Implementation of a new ‘community’ laboratory CD4 service in a rural health district in South Africa extends laboratory services and substantially improves local reporting turnaround time

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Background. The CD4 integrated service delivery model (ITSDM) provides for reasonable access to pathology services across South Africa (SA) by offering three new service tiers that extend services into remote, under-serviced areas. ITSDM identified Pixley ka Seme as such an under-serviced district.

Objective. To address the poor service delivery in this area, a new ITSDM community (tier 3) laboratory was established in De Aar, SA. Laboratory performance and turnaround time (TAT) were monitored post implementation to assess the impact on local service delivery.

Methods. Using the National Health Laboratory Service Corporate Data Warehouse, CD4 data were extracted for the period April 2012 - July 2013 (n=11 964). Total mean TAT (in hours) was calculated and pre-analytical and analytical components assessed. Ongoing testing volumes, as well as external quality assessment performance across ten trials, were used to indicate post-implementation success. Data were analysed using Stata 12.

Results. Prior to the implementation of CD4 testing at De Aar, the total mean TAT was 20.5 hours. This fell to 8.2 hours post implementation, predominantly as a result of a lower pre-analytical mean TAT reducing from a mean of 18.9 to 1.8 hours. The analytical testing TAT remained unchanged after implementation and monthly test volumes increased by up to 20%. External quality assessment indicated adequate performance. Although subjective, questionnaires sent to facilities reported improved service delivery.

Conclusion. Establishing CD4 testing in a remote community laboratory substantially reduces overall TAT. Additional community CD4 laboratories should be established in under-serviced areas, especially where laboratory infrastructure is already in place.


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provides some basic pathology services across a service precinct of 103 410 km² and eight municipalities. The local population of ~186 000 people attend multiple health facilities within a radius of up to 260 km, including ten local hospitals (hospital wards) and 34 PHC clinics. More than 70% of the population is under the age of 40 years, and the district has an HIV prevalence of 18.4% (2012 statistics). A tier 3/community laboratory CD4 service was piloted at the existing De Aar NHLS laboratory to address the historically poor CD4 TAT and gaps in service. The impact of the newly implemented tier 3/community CD4 laboratory service in De Aar was assessed by reviewing the TAT before and after the implementation. Success of the implementation was measured through review of ongoing monthly CD4 test volumes and external quality assessment performance of the De Aar laboratory.

Methods
CD4 testing was implemented and training given at the De Aar laboratory in December 2012 using the Beckman Coulter Epics XL™ flow cytometry platform and the PanLeucogating method. Using Corporate Data Warehouse (CDW), CD4 sample level data were extracted for the period April 2012 - July 2013 (N=11 964) for the wider Pixley ka Seme health area. This included pre-implementation data of CD4 workload referred to regional CD4 testing laboratories for testing (Kimberly or Pelonomi) as well as data from the De Aar laboratory itself post implementation (sample volume data were extended to December 2014). The specimen data extract included the laboratory information system (LIS) episode number (which provided evidence of volumes of tests), province, health district and the health facility location name and code from which the sample was sent. Data that were collected with a listed LIS 'tested date' between 1 April and 30 November 2012 were categorised as 'pre-implementation'. Data collected after January 2013 were categorised as 'post-implementation'. The reporting measures included CD4 monthly test volumes and mean TATs. To establish TAT, the following LIS date and time fields were used, including: (i) ‘registered date’, when samples are registered (order entry) at the referring hub or laboratory; (ii) ‘test registered date’ describing when samples are registered/received at the CD4 testing laboratory; (iii) ‘tested date’, when the CD4 Epics XL™ CD4 result data are downloaded into the LIS; and lastly, (iv) ‘reviewed date’, when a medical technologist authorises the CD4 results for printing (by SMS (text message)) at the original referring site. Using the LIS date and time fields, four distinct TAT measures were identified for this analysis: (i) ‘PRE-LAB TAT’ (pre-analytical component), test registered date minus registered date (interlaboratory or processing hub referral time); (ii) ‘IN-LAB TAT’, tested date minus test registered date (analytical processing component); (iii) ‘PRE-LAB TAT’ (pre-analytical component), test registered date minus registered date (interlaboratory or processing hub referral time); (ii) ‘IN-LAB TAT’, tested date minus test registered date (analytical processing component).
(iii) ‘RVW-TAT’, reviewed date minus tested date (time to authorised results following analysis); and lastly (iv) ‘TOTAL-TAT’, reviewed date minus registered date. Anomalies, due to a computer internal data and time setting for the decentralised Disa*Lab LIS used previously, that were noted in the LIS date and time fields were excluded (e.g. a reviewed date in the year 1800 (1800/01/01 00:01). This affected only 0.3% (36/11 964) of the total data set. All data were analysed using Microsoft Excel and Stata 12.

External quality assessment (EQA) performance of the De Aar laboratory on the NHLS CD4 Proficiency Testing (PT)/African Regional External Quality Assessment (AFREQAS) programme was obtained from the De Aar laboratory from trial 19 (May 2013) through to trial 30. Each AFREQAS trial consists of two proficiency material samples, i.e. one normal (CD4 count >500 cells/µL) and one low (CD4 count <200 cells/µL) sample. Performance of the laboratory is measured against the consensus pooled mean result of each trial, and is regarded as adequate if the CD4 result reported by the laboratory falls within 2 standard deviations (SDs) of the consensus pooled trimmed-mean CD4 result.

Results

The geographical relationship of 44 referring health facilities and the De Aar laboratory in the Pixley ka Seme district to the pre-implementation referral CD4 labs in the Free State (Pelonomi) or Kimberly (North West) is shown in Fig. 1. This figure also gives information about existing CD4 service coverage in SA, showing paucity of local services in the Pixley ka Seme district, in contrast to the relatively well-serviced areas elsewhere in the country. Fig. 2 reveals the impact on TAT of longer (shorter) distances required to refer samples for testing elsewhere; the monthly mean TOTAL-TAT for Pixley ka Seme before implementation ranged from 12 to 38 hours (the majority of samples were referred from the De Aar laboratory to the Kimberley laboratory for testing). A marked difference in mean TOTAL-TAT was noted, reducing from a median of 20.45 hours before implementation to 8.17 hours after December 2012 (Fig. 2). Following a 2-month settling-in period, by March 2013 (mean TOTAL-TAT of 16 hours was recorded), the TOTAL-TAT steadily decreased to a mean of 12 hours and stabilised at 5 hours or less by May 2013. This trend has continued beyond the TAT data extraction period (data not shown).

To assess what component of TAT contributed to TOTAL-TAT, the various components of TAT were assessed including PRE-LAB (pre-analytical), IN-LAB and RVW-TAT (both analytical), before (pre-) and after (post-) implementation (Fig. 2). The pre-analytical component of the TOTAL-TAT was noted to contribute the largest proportion of TOTAL-TAT in the pre-implementation time period. During these pre-implementation months, April - November 2012, pre-analytical TAT showed marked daily variability (with a range of TAT from 4 hours in April 2012 extending to 45 hours in October 2012). After the De Aar CD4 lab was established, the mean PRE-LAB decreased from an average of 18.99 hours to 1.82 hours, reducing the mean monthly TOTAL-TAT substantially. By June 2013, the mean pre-analytical TAT had stabilised at ~2 hours. Evidence of an initial learning curve was revealed during the first 2 months, i.e. January
and February 2013, where an IN-LAB TAT of 11 and 13 hours was noted respectively (highlighted in Fig. 2). However, the IN-LAB/analytical TAT subsequently decreased to that expected of a typical CD4 laboratory processing time as staff became more familiar and integrated the new CD4 testing into their existing workflow (attributable to the adherence to the standardised NHLS CD4 testing procedures that are used irrespective of where testing is performed across the NHLS CD4 network). Further analysis of the TOTAL-TAT data revealed that before implementation of the De Aar CD4 site, 54% of CD4 samples were reported within a TOTAL-TAT of ≤12 hours. After implementation however, 81% of samples had a TOTAL-TAT of ≤12 hours (Fig. 2). In a detailed subanalysis, CD4 sample results with a TOTAL-TAT between 13 and 24 hours decreased from 26% to 13%; those with a TOTAL-TAT >24 hours, decreased by 150%, from 19% to 6% of all samples tested. Overall, the percentage of samples with a ≤24 hours TOTAL-TAT increased from 81% before implementation to 94% post implementation.

Details of month-to-month variation of workload volume post implementation can be seen in Fig. 3, extending to December 2014. Pre-implementation volumes were divided into 'referred to and tested at Kimberley' (mean of 571 samples/month) and ‘referred by De Aar and tested in Kimberley’ (mean 112 samples per month). After CD4 testing started at De Aar in December 2012, some samples were still referred through De Aar to be tested in Kimberley (~125 per month). This was attributed to periods of downtime on the instrument (owing to long distances, reaction time of engineers to get instruments operational is prolonged) and/or availability of staff to operate the system. However, the majority of samples from the district are now tested at the De Aar laboratory (monthly mean of 358 samples). Some samples are still referred directly to Kimberley at a mean rate of 287 per month and reflect NHLS business decisions to optimally refer testing from sites that lie far north in Pixley ka Seme to the nearest testing facility (in this instance Kimberley).

**External quality assessment performance**

During the post-implementation period of this study, ten trials (20 EQA samples) were tested by the De Aar laboratory. Across each trial, proficiency panel results (CD4 normal and CD4 low, as CD4 count and CD4% of lymphocytes (CD4%L)) were recorded within the required 2 SDI range (Fig. 4, shaded area; −2 - +2, with zero as the ideal target). There was an initial outlier EQA result on the normal CD4%L (intervention revealed a transcription error due to inexperience of staff filling in the EQAS submission form). The average SDI of the De Aar CD4 laboratory across all trials was −0.21, confirming that performance was well within the expected range. However, SDI results from Trial 23 fell outside the expected range for both absolute count and CD4%L (intervention and corrective action revealed that new staff had been appointed without proper EQAS training). The laboratory has sustained acceptable performance on the scheme for the remaining reported trials (Fig. 4).

**Discussion**

CD4 testing has been used for establishing immune suppression and disease progression in HIV-positive patients, enabling identification of those patients who are eligible for antiretroviral therapy and/or treatment for opportunistic infection. Current SA treatment guidelines\[9\] require that newly identified HIV-positive patients return to the clinic facility within 7 days of testing HIV-positive\[12\] to

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**Fig. 3.** Total volumes of CD4 tests performed per month before (pre-) and after (post-) CD4 laboratory implementation at De Aar. Volumes of samples ‘referred to and tested at Kimberley’ (‘RT Kimberley’), are shown in contrast to samples ‘referred to and tested at De Aar’ (‘RT De Aar’). A small fraction of samples referred to De Aar for testing is still tested at Kimberley (‘R De Aar T Kimberley’/‘RT Kimberley’).
receive the results of the required pathology work-up, including CD4 results outlined in the treatment guidelines. Since the commencement of the South African HIV/AIDS Comprehensive Care, Management and Treatment programme, laboratory-based CD4 testing has been the mainstay of the CD4 services provided. Locally, the NHLS provides access to the majority of these CD4 results within a 24-hour TAT to facilitate algorithm management. In the rural areas of the Northern Cape Province, the closest tier 4/regional and tier 5/metro centralised laboratories are too far (up to 300 - 400 km) away to ensure reliable CD4 TATs.

Several service delivery options were considered for Pixley ka Seme area that would improve TAT, while taking into account the vast distances between facilities in the district and proximity to the closest testing facilities. The first service option considered CD4 testing on a mobile unit, with various scenarios proposed. These included a once-per-month service option (costing ZAR125.47/test), a fortnightly option (ZAR201.22/test/visit), a weekly option (ZAR352.73) as well as a daily service delivery option (ZAR1 564.75/test). This approach was abandoned because of the prohibitively high, unaffordable cost per test required to maintain reasonable access to services on at least 4 days a week.

Implementation of widespread POC CD4 testing services in all 44 referring facilities was also considered as the second option, potentially providing immediate patient access to results and purporting the added advantage of improving the number of patients enrolled into care. Aside from the logistical and other problems of widespread POC technology implementation, overall voluntary counselling and testing process compliance has been reportedly poor, with specific areas of concern including specimen collection methodology and availability of equipment, e.g. gloves, etc., to perform the test (procurement challenges), poor or misuse of available consumables, poor adherence to stipulated incubation times before reading HIV rapid test results, lack of staff training and inconsistent or absent quality assurance practices. This approach was also abandoned in the NHLS because of the substantively higher programmatic costs that could be incurred by implementing POC across multiple clinics; a CD4 test performed at the POC was shown to cost 5 - 7 times more than providing a CD4 test in a laboratory.

Fig. 4. Performance on EQA after tier 3 CD4 service implementation at De Aar. Radial performance plots show laboratory performance on trials 19 through 30. A1 and A2 represent data from the normal CD4 count proficiency material while B1 and B2 depict results from low CD4 count EQA material. The acceptable range of −2 - +2 SDI is shaded with a solid line indicating the ideal of zero. An initial outlier on trial 19 was shown to be a transcription error. Another outlier was noted for trial 23 (both parameters). Intervention and corrective action ensured that future performance returned to expected.
The third service delivery of providing a local small community laboratory was provided for in the ITSDM. This study confirms that the establishment of an ITSDM tier 3 laboratory in a remote, under-serviced area of SA with historically poor TAT can lead to a dramatic improvement of local service delivery. There was a notable reduction in the pre-analytical TAT after CD4 testing was started in the De Aar laboratory, from 18 hours when samples were referred in centrally for testing to the Kimberley laboratory, down to a mean of 1.82 hours post implementation. Although anecdotal, the responses from local clinics were favourable; a moderate increase in the number of CD4 tests requested post implementation (see Fig. 3) attested to this. Despite an initial hiccup of increased analytical/IN-LAB TAT attributable to learning curve, IN-LAB TAT fell to expected levels as staff became better acquainted with NHLS standardised operating procedures. This is an encouraging aspect, and together with the satisfactory EQA review of the performance of the site, suggests that any small laboratory, with few staff members, is likely to cope well with an add-on tier 3 service. Although some CD4 testing was still referred to the larger metropolitan tier 4 lab at Kimberley after implementation, because referring clinic sites were in closer proximity to this centre (see Fig. 3), a moderately increased local workload (Fig. 3) was noted post implementation at De Aar. This is also an important outcome. If smaller laboratories are capable of an additional modest workload, without major implementation, overhead and staffing costs, this could potentially save the country millions of rands while extending the footprint of laboratory services.[8]

Previous cost modelling work in Pixley ka Seme district[8] reveals that establishing this small tier 3 site may cost marginally more per CD4 test than a CD4 tested in a centralised larger laboratory (ZAR28.62/USD2.06 at exchange rate of ZAR13.90 per USD on 20 November 2015). This is, however, substantially cheaper than implementing widespread use of POC technology (44 tier 1 facilities) across the district’s 44 clinic sites, where it has been estimated that widespread CD4 provided at the POC could cost Pixley ka Seme in excess of ZAR4.5 million/USD325700 (at ZAR49.25/USD32.32 per test) as opposed to an estimated ZAR1.03 million/USD74 000 needed to provide a tier 3 laboratory-based service offering the same number of tests performed, but at the De Aar facility. The additional benefit, as a trade-off for the small incremental cost, is that there are less local logistics costs incurred, as testing is performed on-site, coupled with better sample integrity and significantly improved local TAT – all of which has positive implications for better patient management.

Conclusion
The extended decentralised 3/community laboratory implementation at the De Aar laboratory in Pixley ka Seme offers a compromise between cost and accessibility, with a small incremental cost (ZAR28.63/USD2.06)[8] but substantially lower TAT, while maintaining a quality of testing expected from a larger laboratory. Owing to the success of the De Aar pilot site, additional implementation sites, also identified through ITSDM planning, have been proposed to improve and extend services elsewhere in under-serviced districts using existing NHLS laboratories, including Alivil North (Eastern Cape), Lephalele (Limpopo), St Patricks (Eastern Cape) and Vredenburg (Western Cape).

Study limitations
Owing to the absence of an end-to-end sample tracking system, it was not possible to report TAT for afferent (time from sample collection by the courier at the health facility to registration on the LIS) and efferent (time from result authorisation to result delivery to the health facility) phases of the laboratory value chain. Additionally, for interlaboratory referrals we were unable to identify whether delays were due to courier delays or to pre-analytical receiving office delays.

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Conflict of interest. DKG declares that the employer, the NHLS, is the sole owner of the patent held for the PanLeucogated CD4 method, currently licensed to Beckman Coulter International and receives royalties. Through open public tender, Beckman Coulter was awarded a service level agreement to provide CD4 testing services in the NHLS. All LIS sample-related CD4 data were used with permission from the NHLS CEO, Dr Seagie Pillay.

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