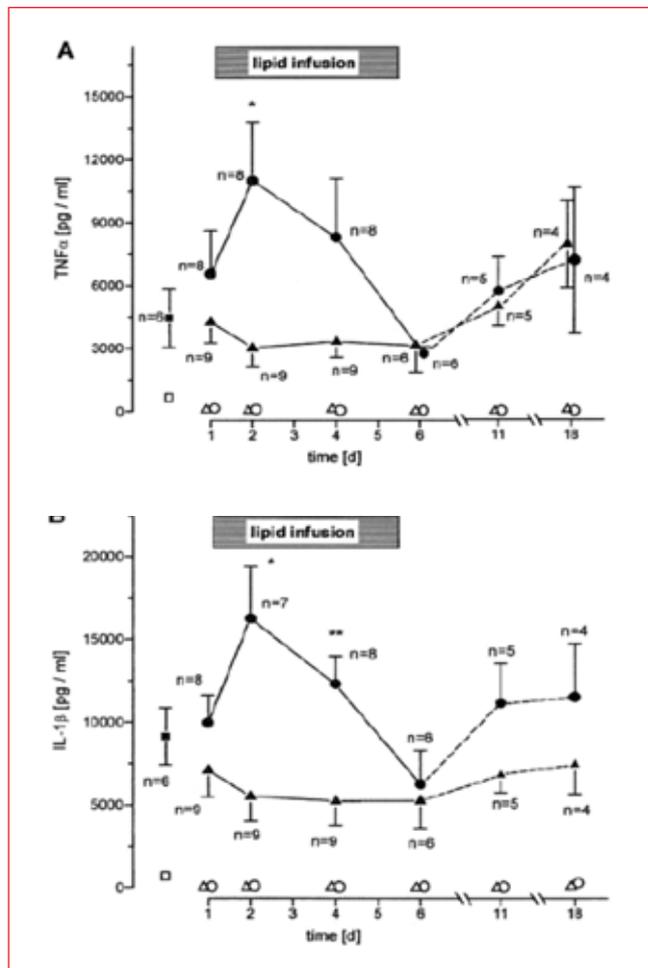
As the  $\omega$ -3 fatty acids emulsion ( $\omega$ -3) was never licensed for a standalone use but had to be administered in addition to another lipid emulsion, there are only few experimental clinical studies on the use of pure  $\omega$ -3.<sup>69-71</sup> In these, its main effect was to modulate cytokine and leukotriene release (Figure 3).

As far as the admixture of  $\omega$ -3 to other lipids is concerned, an important experiment was performed by Grimm et al<sup>72</sup> in 1994. They performed allogeneic heart transplantations in inbred rats and fed them postoperatively with different lipid emulsions: fish oil, soybean oil, safflower oil or a mixture of safflower with fish oil with a ratio of 2.1:1. The rejection time of the allograft was significantly prolonged with fish oil but also with safflower or soybean oil. However, the mixture of safflower with fish oil induced a rejection time which was comparable to control animals (Figure 4). This experiment was the base for the  $\omega$ -6: $\omega$ -3 ratio in the supplemental use of  $\omega$ -3 with  $\omega$ -6 and also for all lipid emulsions containing fish oil which were introduced after 2005.

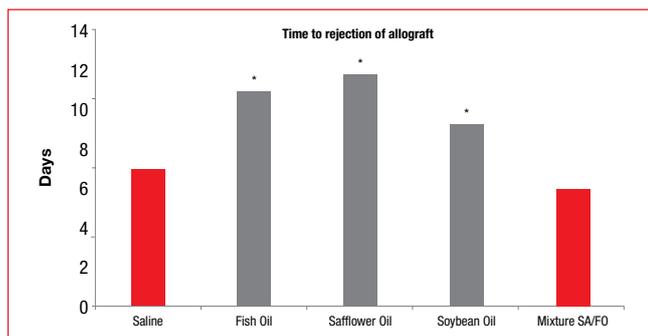
It might be concluded from these data that an admixture with the described  $\omega$ -6: $\omega$ -3 ratio is mostly immune-neutral. However, experimental data on the immunologic effect of these latest lipid emulsions are very rare. De Nardi et al<sup>73</sup> found yet a significant increase in macrophages that had engaged in phagocytosis with  $\omega$ -6/MCT and  $\omega$ -6/MCT/FO. However, there was no increase with  $\omega$ -6/MCT/FO/OO.

There are many clinical studies concerning the addition of fish oil to lipid emulsions. They can be divided into studies using the admixture of pure  $\omega$ -3 to other lipids, studies using  $\omega$ -6/MCT/FO, or studies using  $\omega$ -6/MCT/FO/OO.

**Figure 3 : Meyer K et al.<sup>67</sup> Parenteral nutrition with fish oil modulates cytokine response in patients with sepsis**



**Figure 4: Grimm H et al.<sup>72</sup> Immunoregulation by parenteral lipids (Figure was created using data from this publication)**



There are two observational<sup>74,75</sup> and nine randomised controlled<sup>76-83</sup> trials on the admixture of pure  $\omega$ -3 FA to other lipids. While none of the randomised trials showed a significant improvement in survival or length of stay, the observational trial reported by Heller et al<sup>74</sup> showed that FO had the most favorable effects on survival, infection rates and length of stay when administered in doses between 0.1 and 0.2 g/kg/day.

The eight studies which evaluated  $\omega$ -6/MCT/FO<sup>84-91</sup> also didn't show any significant effect on outcome except the trial reported by Wichmann et al<sup>87</sup> which showed a significant decrease in the length of stay (17.2 vs 21.9 days).

Of the three studies evaluating  $\omega$ -6/MCT/FO/OO,<sup>92-94</sup> the study reported by Mertes et al<sup>93</sup> showed a trend towards a reduced length of hospital stay (15.7 +/- 6.3 vs 17.8 +/- 13.2 days). Piper et al<sup>92</sup> compared  $\omega$ -6/MCT/FO/OO to  $\omega$ -9/ $\omega$ -6 and found at day two and five significantly lower levels of liver enzymes in the  $\omega$ -6/MCT/FO/OO group. However, the importance of this result is limited by the fact that the higher levels found with  $\omega$ -9/ $\omega$ -6 also were only slightly above normal range.

In conclusion,  $\omega$ -9/ $\omega$ -6 with its relative immune-neutrality is a lipid emulsion that can be used in most critically ill and non-critically ill patients. The addition of fish oil in the newer  $\omega$ -6/MCT/FO or  $\omega$ -6/MCT/FO/OO emulsions certainly has advantages compared to pure  $\omega$ -6 or  $\omega$ -6/MCT and these emulsions can also be used in most patients. However, the drawback is that the immune-modulatory potential of these emulsions is rather low as the fish oil content is too low to exert a greater effect. So there is still need for a lipid emulsion that could be advantageous in patients with real hyperinflammation.

### References

- Mertin J, Hunt R. Influence of polyunsaturated fatty acids on survival of skin allografts and tumor incidence in mice. *Proc Natl Acad Sci U S A* 1976;73(3):928-931.
- Wan JM, Teo TC, Babayan VK, Blackburn GL. Invited comment: lipids and the development of immune dysfunction and infection. *JPEN J Parent Ent Nutrition* 1988;12(6 Suppl):43S-52S.
- Utermohlen V, Tucker MA. Possible effects of dietary n-6 series polyunsaturated fatty acids on the development of immune dysfunction and infection. *Proc Nutr Soc* 1986;45(3):327-331.
- Calder PC, Sherrington EJ, Askanazi J, Newsholme EA. Inhibition of lymphocyte proliferation in vitro by two lipid emulsions with different fatty acid compositions. *Clin Nutr* 1994;13(2):69-74.
- Soyland E, Nenseter MS, Braathen L, Drevon CA. Very long chain n-3 and n-6 polyunsaturated fatty acids inhibit proliferation of human T-lymphocytes in vitro. *Eur J Clin Invest* 1993;23(2):112-121.
- Yaqoob P, Newsholme EA, Calder PC. Inhibition of natural killer cell activity by dietary lipids. *Immunol Lett* 1994;41(2-3):241-247.
- Monson JR, Sedman PC, Ramsden CW, Brennan TG, Guillou PJ. Total parenteral nutrition adversely influences tumour-directed cellular cytotoxic responses in patients with gastrointestinal cancer. *Eur J Surg Oncol* 1988;14(5):935-943.
- Cury-Boaventura MF, Gorjao R, de Lima TM et al. Toxicity of a soybean oil emulsion on human lymphocytes and neutrophils. *JPEN J Parenter Enteral Nutr* 2006;30:115-123.
- Fischer GW, Hunter KW, Wilson SR, Mease AD. Diminished bacterial defences with intralipid. *Lancet* 1980;2:819-820.
- Nordenstrom J, Jarstrand C, Wiernik A. Decreased chemotactic and random migration of leukocytes during Intralipid infusion. *Am J Clin Nutr* 1979;32:2416-2422.
- Katz S, Plaisir BR, Folkering WJ, Grosfeld JL. Intralipid adversely affects reticuloendothelial bacterial clearance. *J Pediatr Surg* 1991;26(8):921-924.
- Seidner DL, Mascioli EA, Istfan NW et al. Effects of long-chain triglyceride emulsions on reticuloendothelial system function in humans. *JPEN J Parenter Enteral Nutr* 1989;13:614-619.
- Li X, Ying J, Zeng S et al. The effects of a short-term long-chain-triglyceride infusion on the postoperative immune function of pediatric patients receiving a gastrointestinal surgical procedure. *JPEN J Parenter Enteral Nutr* 2008;32(1):72-77.
- Dionigi P, Dionigi R, Prati U, et al. Effect of Intralipid(Reg,trademark) on some immunological parameters and leukocyte functions in patients with esophageal and gastric cancer. *Clin Nutrition* 1985;4(4):229-234.
- Ota DM, Jessup JM, Babcock GF et al. Immune function during intravenous administration of a soybean oil emulsion. *JPEN J Parenter Enteral Nutr* 1985;9(1):23-27.
- Lenness P, Bruemmer BA, Bowden RA, Gooley T, Aker SN, Mattson D. Intravenous lipid dose and incidence of bacteremia and fungemia in patients undergoing bone marrow transplantation. *Am J Clin Nutr* 1998;67(5):927-933.
- Monson JR, Ramsden CW, MacFie J, Brennan TG, Guillou PJ. Immunorestorative effect of lipid emulsions during total parenteral nutrition. *Br J Surg* 1986;73:843-846.
- Battistella FD, Widergren JT, Anderson JT, Siepler JK, Weber JC, MacColl K. A prospective, randomized trial of intravenous fat emulsion administration in trauma victims requiring total parenteral nutrition. *J Trauma* 1997;43(1):52-58.
- Gelas P, Cotte L, Poitevin-Later F et al. Effect of parenteral medium- and long-chain triglycerides on lymphocytes subpopulations and functions in patients with acquired immunodeficiency syndrome: a prospective study. *JPEN J Parent Ent Nutrition* 1998;22(2):67-71.
- Sedman PC, Ramsden CW, Brennan TG, Guillou PJ. Pharmacological concentrations of lipid emulsions inhibit interleukin-2-dependent lymphocyte responses in vitro. *JPEN J Parenter Enteral Nutr* 1990;14(1):12-17.
- Versleijen M, Roelofs H, Preijers F, Roos D, Wanten G. Parenteral lipids modulate leukocyte phenotypes in whole blood, depending on their fatty acid composition. *Clin Nutr* 2005;24(5):822-829.
- Wanten GJ, Roos D, Naber AH. Effects of structurally different lipid emulsions on human neutrophil migration. *Clin Nutr* 2000;19:327-331.
- Hamawy KJ, Moldawer LL, Georgieff M et al. The Henry M. Vars Award. The effect of lipid emulsions on reticuloendothelial system function in the injured animal. *JPEN J Parent Ent Nutrition* 1985;9(5):559-565.
- Sobrado J, Moldawer LL, Pomposelli JJ et al. Lipid emulsions and reticuloendothelial system function in healthy and burned guinea pigs. *Am J Clin Nutr* 1985;42:855-863.
- Wanten GJ, Netea MG, Naber TH et al. Parenteral administration of medium- but not long-chain lipid emulsions may increase the risk for infections by *Candida albicans*. *Infect Immun* 2002;70(11):6471-6474.
- Wanten GJ, Curfs JH, Meis JF, Naber AH. Phagocytosis and killing of *Candida albicans* by human

- neutrophils after exposure to structurally different lipid emulsions. *JPEN J Parenter Enteral Nutr* 2001;25(1):9–13.
27. Iovinelli G, Marinangeli F, Ciccone A et al. Parenteral nutrition in ventilated patients with chronic obstructive pulmonary disease: long chain vs medium chain triglycerides. *Minerva Anestesiologica* 2007;73(1–2):65–76.
  28. Lekka ME, Lioikatis S, Nathanael C, Galani V, Nakos G. The impact of intravenous fat emulsion administration in acute lung injury. *Am J Respir Crit Care Med* 2004;169(5):638–644.
  29. Mascians JR, Iglesia R, Bermejo B, Pico M, Rodríguez-Roisin R, Planas M. Gas exchange and pulmonary haemodynamic responses to fat emulsions in acute respiratory distress syndrome. *Int Care Med* 1998;24(9):918–923.
  30. Ball MJ. Parenteral nutrition in the critically ill: use of a medium chain triglyceride emulsion. *Intensive Care Med* 1993;19(2):89–95.
  31. Radermacher P, Santak B, Strobach H, Schror K, Tarnow J. Fat emulsions containing medium chain triglycerides in patients with sepsis syndrome: effects on pulmonary hemodynamics and gas exchange. *Int Care Med* 1992;18(4):231–234.
  32. Ball MJ, White K. Comparison of medium and long chain triglyceride metabolism in intensive care patients on parenteral nutrition. *Intensive Care Med* 1989;15(4):250–254.
  33. Grau T, Ruiz de Adana JC, Zubillaga S, Fuente S, Giron C. [Randomized study of two different fat emulsions in total parenteral nutrition of malnourished surgical patients; effect of infectious morbidity and mortality]. *Nutr Hosp* 2003;18(3):159–166.
  34. Garnacho-Montero J, Ortiz-Leyba C, Jimenez-Jimenez FJ et al. Clinical and metabolic effects of two lipid emulsions on the parenteral nutrition of septic patients. *Nutrition* 2002;18(2):134–138.
  35. Munck A, Navarro J. Tolerability and efficacy of ClinOleic® lipid emulsion in children reliant exclusively on parenteral nutrition. *Nutr Clin Métabol* 1996;10:455–475.
  36. Granato D, Blum S, Rossle C, Le Boucher J, Malnoe A, Dutot G. Effects of parenteral lipid emulsions with different fatty acid composition on immune cell functions *in vitro*. *JPEN J Parent Ent Nutrition* 2000;24(2):113–118.
  37. Buenestado A, Cortijo J, Sanz MJ et al. Olive oil-based lipid emulsion's neutral effects on neutrophil functions and leukocyte-endothelial cell interactions. *JPEN J Parenter Enteral Nutr* 2006;30(4):286–296.
  38. Leite MS, Pacheco P, Gomes RN et al. Mechanisms of increased survival after lipopolysaccharide-induced endotoxic shock in mice consuming olive oil-enriched diet. *Shock* 2005;23(2):173–178.
  39. Reimund JM, Scheer O, Muller CD, Pinna G, Duclos B, Baumann R. *In vitro* modulation of inflammatory cytokine production by three lipid emulsions with different fatty acid compositions. *Clin Nutr* 2004;23(6):1324–1332.
  40. Sadeghi S, Wallace FA, Calder PC. Dietary lipids modify the cytokine response to bacterial lipopolysaccharide in mice. *Immunology* 1999;96(3):404–410.
  41. Yaqoob P, Calder P. Effects of dietary lipid manipulation upon inflammatory mediator production by murine macrophages. *Cell Immunol* 1995;163(1):120–128.
  42. Deshpande GC, Simmer K, Mori T, Croft K. Parenteral lipid emulsions based on olive oil compared with soybean oil in preterm (<28 weeks' gestation) neonates: a randomised controlled trial. *J Pediatr Gastroenterol Nutr* 2009;49:619–625.
  43. Roggero P, Mosca F, Gianni ML et al. F2-isoprostanes and total radical-trapping antioxidant potential in preterm infants receiving parenteral lipid emulsions. *Nutrition* 2009.
  44. Gawecka A, Michalkiewicz J, Kornacka MK, Luckiewicz B, Kubiszewska I. Immunologic properties differ in preterm infants fed olive oil vs soy-based lipid emulsions during parenteral nutrition. *JPEN J Parenter Enteral Nutr* 2008;32(4):448–453.
  45. Webb AN, Hardy P, Peterkin M et al. Tolerability and safety of olive oil-based lipid emulsion in critically ill neonates: a blinded randomized trial. *Nutrition* 2008;24(11–12):1057–1064.
  46. van Kempen AA, van der Crabben SN, Ackermans MT, Ender E, Kok JH, Sauerwein HP. Stimulation of gluconeogenesis by intravenous lipids in preterm infants: response depends on fatty acid profile. *Am J Physiol Endocrinol Metab* 2006;290(4):E723–E730.
  47. Pitkänen OM, Luukkainen P, Andersson S. Attenuated lipid peroxidation in preterm infants during subsequent doses of intravenous lipids. *Biol Neonate* 2004;85(3):184–187.
  48. Göbel Y, Koletzko B, Bohles HJ et al. Parenteral fat emulsions based on olive and soybean oils: a randomized clinical trial in preterm infants. *J Pediatr Gastroenterol Nutr* 2003;37(2):161–167.
  49. Hartman C, Ben-Artzi E, Berkowitz D et al. Olive oil-based intravenous lipid emulsion in pediatric patients undergoing bone marrow transplantation: A short-term prospective controlled trial. *Clin Nutr* 2009.
  50. Goulet O, de Pottier S, Antebi H et al. Long-term efficacy and safety of a new olive oil-based intravenous fat emulsion in pediatric patients: a double-blind randomized study. *Am J Clin Nutr* 1999;70(3):338–345.
  51. Antébi H, Zimmerman L, Bourcier C et al. Peroxydation *in vitro* et effet de l'administration en nutrition parentérale totale d'une émulsion lipidique à base d'huile d'olive sur la peroxydabilité des lipoprotéines de basse densité chez l'enfant. *Nutr Clin Métabol* 1996;10:415–435.
  52. Reimund JM, Rahmi G, Escalin G et al. Efficacy and safety of an olive oil-based intravenous fat emulsion in adult patients on home parenteral nutrition. *Aliment Pharmacol Ther* 2005;21(4):445–454.
  53. Vahedi K, Atlán P, Joly F et al. A 3-month double-blind randomised study comparing an olive oil- with a soybean oil-based intravenous lipid emulsion in home parenteral nutrition patients. *Br J Nutr* 2005;94(6):909–916.
  54. Thomas-Gibson S, Jawhari A, Atlán P, Brun AL, Farthing M, Forbes A. Safe and efficacious prolonged use of an olive oil-based lipid emulsion (ClinOleic) in chronic intestinal failure. *Clin Nutr* 2004;23(4):697–703.
  55. Mateu-de Antonio J, Grau S, Luque S, Marin-Casino M, Albert I, Ribes E. Comparative effects of olive oil-based and soybean oil-based emulsions on infection rate and leukocyte count in critically ill patients receiving parenteral nutrition. *Br J Nutr* 2008;99(4):846–854.
  56. Garcia-de-Lorenzo A, Denia R, Atlán P et al. Parenteral nutrition providing a restricted amount of linoleic acid in severely burned patients: a randomised double-blind study of an olive oil-based lipid emulsion v. medium/long-chain triacylglycerols. *Br J Nutr* 2005;94(2):221–230.
  57. Huschak G, Zur NK, Hoell T, Riemann D, Mast H, Stuttmann R. Olive oil based nutrition in multiple trauma patients: a pilot study. *Intensive Care Med* 2005;31(9):1202–1208.
  58. Endres S, Meydani SN, Ghorbani R, Schindler R, Dinarello CA. Dietary supplementation with n-3 fatty acids suppresses interleukin-2 production and mononuclear cell proliferation. *J Leukoc Biol* 1993;54:599–603.
  59. Kelley DS, Branch LB, Love JE, Taylor PC, Rivera YM, Iacono JM. Dietary alpha-linolenic acid and immunocompetence in humans. *Am J Clin Nutr* 1991;53:40–46.
  60. Meydani SN, Endres S, Woods MM et al. Oral (n-3) fatty acid supplementation suppresses cytokine production and lymphocyte proliferation: comparison between young and older women. *J Nutr* 1991;121:547–555.
  61. Molvig J, Pociot F, Worsaae H et al. Dietary supplementation with omega-3-polyunsaturated fatty acids decreases mononuclear cell proliferation and interleukin-1 beta content but not monokine secretion in healthy and insulin-dependent diabetic individuals. *Scand J Immunol* 1991;34:399–410.
  62. Kelley DS, Taylor PC, Nelson GJ et al. Docosahexaenoic acid ingestion inhibits natural killer cell activity and production of inflammatory mediators in young healthy men. *Lipids* 1999;34:317–324.
  63. Mayer K, Merfels M, Muhly-Reinholz M et al. Omega-3 fatty acids suppress monocyte adhesion to human endothelial cells: role of endothelial PAF generation. *Am J Physiol Heart Circ Physiol* 2002;283(2):H811–H818.
  64. Fisher M, Levine PH, Weiner BH et al. Dietary n-3 fatty acid supplementation reduces superoxide production and chemiluminescence in a monocyte-enriched preparation of leukocytes. *Am J Clin Nutr* 1990;51:804–808.
  65. Schmidt EB, Pedersen JO, Ekelund S, Grunnet N, Jersild C, Dyerberg J. Cod liver oil inhibits neutrophil and monocyte chemotaxis in healthy males. *Atherosclerosis* 1989;77:53–57.
  66. Pluess TT, Hays D, Berger MM et al. Intravenous fish oil blunts the physiological response to endotoxin in healthy subjects. *Intensive Care Med* 2007;33(5):789–797.
  67. Mayer K, Meyer S, Reinholz-Muhly M et al. Short-time infusion of fish oil-based lipid emulsions, approved for parenteral nutrition, reduces monocyte proinflammatory cytokine generation and adhesive interaction with endothelium in humans. *J Immunol* 2003;171(9):4837–4843.
  68. Endres S, Ghorbani R, Kelley VE et al. The effect of dietary supplementation with n-3 polyunsaturated fatty acids on the synthesis of interleukin-1 and tumor necrosis factor by mononuclear cells. *N Engl J Med* 1989;320:265–271.
  69. Mayer K, Fegbeutel C, Hattar K et al. Omega-3 vs. omega-6 lipid emulsions exert differential influence on neutrophils in septic shock patients: impact on plasma fatty acids and lipid mediator generation. *Intensive Care Med* 2003;29(9):1472–1481.
  70. Mayer K, Gokorsch S, Fegbeutel C et al. Parenteral nutrition with fish oil modulates cytokine response in patients with sepsis. *Am J Respir Crit Care Med* 2003;167(10):1321–1328.
  71. Weiss G, Meyer F, Matthies B, Pross M, Koenig W, Lippert H. Immunomodulation by perioperative administration of n-3 fatty acids. *Br J Nutr* 2002;87 Suppl 1:S89–S94.
  72. Grimm H, Tibell A, Norrind B, Blecher C, Wilker S, Schwemmler K. Immunoregulation by parenteral lipids: impact of the n-3 to n-6 fatty acid ratio. *JPEN J Parent Ent Nutrition* 1994;18(5):417–421.
  73. De Nardi L, Bellinati-Pires R, Torrinhas RS, Bacchi CE, Arias V, Walitzberg DL. Effect of fish oil containing parenteral lipid emulsions on neutrophil chemotaxis and resident-macrophages' phagocytosis in rats. *Clin Nutr* 2008;27(2):283–288.
  74. Heller AR, Rossler S, Litz RJ et al. Omega-3 fatty acids improve the diagnosis-related clinical outcome. *Crit Care Med* 2006;34(4):972–979.
  75. Teskes E, Reuter C, Stehle P, Boeden G. Perioperative administration of parenteral fish oil supplements in a routine clinical setting improves patient outcome after major abdominal surgery. *Clin Nutr* 2004;23(3):325–330.
  76. Liang B, Wang S, Ye YJ et al. Impact of postoperative omega-3 fatty acid-supplemented parenteral nutrition on clinical outcomes and immunomodulations in colorectal cancer patients. *World J Gastroenterol* 2008;14(15):2434–2439.
  77. Wang X, Li W, Li N, Li J. {omega}-3 Fatty Acids-Supplemented Parenteral Nutrition Decreases Hyperinflammatory Response and Attenuates Systemic Disease Sequelae in Severe Acute Pancreatitis: A Randomized and Controlled Study. *JPEN J Parenter Enteral Nutr* 2008;32(3):236–241.
  78. Tappy L, Berger MM, Schwarz JM et al. Metabolic effects of parenteral nutrition enriched with n-3 polyunsaturated fatty acids in critically ill patients. *Clin Nutr* 2006;25(4):588–595.
  79. Heller AR, Rossel T, Gottschlich B et al. Omega-3 fatty acids improve liver and pancreas function in postoperative cancer patients. *Int J Cancer* 2004;111(4):611–616.
  80. Heller AR, Fischer S, Rossel T et al. Impact of n-3 fatty acid supplemented parenteral nutrition on haemostasis patterns after major abdominal surgery. *Br J Nutr* 2002;87 Suppl 1:S95–101.
  81. Schauder P, Rohn U, Schafer G, Korff G, Schenk HD. Impact of fish oil enriched total parenteral nutrition on DNA synthesis, cytokine release and receptor expression by lymphocytes in the postoperative period. *Br J Nutr* 2002;87 Suppl 1:S103–S110.
  82. Abribat T, Nedelec B, Jobin N, Garrel DR. Decreased serum insulin-like growth factor-I in burn patients: relationship with serum insulin-like growth factor binding protein-3 proteolysis and the influence of lipid composition in nutritional support. *Crit Care Med* 2000;28(7):2366–2372.
  83. Morlion BJ, Torwesten E, Lessire H et al. The effect of parenteral fish oil on leukocyte membrane fatty acid composition and leukotriene-synthesizing capacity in patients with postoperative trauma. *Metabolism* 1996;45(10):1208–1213.
  84. Berger MM, Tappy L, Revelly JP et al. Fish oil after abdominal aorta aneurysm surgery. *Eur J Clin Nutr* 2008;62(9):1116–1122.
  85. Friesecke S, Lotze C, Kohler J, Heinrich A, Felix SB, Abel P. Fish oil supplementation in the parenteral nutrition of critically ill medical patients: a randomised controlled trial. *Intensive Care Med* 2008;34(8):1411–1420.
  86. Senkal M, Geier B, Hannemann M et al. Supplementation of omega-3 fatty acids in parenteral nutrition beneficially alters phospholipid fatty acid pattern. *JPEN J Parenter Enteral Nutr* 2007;31(1):12–17.
  87. Wichmann MW, Thul P, Czarnetzki HD, Morlion BJ, Kemen M, Jauch KW. Evaluation of clinical safety and beneficial effects of a fish oil containing lipid emulsion (Lipolip, MLF541): Data from a prospective, randomized, multicenter trial. *Crit Care Med* 2007;35:700–706.
  88. Koller M, Senkal M, Kemen M, König W, Zumbel V, Muhr G. Impact of omega-3 fatty acid enriched TPN on leukotriene synthesis by leukocytes after major surgery. *Clin Nutr* 2003;22(1):59–64.
  89. Linseisen J, Hoffmann J, Lienhard S, Jauch KW, Wolfram G. Antioxidant status of surgical patients receiving TPN with an omega-3- fatty acid-containing lipid emulsion supplemented with alpha-tocopherol. *Clin Nutr* 2000;19(3):177–184.
  90. Wachtler P, König W, Senkal M, Kemen M, Koller M. Influence of a total parenteral nutrition enriched with omega-3 fatty acids on leukotriene synthesis of peripheral leukocytes and systemic cytokine levels in patients with major surgery. *J Trauma* 1997;42(2):191–198.
  91. Barbosa VM, Miles EA, Calhau C, Lafuente E, Calder PC. Effects of a fish oil containing lipid emulsion on plasma phospholipid fatty acids, inflammatory markers, and clinical outcomes in septic patients: a randomized, controlled clinical trial. *Crit Care* 2010;14:R5.
  92. Piper SN, Schade I, Beschmann RB, Maleck WH, Boldt J, Rohm KD. Hepatocellular integrity after parenteral nutrition: comparison of a fish-oil-containing lipid emulsion with an olive-soybean oil-based lipid emulsion. *Eur J Anaesthesiol* 2009;26:1076–1082.
  93. Mertes N, Grimm H, Furst P, Stehle P. Safety and efficacy of a new parenteral lipid emulsion (SMOFlipid) in surgical patients: a randomized, double-blind, multicenter study. *Ann Nutr Metab* 2006;50(3):253–259.
  94. Antebi H, Mansoor O, Ferrier C et al. Liver function and plasma antioxidant status in intensive care unit patients requiring total parenteral nutrition: comparison of 2 fat emulsions. *JPEN J Parenter Enteral Nutr* 2004;28(3):142–148.