The importance of animals in human schistosomiasis in South Africa

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Summary

The importance of animals in human schistosomiasis in South Africa is reviewed. The prevalence of animal schistosome species in humans, the role of animals as reservoir hosts of Schistosoma haematobium and S. mansoni, and the possibility of falsepositive serological reactions in humans following exposure to animal or bird bilharzia are considered.

It is concluded that, as regards animal schistosomes, at present only S. mattheei and a hybrid of S. mattheei and S. haematobium pose a potential threat to human health in South Africa.

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In southern Africa schistosomiasis is practically as widely disseminated in animals as in humans. The prevalence in animals is very high in certain areas, for instance up to 90% in parts of the lowveld of the Transvaal.

Schistosoma mattheei is the only schistosome of importance in animals in South Africa; with the exception of S. mansoni, which has been recovered from primates and rodents and a single waterbuck, and S. haematobium, recovered from one buffalo, it is probably the only schistosome of mammals occurring naturally in domestic and wild animals in the country.

S. mattheei was discovered in 1926 by Veglia and Le Roux2 in sheep near Humansdorp in the eastern Cape Province, and Le Roux3 recovered S. mattheei ova from cattle faeces on the same farm. Although regarded by some 4.5 as equivalent to or a variant of *S. bovis*, differences in the ova 6.8 and biological and biochemical differences 9-11 have confirmed it as a separate species. Occurrence of S. bovis has not been confirmed in South Africa.

In this paper the following aspects were discussed: (i) the prevalence and importance of S. mattheei in humans; (ii) the role of animals as reservoir hosts of S. mansoni and S. haematobium; (iii) animal schistosome species beyond our borders; (iv) serological reactions in humans; and (v) avian bilharzia.

The prevalence and importance of S. mattheei in humans 12

S. mattheei, which shares the intermediate snail host, Bulinus (Physopsis) spp., with the human schistosome, S. haematobium, is primarily of veterinary importance. Apart from South Africa, it occurs in most neighbouring territories and as far north as parts of Tanzania, Chad and Nigeria.1

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The wide definitive host range includes man, cattle, sheep, goats and horses among the domestic animals and a large number of wild animals, including primates (baboons and monkeys), antelopes (kudu, waterbuck, impala, nyala, puku, lechwe, etc.), zebra, buffalo, giraffe, bushpig, warthog and various

As can be expected from the fact that they share a common intermediate host, S. mattheei and S. haematobium have a very similar distribution in South Africa - mainly in the lowveld of the Transvaal, in large parts of Natal and coastal areas as far south as Humansdorp in the Cape. They also occur to a lesser extent in the Transvaal highveld. C. J. Visagie (personal communication) estimates that 3-4 million people in South Africa are infected with S. haematobium and/or S. mansoni and Pitchford13 has reported that in the areas of highest prevalence (more than 90% S. haematobium in humans and up to 90% S. mattheei in cattle, for example in the Transvaal lowveld) man and domestic and wild animals often share the same water sources infested with the intermediate snail host.

Man is often exposed to S. mattheei infection. The parasite has the ability to develop in humans, as cattle schistosome ova have been demonstrated in both urinary and intestinal infections in humans in southern Africa by numerous workers. 6,13-17 With one exception,16 cattle schistosome ova were always accompanied by S. haematobium and/or S. mansoni. 13

Nevertheless, it is difficult to gauge just how susceptible man is to S. mattheei. In some well-documented cases man appears to be practically refractory to infection. While Le Roux3 had good evidence that people were exposed to S. mattheei, by swimming in and drinking infested water, he could not demonstrate infection in them: 'Judging from the fact that the pool responsible for the infection of the sheep is used by man and beast alike, it is only natural to assume that if man could be infected, the human beings on the farm should have been. Both urine and faeces were examined with negative results.' He also examined 2 workers who had swum in the water but obtained consistently negative faecal and urine examinations 1 day and 2 and 3 months after exposure.

On the other hand, S. mattheei infections in humans are sometimes very prevalent, high infection rates having been reported from parts of the Transvaal, for instance. Pitchford13 surveyed 275 people from different parts of the Transvaal and found cattle schistosome ova in the rectum and bladder in humans from the whole of the eastern Transvaal, east of the Drakensberg, from the western Transvaal, and Swaziland. He concluded: 'The distribution of the parasite in man in South Africa therefore probably follows very closely that of S. haematobium, and though not many cases have been found in the north-western and central Transvaal, this may be due to insufficient search being made.' He found a prevalence of up to 40% of cattle schistosomiasis in humans on one farm in the Komatipoort area in the eastern Transvaal (R. J. Pitchford - personal communication).

The apparent anomaly therefore exists that up to 40% of humans have been found to be infected with cattle schistosomes in certain areas, while in other areas they do not appear to be susceptible.

A comparable situation exists in the case of S. japonicum in the Far East. While a zoophilic strain of this parasite in Taiwan is

apparently not infective to man, the same species from other localities is known to be highly infective to man. 18

Van Wyk¹² suggests that an answer to these apparent strain differences in South Africa may lie in the fact that some of the schistosome ova recovered from man were not pure S. mattheei, but a hybrid between S. mattheei and S. haematobium, as demonstrated by Pitchford,¹⁹ who recovered ova indistinguishable from S. mattheei from man, which, when passaged in rodents and a calf gave rise to ova intermediate in size between S. mattheei and S. haematobium. In another case typical-looking S. haematobium ova recovered from a naturally infected rodent from Komatipoort gave rise to typical but short S. mattheei ova when passaged in white mice.¹⁹

Subsequently Taylor¹⁰ demonstrated that *S. mattheei* and *S. haematobium* can cross, giving rise to a viable hybrid. Wright and Ross²⁰ confirmed (by iso-electric focusing of enzymes of worms obtained by passage from 2 patients) that this hybrid does occur naturally in man in South Africa and concluded that the shape of the eggs produced is: '... not necessarily a guide to the genetic constitution of the enclosed larvae'.

The possibility therefore exists that the apparent *S. mattheei* ova demonstrated in man in areas of high prevalence may be from hybrid parents producing ova morphologically indistinguishable from those of *S. mattheei*. Furthermore, by inference, some of the typical-looking *S. haematobium* ova in these people may also be hybrids. While not confirmed in the limited study by Wright and Ross²⁰ this inference is perhaps not too farfetched if one considers the abovementioned typical-looking *S. haematobium* ova from a rodent giving rise, on passage in mice, to almost typical *S. mattheei* ova.¹⁹

Van Wyk¹² suggested that patent infections of *S. mattheei* may develop in man only if exposure occurred for a certain minimum time, and that this may tie up with Pitchford's^{13,19} surmise that *S. mattheei* females can give rise to patent infections in man only if carried by foreign or hybrid males. Continued exposure of man to large numbers of *S. mattheei* cercariae may improve the chances of female *S. mattheei* pairing with male *S. haematobium*.

The exact character of what has thus far been regarded as S. mattheet in man remains to be determined, which may be quite difficult despite the availability of sophisticated techniques such as iso-electric focusing. If an S. mattheet-like ovum is an F_1 hybrid, identification would be relatively easy. Where, however, the hybrid has been bred back repeatedly to one of the parent species, the shape of the ovum may be typical of one parent species, but will passage and enzyme comparisons reveal the fact that it is not of pure origin? More work is necessary, including cross-breeding studies proceeding further than the F_1 hybrid or even the first 2 or 3 crosses to determine up to what point the hybrids can be identified by any presently available means.

We have ample evidence that man is susceptible to cattle schistosomes and hybrids, but no positive suggestions have been forthcoming concerning the possible clinical effect of *S. mattheei* or the *S. mattheei/S. haematobium* hybrid, either on their own or together with the human schistosomes.

At present *S. mattheei* or hybrid infections are probably not a serious health problem in South Africa. It is, however, difficult to estimate the clinical effect because *S. mattheei* is invariably associated with *S. haematobium* or *S. mansoni* in human cases. Nevertheless, Pitchford¹⁹ was able to conclude: '. . . it appears that the cattle schistosome *S. mattheei* will infect man, but that man is not a particularly good host; if he were, there seems no reason why the incidence of *S. mattheei* in man should not be as high as that of *S. haematobium* in those areas where there is close association between the two definitive hosts'.

Furthermore, in cases where S. mattheei succeeds in infecting man, the infection does not seem to be sustained. Although it must be borne in mind that they examined only urine, Pitchford and Visser²¹ found only relatively low S. mattheei egg counts with no sustained rise in 10 children examined regularly over 14 months.

A prevalence as high as 40% in some areas, however, indicates that the parasite (pure strain or hybrid) may be adapting to the human host. As stated by Pitchford, 19 . . . there is every likelihood that the incidence of S. mattheei in man might increase. The resulting hybrids will in time possibly supplant S. mattheei and S. haematobium with a schistosome infecting man and cattle with equal ease. Already Pitchford and Lewis 22 have suggested that the poor response of S. mattheei in humans to oxamniquine treatment may be due to hybridization with S. haematobium, which is not susceptible to the drug.

The role of animals as reservoir hosts of S. mansoni and S. haematobium

For a reservoir host (defined by Nelson *et al.* ²³ as: 'an animal which maintains under natural conditions an infection transmissible to man') to be an important source of *Schistosoma* infection in man it must be readily susceptible, must occur in sufficiently large numbers, so that a high level of transmission is maintained, and must frequent water sufficiently so that substantial amounts of excreta containing large numbers of schistosome ova are deposited in or near water. As the two human schistosomes in South Africa differ markedly in their infectivity for various definitive hosts, they need to be considered separately.

S. mansoni

Pitchford¹ lists the following animals as susceptible to *S. mansoni:* chimpanzee, various species of baboon, grivet monkey, sheep, dog, waterbuck, various species of rodent and two species of shrew, of which the chimpanzees, baboons and perhaps the monkeys are of potential practical significance as hosts.

Elsewhere in Africa it appears that baboons are able to maintain *S. mansoni* in nature for at least 18 months and that the *S. mansoni* is infective to man.²⁵ In South Africa Pitchford *et al.*²⁶ found only 1 out of 280 samples of baboon faeces containing eggs of *S. mansoni* (compared with 22 with *S. mattheei* ova); they found no infected humans, but this is probably inconclusive as the survey was conducted in the Kruger National Park with only a very low human population. No infected monkeys have as yet been found, but perhaps insufficient search has been made.

Although *S. mansoni* has been recovered from a few rodents, this is probably of little importance in this country. ^{13,27} Infection in sheep and dogs is very rare in nature, and all the ova recovered by Pitchford *et al.* ²⁸ from a single waterbuck were dead.

S. haematobium

The following records of natural infection are listed for *S. haematobium*: 1 two species of baboon, chimpanzee, vervet monkey, pig, sheep and the Cape buffalo. Nelson *et al.* 23 consider man to be the only true host of *S. haematobium* and that the few records of this parasite from animals represent incidental or dead-end infections. No evidence exists to dispute this contention

Animal schistosome species beyond our borders

The two species concerned are *S. margrebowiei* and *S. leiperi*, referred to as 'lechwe schistosomes' by Pitchford.²⁹ He considers the reports on *S. spindale* and *S. japonicum* in southern Africa to represent probable records of lechwe schistosomes. He lists man

as being susceptible to S. margrebowiei but not to S. leiperi. Man, however, appears to be a poor host, and cases of infection are rare. According to Pitchford and Wolstenholme 30 '... the low S. margrebowiei egg counts . . . suggest that man is a poor host with low grade short lived infections . . . Alternatively man acts as a host in the same manner as cattle, i.e. unisexual male infections occur with an occasional female reaching maturity, which may or may not be short lived.' It is therefore very unlikely that these animal schistosomes will be important in humans. On the contrary, there is circumstantial evidence that the animal schistosomes (and specifically the two lechwe schistosomes) may limit or prevent the spread of the human schistosomes (S. mansoni and S. haematobium) presumably by stimulating resistance to infection.

Le Roux, cited by Pitchford,29 suggested that animal schistosome cercariae may immunize man against human species and vice versa. Nelson et al.23 coined the term 'zooprophylaxis' for instances where zoonoses '. . . stimulate man's immunological mechanisms so that he is able to resist the more pathogenic organisms in his environment'. An example given is the previously mentioned zoonotic strain of S. japonicum in Taiwan that fails to reach maturity in man, possibly the reason why the pathogenic strain of S. japonicum has not become established in that country.31

Indications of an inverse relationship between human schistosomiasis and the presence of lechwe schistosomes have been seen in the Okavango and Caprivi, 29,30 and very little human bilharzia was found in all areas where lechwe (shown to be commonly infested with the lechwe schistosomes) were common, a moderate amount where lechwe were scarce, and high rates where lechwe and puku were absent. Furthermore, over a number of years the rate of infection at Maun in Botswana has increased very greatly, in parallel with a simultaneous decrease in the lechwe population in the area. There are also indications that the lechwe schistosomes may have blocked the advance of S. mattheei in animals and man in these areas.

Serological reactions in humans

There are indications that false-positive serological reactions result in man from infection with animal schistosomes. Pitchford and Wolstenholme30 surveyed 77 children from areas of the eastern Caprivi not endemic for human schistosomiasis and obtained 62% positive serological reactions. They found no schistosome ova and concluded that the children had been exposed to lechwe schistosomes. In two other areas where human schistosomiasis is endemic, positive serological reactions corresponded closely with the occurrence of ova in the excreta of the patients.

Apart from this limited evidence in southern Africa, similar results were obtained by Sadun and Biocca³² with S. bovis in Sardinia. Using S. mansoni antigen in fluorescent antibody tests, they reported a 40% positivity rate in humans with a history of cercarial dermatitis. They posed the question whether this phenomenon perhaps signifies the presence of cross-immunity between S. bovis and human schistosomes. Wright 33 doubts this possibility because of a high prevalence of S. bovis in cattle and S. haematobium in humans in Gambia where the two parasites share the same intermediate host. The circumstantial evidence supplied by Pitchford and co-workers, however, seems to point to a different situation in the Caprivi and the Okavango.

Avian bilharzia

Exposure to avian schistosomes also needs to be considered as a possible cause of cercarial dermatitis and false-positive serological reactions in humans.

Conclusion

The only animal schistosome that at present constitutes a threat to human health in South Africa is S. mattheei, a parasite with such a wide host range (including cattle and game) that control will be extremely difficult, especially in areas where man and cattle share the same water sources. Control in cattle is much more feasible than in game, and reduction of the prevalence and the level of infestation of cattle should greatly reduce the threat to which man is exposed.

The S. mattheei/S. haematobium hybrid poses a potential but as yet unidentified threat to human health. If this hybrid can successfully maintain itself in man in the absence of animals, it could prove to be of considerable importance. It is suspected to be in the process of adaptation to man, and therefore this situation should be closely monitored.

This article is dedicated to Dr R. J. Pitchford, world-renowned for his work on the epidemiology of both human and animal schistosomiasis in southern Africa. Without the work of R. J. Pitchford our knowledge of the epidemiology of bilharziasis in this country as well as in some neighbouring territories would have been scanty to the point of being well-nigh non-existent.

REFERENCES

- Pitchford RJ. A check list of definitive hosts exhibiting evidence of the genus Schistosoma Weinland, 1858 acquired naturally in Africa and the Middle East. J Helminthol 1977; 51: 229-252.
- Veglia F, Le Roux PL. On the morphology of a schistosome (Schistosoma mattheei, sp. nov.) from the sheep in the Cape Province. Annual Report, Director of Veterinary Services, Union of South Africa 1929. Pretoria: Government Printer, 15: 335-346.
 Le Roy, Pl. 2019.
- Le Roux PL. Remarks on the habits and the pathogenesis of Schistosoma mattheei together with notes on the pathological lesions observed in infected sheep. Annual Report, Director of Veterinary Services, Union of South Africa 1000. Pretoria: Government Printer. 15: 347-406.

- 1929. Pretoria: Government Printer. 15: 347-406.
 MacHattie C, Chadwick CR. Schistosoma boris and S. mattheei in Irak with notes on the development of eggs of the S. haematobium pattern. Trans R Soc Trop Med Hyg 1932; 26: 147-156.
 Van den Berghe L. A morphological study of bovine schistosomes. J. Helminthol 1937; 15: 125-132.
 Alves W. The eggs of Schistosoma bovis, S. mattheei and S. haematobium. J. Helminthol 1949; 23: 127-134.
 Dinnik JA, Dinnik NN. The schistosomes of domestic ruminants in Eastern Africa. Bull Epizoot Dis Afr 1965; 13: 341-359.
 Pitchford RJ. Differences in the egg morphology and certain biological characteristics of some African and Middle Eastern schistosomes. Genus Schistosoma, with terminal-spined eggs. Bull WHO 1965; 32: 105-120.
 Pitchford RJ, Meyling AH, Meyling J, Du Toit JF. Cercarial shedding patterns of various schistosome species under outdoor conditions in the Transvaal.
- terns of various schistosome species under outdoor conditions in the Transvaal.

 Ann Trop Med Parasitol 1969; 63: 359-371.

 Taylor MG. Hybridisation experiments on five species of African schistosomes. J Helminthol 1970; 44: 253-314.

 Ross GC, Southgate VR, Knowles RJ. Observations on some isoenzymes of strains of Schistosome hours. Support Schistosome hours.
- strains of Schistosoma bovis, S. mattheei, S. margrebowiei and S. leiperi. Z. Parasitenka 1978: 57: 49-56.
- Van Wyk JA. Transmission of Schistosoma mattheei from animals to man. In: Gear JHS, ed. Medicine in a Tropical Environment. Proceedings of the Interna-tional Symposium South Africa 1976. Cape Town: AA Balkema, 1976: 705-717.
- tional Symposium South Africa 1976. Cape Town: AA Balkema, 1976: 705-717.

 13. Pitchford RJ. Cattle schistosomiasis in man in the Eastern Transvaal. Trans R Soc Trop Med Hyg 1959; 53: 285-290.
- Cawston FG. The experimental infestation of Physopsis africana. Ann Trop Med Parasitol 1922; 16: 207-211.
- Blackie WK. A helminthological survey of Southern Rhodesia. Mem Lond Sch Hyg Trop Med 1932; 5: 1-91.
 Kisner CD, Stoffberg N, De Meillon B. Human infection with Bilharzia bovis. S Afr Med J 1953; 27: 357-358.
- Cruz e Silva JA. Contribuição para o estudo dos helmintes parasitas dos vertebrados de Moçambique. Mem Junta Invest Ultramar 1971; No. 61: 50-64.
- Hsi HF, Hsii SY Li. On the infectivity of the Formosan 1strii, 30.6.1.305.
 Hsi HF, Hsii SY Li. On the infectivity of the Formosan 1striin of Schistosoma japonicum in Homo sapiens. Am J Trop Med Hyg 1956; 5: 521-528.
 Pitchford RJ. Observations on a possible hybrid between the two schistosomes. S. haematobium and S. mattheei. Trop Med Hyg 1961; 55: 44-51.
 Wright CA, Ross GC. Hybrids between Schistosoma Faematobium and S. mattheei and their identification by isoelectric focusing of enzymes. Trans R Soc Trop Med Hyg. 1980: 74: 326-332. Trop Med Hyg 1980; 74: 326-332.
 21. Pitchford RJ, Visser PS. Excretion of Schistosoma mattheei eggs from man,
- baboons and cattle living in their normal environment. J Helminthol 1975; 49: 137-142
- Pitchford RJ, Lewis M. Oxamniquine in the treatment of various schistosome infections in South Africa. S Afr Med J 1978; 53: 677-680.

 Nelson GS, Teesdale C, Highton RB. The rôle of animals as reservoirs of bilharziasis in Africa. In: Wolstenholme GEW, O'Connor M, eds. Ciba Foundation Symposium. Bilharziasis. London: J & A Churchill, 1962: 127-156.

 Nelson GS. Schistosome infections as zoonoses in Africa. Trans R Soc Trop Med Hyg 1960; 54: 301-324.

 Fenwick A. Baboons as reservoir hosts of Schistosoma mansoni. Trans R Soc Trop Med Hvg 1969; 63: 557-567.

 Pitchford RJ, Visser PS, du Toit JF, Pienaar U de V, Young E. Observations on the ecology of Schistosoma mattheet Veglia and Le Roux 1929, in portion of the Kruger National Park and surrounding area using a new quantitative technique for egg output. 7 S Afr Vet Assoc 1973; 44: 405-420.

Pitchford RJ, Visser PS. The role of naturally infected wild rodents in the epidemiology of schistosomiasis in the Eastern Transvaal. Trans R Soc Trop Med Hyg 1962; 56: 126-135.

- Pitchford RJ, Visser PS, Pienaar U de V, Young E. Further observations on Schistosoma mattheei, Veglia and Le Roux, 1929, in the Kruger National Park. J S Afr Vet Assoc 1974: 45: 211-218.
- 29. Pitchford RJ. Preliminary observations on the distribution, definitive hosts and

- possible relation with other schistosomes, of Schistosoma margrebowiei, Le Roux, 1933 and Schistosoma leiperi, Le Roux, 1955. J Helminthol 1976; 50: 111-123.
- Pitchford RJ, Wolstenholme B. Further observations on the relationship and distribution of Schistosoma margrebowiei and S. leiperi in central southern Africa. J Helminthol 1977; 51: 327-336.
- Nelson GS. Zooprophylaxis with special reference to schistosomiasis and filariasis. In: Soulsby EJL, ed. Parasitic Zoonoses: Clinical and Experimental Studies. London: Academic Press, 1974: 273-285.
- Sadun EH, Biocca E. Intradermal and fluorescent antibody tests on humans exposed to Schistosoma boeis cercariae from Sardinia. Bull WHO 1962; 27: 810-814.
- Wright CA. General discussion. In: Wolstenholme GEW, O'Connor M, eds. Ciba Foundation Symposium: Bilharziasis. London: J & A Churchill, 1962: 154.