



FACULTY OF VETERINARY MEDICINE
USMANU DANFODIYO UNIVERSITY
P.M.B. 2346, SOKOTO, NIGERIA



Sokoto Journal of Veterinary Sciences

ISSN 1595-093X

Nwanta *et al.* /*Sokoto Journal of Veterinary Sciences* (2008). 7(2): 42-45.

Field trial of Malaysian thermostable Newcastle disease vaccine in village chickens in Kaduna State, Nigeria

JA Nwanta^{1*}, JU Umoh¹, PA Abdu¹, I Ajogi¹, SC Egege¹ and AA Adeiza²

¹*Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria, Nigeria*

²*College of Agriculture and Animal Science, Ahmadu Bello University, Mando, Kaduna, Nigeria*

Correspondence Author:

Abstract

Village chickens in Kaduna State, Nigeria were vaccinated once with a Malaysian heat-resistant Newcastle disease vaccine (NDV4HR) given in feed. In all, 1605 chickens in 223 households covering 33 villages and 13 Local Government Areas were tagged and bled before vaccination and two weeks after vaccination. Antibodies to Newcastle disease virus were titrated by haemagglutination inhibition test and titres $\geq 3(\log_2)$ were assumed to be protective. Presumed protective titres

were recorded in 143 (8.9%) of chickens before vaccination and in 957 (65.5%) after vaccination. Recommendation is made for the widespread adoption of this technology.

Keywords: Newcastle disease vaccine; Antibody response; Vaccination; Village chicken; Thermostability.

Introduction

Newcastle disease (ND) is reported as the most important viral disease of poultry in the world including developing countries (Nawathe et. al., 1975; Adu et. al., 1986; Adene, 1990; Spradbrow, 1997). It has a devastating effect on commercial as well as village poultry industries (Philips, 1973; Nawathe et. al., 1988; Okeke and Lamorde, 1988; Shamaki et. al., 1989; Adene, 1997). The resources derivable from the chickens cannot be fully utilized unless the disease is controlled particularly in the village poultry flocks that are believed to keep the virus circulation and act as reservoirs and carriers to themselves and the more susceptible exotic breeds in commercial farms (Nawathe et. al., 1975; Ezeokoli et. al., 1984; Gomwalk et. al., 1985; Adu et. al., 1986; Nwosu and Okeke, 1989; Olabode et. al., 1992).

Vaccination has been reported as the only safeguard against endemic ND (Orajaka et. al., 1999; Usman, 2002). The village chickens, in multi-age flocks scattered in small number over villages, are difficult to catch for formal vaccination as adopted in commercial chicken enterprise (Aini et. al., 1990; Orajaka et. al., 1999; Usman, 2002). It therefore takes great effort and time trying to vaccinate them. More importantly, these conventional vaccines are not heat-labile and therefore require complex cold-chains to link the vaccine producers and users (Aini et. al., 1990; Usman, 2002).

Preliminary work conducted elsewhere indicates that heat-stable a virulent V4 or 1-2 strains selected for heat resistance and applied through eye drop, drinking water or food and feed particles has been found to be a suitable oral vaccine for village chickens (Saglid and Spalatin, 1982; Spradbrow, 1988; Spradbrow, 1992; Bell et. al., 1995; Tu et. al., 1998). This V4 or 1-2 vaccine has been successfully used in many African countries such as The Gambia (Jarra et. al., 1991), Cameroon (Bell et. al., 1995), Ghana (Amakye – Anim et. al., 1998) and many countries of South-East Asia (Copland, 1987). Maize byproducts after processing have been found to be good carriers of V4 in the vaccination of birds against ND in Nigeria (Olabode, 1996).

In Malaysia, a 60% protection rate was recorded on challenge of village chickens with virulent NDV strain and farmers in Malaysia have benefited from the oral food vaccination of their flocks with NDV4HR (Aini et. al., 1990). It therefore became necessary to assess the immunological status of Nigerian village chickens and to conduct field trails with this newly developed vaccine for the purpose of determining its suitability as a rural poultry vaccine in Kaduna State, Nigeria. In view of the economic importance of ND in Nigerian village chickens, indications are that local poultry farmers would welcome ND vaccinations capable of protecting their flocks. The objective of this study was to investigate the seroprevalence antibody status of ND in Nigerian local chickens and to conduct field vaccination trails with ND4HR in village chicken flocks in Kaduna State, Nigeria. The outcome of the results obtained would form the basis for recommendations on whether the new vaccination technology will be adopted in Kaduna State, Nigeria.

Materials and Methods

The study area

The study covered thirteen (13) Local Government Areas of Kaduna State located within the semi-arid and sub-humid zones of North Central Zone of Nigeria. Kaduna State is situated between 8° 45' and 11° 30' N and 6° 10' and 9° E. The mean annual temperature is about 34°C with the hottest months being from March – April (40°C) and the coldest period (13.2°C) is between December and January during the severe harmattan. Rainfall varies between 1,000mm and 1,500mm and rainy season lasts for 150-200 days (Mid April-end to October). The dry season occurs from late October to early April (RIM, 1993).

Flocks Sampling Procedure

Prior to the survey, group meetings were held with the Management Staff of the State Veterinary Department and the Heads of Agricultural Department of the 23 Local Government Areas (L.G.A.'s) of the State for discussion on the project objective, benefits, sample size and areas to be covered. At the meeting, it was decided that a total of 2,000 village chickens in 250 households and 40 villages be sampled in two local government areas of the State (Kaduna North and South) based on high population density of village chickens in the areas. Out of the list of 60 villages supplied by the State, 40 villages were randomly selected through balloting and out of a total of 400 households listed by the village heads of the 40 villages selected, 250 households were also randomly selected.

Due to poor response from the majority of the farmers who refused to allow their chickens to be bled for fear of death, despite the offer of free poultry drugs and feeds as incentives to them, only 300 village chickens in 50 households and 10 villages were sampled initially. Additional 11 Local Government Areas consisting of 50 villages, 250 households and 2000 village chickens were listed for sampling based on the areas with high concentration of village chickens and presence of United Nations Development Programme (UNDP) activities on agriculture.

Scheduled visits were made to the additional proposed Local Government Areas and villages in the company of the LGAs representatives and meetings were held with the village heads and farmers to discuss the objectives of the project, its benefit and the areas to be sampled. The list of the villages and households were supplied by the village heads. From the list, additional number of 1,305 village chickens in 173 households of 23 villages were selected from the 11 Local Government Areas that responded. Thus a total of 1,605 village chickens in 223 households covering 33 villages and 13 Local Government Areas were selected for the study.

NDV4HR vaccine

The NDV4HR used for the field trails in a freeze-dried live thermostable strain vaccine imported from Malaysian Vaccines and Pharmaceutical SNP, BHD. Each vial of the vaccine contains 100 bird doses. It is a lentogenic live virus selected for its heat resistance. The vaccine dose per bird was 106.0 EID₅₀ (50% embryo infective doses).

Vaccination of Chickens and Blood Sampling

Vials of the NDV4HR were reconstituted in the feed as recommended by the manufacturer and administered to the birds. The chick mash used was obtained from Sanders Feeds (SEEPIC Nigeria Limited), Kaduna Depot, Nigeria. The chick mash contains 19.00% crude protein, 6.00% crude fat, 1.25% calcium, 0.65% phosphorus, 1.00% lysine, 0.80% methionine, 5.00% crude fibre and 2700 kilocalorie per kg metabolisable energy. The required dose per flock in each household was calculated based on the recommendation of 10g of the feed per bird (Aini et. al., 1990). In the calculated amount of feed was added an equivalent amount of diluent (well water), 10ml of well water containing 106.0 EID₅₀ of vaccine per 10g of feed. The moist mixture was then put in a clean feeding trough under a shade for the birds to consume. All the chickens in the flocks selected were vaccinated. About 50% of the chickens in each vaccinated flock were caught and wing-tagged, bled and released. Two weeks post vaccination, the wing-tagged birds were bled again. A total of 1,605 wing-tagged village chickens were bled before and after vaccination. Both pre and post vaccination sera were tested for NDV haemagglutination inhibition (HI) antibodies.

Newcastle Disease Virus Antigen Preparation

The antigen was prepared from NDV-LaSota vaccine obtained from the National Veterinary Research Institute, Kaduna Field Station. The 200 dose vial of the NDV-LaSota vaccine was reconstituted in 8ml distilled water. The haemagglutination (HA) titre was determined as described by Beard (1980) and diluted to contain 4 HA

units for use in the HI test as described by Allan and Gough (1974).

Data Analysis

Number of birds with detectable NDV antibody titres were calculated. Sera with HI titres $\geq 3(\log_2)$ were considered positive or protective based on the reports of Allan et. al., (1974); Westbury et. al., (1984); Aini et. al., (1990), Chi-square was used to determine the relationship between the number of village chickens that had antibody titre $\geq 3(\log_2)$ pre and post vaccination periods.

Results

Out of a total of 1,605 village chickens screened for NDV antibody, 143 (8.90%) had antibody titre $\geq 3(\log_2)$ before vaccination (Table 1). Also out of the 143 birds that had protective antibody titre $\geq 3(\log_2)$ prevaccination, only 135 (94.41%) had protective titre $\geq 3(\log_2)$ post vaccination. Very few proportion (5.59%) of the birds that tested positive before vaccination tested negative after vaccination.

About 65.46% (95% CI = 63.02 % < P < 67.88%) of the birds that had antibody titre $\leq 3(\log_2)$ before vaccination sero converted to higher HI titre $\geq 3(\log_2)$ post vaccination (Table 1). Presumed protective titres were recorded in 1092 (68.04%) of birds post vaccination (Table 1). There was a statistically significant difference in the frequency of protective titre pre- and postvaccination ($X^2 = 48.87$, 1df, $P < 0.001$).

Table 1: Number of birds with protective antibody titres to ND pre and post vaccination

		Post vaccination		
		Titre $\geq 2^3$	Titre $\leq 2^3$	Total
Pre-vaccination	Titre $\geq 2^3$	135 (94.4%)	8 (5.6%)	143
	Titre $\leq 2^3$	957 (65.5%)	505 (44.5)	1462
	Total	1092 (68.0%)	513 (32.0%)	1605

Discussion

In this study, the HI titre Titre $\geq 3(\log_2)$ were considered positive (protective) based on the findings of Allan et. al., (1974); Westbury (1984) and Bell et. al., (1991a) who reported that birds with HI titres Titre $\geq 3(\log_2)$ were protective against challenge with a virulent strain of ND virus. The protective antibody titres $\geq 3(\log_2)$ recorded in 143 (8.9%) out of 1605 village chickens screened for HI antibody before vaccination suggested previous exposure of the birds to field strains of ND virus (velogenic). This is in agreement with the report of Alexander (1998) who showed that in all paramyxoviruses, birds that have not been immunized or infected usually have HI titres $\geq 3(\log_2)$ and that non-specific titres above this levels are rare in avian species. Also farmers and veterinary staff interviewed reported that no ND vaccination has been conducted in these villages and their respective neighbourhoods during the past 12 months prior to the study. Reports of ND outbreaks in the monthly returns from the Local Government Veterinary Officers, Ezeokoli et. al., (1984) and personal communications with the farmers confirmed ND as a widespread problem in the study area. It has also

observed that a small proportion (5.6%) of the birds that were protected prevaccination, tested negative post vaccination. This observation contrasts with the previous findings of Bell et. al., (1991b) who found that positive breeders (naturally exposed) responded serologically and that the number of birds with titres $\geq 3(\log_2)$ increased significantly following vaccination. In addition, about 65.5% of the birds that had antibody titres $\geq 3(\log_2)$ before vaccination seroconverted to protective level ($\geq 3(\log_2)$) post vaccination. The statistically significant difference in the frequency of protective titre prevaccination and post vaccination signifies that the vaccine, NDV4HR, is immunogenic and capable of provoking antibody response (Aini et.al., 1990).

Conclusion

Recommending the use of NDV4HR to Nigerian rural farmers requires that the local conditions, protective ability of the vaccine and ease of administration is taken into consideration. In Kaduna State of Nigeria, where farming is predominant, many farmers are yet to be convinced of the benefits of ND vaccination. Achieving good level of protection with the use of NDV4HR in vaccination of chicken will stimulate the interest of rural farmers in adopting the new vaccine technology.

The use of NDV4HR in a single oral vaccination of village chickens in this study, has achieved a 65.5%

protection. Therefore, subsequent application of more doses of the vaccine at intervals to be determined by further research may result in the development of a higher protection in greater proportion of chickens and subsequent reduction in the outbreaks of ND. Finally, controlling ND in the endemic rural areas of Nigeria through the use of NDV4HR will subsequently control the spread of the ND virus in the neighbouring countries such as Cameroon, Tchad, Niger and Benin Republic.

Acknowledgements

We are grateful to Prof. (Dr.) Aini Ideris of the Faculty of Veterinary and Animal Science, University Pertanian, Malaysia, Prof. Dr. Abd Latif Ibrahim of the National Biotechnology Directorate, Ministry of Science, Technology and Environment, Malaysia, Dr. Roshidah Ismail and Dr. Mazlan Mohamad of Malaysian Vaccines and Pharmaceuticals SDN, BHD Malaysia for their efforts in making the vaccine, NDV4HR, available to us at no cost. We also acknowledge the cooperation of Mrs. J.C. Atawodi, laboratory technologist, Department of Veterinary Public Health and Preventive Medicine, Ahmadu Bello University, Zaria, Nigeria.

Other people worthy of note for their contribution include Dr. Y.B. Audi, Director Veterinary Services, Kaduna State, Nigeria and Mr. J.A. Alabi of the Federal Department of Livestock and Pest Control Services, Kaduna, Nigeria.

References

- Adene, D.F. (1990). Country report on the management and health problems of rural poultry stock in Nigeria. In: *Proceedings, Centre for Tropical Agriculture (CTA). Seminar on Smallholder Rural Poultry Production. Thessaloniki, Greece, 9-13 October, 1990*, Volume 2, Pp 175-182
- Adene, D.F. (1997). Diseases of Poultry in Nigeria. An overview of the problems and solution. *Trop. Vet.*, **15**:103-110.
- Adu, F.D; Edo, U. and Sokale, B. (1986). Newcastle disease: The immunological status of Nigerian local chickens. *Trop. Vet.*, **4**: 149-152
- Aini, A.; Ibrahim and A. Mustaffa Babjee, (1990). Feed-based Newcastle disease vaccine for village chickens. *Poultry International* Publication, December edition, Pp. 24-28.
- Alexander, D.J. (1998). Newcastle disease virus and other paramyxoviruses. In: *Isolation and identification of Avian Pathogens (4th edition)*. American Association of Avian Pathologists. pp. 156-163.
- Allan, W.H., Gough, R.E., 1974. A standard haemagglutination-inhibition test for Newcastle disease: I.A. comparison of macro-and micro-methods. *Vet. Rec.* **95**:120-123.
- Allan, W.H., Lancaster, J. E. and Toth, B. (1978). Newcastle disease vaccines – their production and use. *Food and Agriculture Organisation Animal Production and Health Series No. 10*, FAO, Rome, Italy. Pp. 51-108
- Amakye-Anim, J., Alders, R.G., and Spradbrow, P.B., (1998). Trials with V4 Newcastle Disease Vaccine in Ghana. In: *Proceedings of Scientific Session at the Fourth Asia Pacific Poultry Health Conference, Melbourne, Australia, November 22-26, 1998*, pp. 124.
- Baba, S.S.; El-Yuguda, A.D. and Baba, M.M. (1998). Serological evidence of mixed infection with Newcastle disease and Egg Drop Syndrome 1976 viruses in village chicken in Borno State, Nigeria. *Trop. Vet.* **16**:137-141.
- Beard, C.W. (1980). Serological procedures. In: *Isolation and Identification of Avian Pathogens*. Edited by S.B. Hitchner, C.M. Domarmuth G. Purchase and J.E. Williams. American Association of Avian Pathologists, College Station Texas. 129-135.
- Bell, I.G.; Nicholas, P.J.; Norman, C.; Cooper, K. and Cross, G.M. (1991a). The serological responses of chickens to mass vaccination with a live V4 Newcastle disease virus vaccine in the field and in the laboratory. 1 Meat chickens. *Aust. Vet. J.*, **68**: 85-89
- Bell, I.G.; Nicholas, P.J.; Norman, C; Cooper, K. and Cross, G.M., (1991b). The serological responses of chickens to mass vaccination with a live V4 Newcastle disease virus vaccine in the field and in the laboratory. 2. Layer pullets. *Aust. Vet. J.*, **68**: 90-92.
- Bell, J.G., Fotzo, T.M.; Amara, A. and Agbede, G., (1995). A field trial of the heat resistant V4 vaccine against Newcastle disease by eye-drop inoculation in village poultry in Cameroon. *Prev. Vet. Med.*, **25**:19-25.
- Copland, J.W., (1987). Newcastle disease in poultry a new food pellet vaccine. The Australian Centre for International Agriculture Research (ACIAR); Canberra, Monograph No. 5:7-9.
- Ezeokoli, D.C.; Umoh, J.U. Adesiyun, A.A. and Abdu, P.A, (1984). Prevalence of Newcastle disease virus antibodies in local and exotic chickens under different management systems in Nigeria. *Bull. Ani. Hlth. Prod. Afri.*, **32**:253-257.
- omwalk, N.E.; Adesiyun, J.T.; Bishu, G. and Adesiyun, A.A., (1985). A serological survey of Newcastle disease virus in domestic poultry around Zaria. *Nig. Vet. J.*, **14**(1):70.
- Ibrahim, A.L.; Chulan, U.; babjee, A.M., (1981). An assessment of the Australian V4 strain of Newcastle disease virus as a vaccine by spray, aerosol and drinking water administration. *Aust. Vet. J.*, **57**(6), 277-280.
- Ibrahim, M.A. and Abdu, P.A., (1992). Ethnoagroveterinary perspectives of poultry management, health and production among Hausa/Fulani of rural Nigeria In: *Proceedings of the 29th Annual General Meeting of Nigeria Veterinary Medical Association, Kaduna*. Pp. 172-181.
- Jarra, J., Ani, I., Schat, K.A. and Tourary, O., (1991). Vaccination of village chickens in Gambia against Newcastle disease using the heat resistant, food-pelleted V4 vaccine. *Avian Path.*, **20**:721-724.
- Jayawardane, G.W.L., Da Alwis, Mcl and Dawwda, B., (1990). Vaccination of chickens against Newcastle disease with V4 vaccine delivered on processed rice grains. *Aust. Vet. J.*, **67**(10): 364-366
- Nawathe, D.R.; Majiyagbe, K.A. and Ayoola, S.O., (1975). Characterization of Newcastle disease isolates from Nigeria. *Bull. Int. Epi.*, **83**: 11-12
- Nawathe, D.R., (1988). Newcastle Disease in Nigeria. National Veterinary Research Institute, Vom, Nigeria. In: *Viral Diseases of Animals in Africa* edited by Olufemi Williams and W.N. Masiga. Pp. 150-151

- Nwosu, I. and Okeke, S.T., (1989). The role of local chickens in the spread of Newcastle disease, in Nigeria. *Zariya Vet.*, **4**(1): 25-29.
- Okeke, A.N and Lamorde, A.G., (1988). Newcastle disease and its control in Nigeria. National Veterinary Research Institute, Vom, Nigeria In: *Viral Diseases of Animals in Africa. Olufemi Williams, A and W.N. Masiga (eds.)*, OAU/CTA Publication, Pp. 283-287.
- Olabode, A.O.; Lamorde, A.G.; Shidali, N.N.; Chukwuodo, A.A., (1992). Village chickens and Newcastle disease in Nigeria. *Australian Centre for International Agricultural Research Proceedings*, No. **39**. Pp. 159-160.
- Olabode, A.O. (1996). Research on NDV4 vaccine in Nigeria. Report submitted to Australian Centre for International Agricultural Research Proceedings at joint Mauritius and Common Wealth Veterinary Association held at Mauritius.
- Orajaka, L.J.E.; Adene, D.F.; Anene, B. M.; Onuoha, E.A. (1999). Seroprevalence of Newcastle disease in local chickens from Southeast derived savannah zone of Nigeria. *Revue Elev. Med. Vet. Pays. Trop*; **52** (3-4), 185-188.
- Oyewola, K.A., Ogundipe, G.A.T. and Durojaiye, O.A., (1996). Seroprevalence of Gumboro and Newcastle disease in local chickens in Ibadan, Nigeria. *Bull. Ani. Hlth. Prod. Afri.*, **34**:57-59
- Philips, J.M., (1973). Vaccination against Newcastle disease: An assessment of haemagglutination inhibition titres obtained from field sample. *Vet. Rec.*, **93**(22): 577-583.
- RIM, (1993). *Nigerian Livestock Reserve Resource Inventory and Management Report, Vol. I-IV*. Federal Department of Livestock and Pest Control Services, Federal Ministry of Agriculture and Rural Development, Garki, Abuja.
- Saglid, I.K. and Spalatin, J., (1982). Newcastle disease vaccination with the V4 strain in Malawi: Laboratory and field studies. *Avian Dis.*, **26**: 625-628.
- Shamaki, D.; Durojaiye, O.A. and Ojeh, C.K., (1989). Immunogenicity of Newcastle disease vaccine. *Zariya Vet.*, **11**(4): 19-24
- Spradbrow, P.B. (1988). Geographical distribution. In: *D.J. Alexander (editor). Newcastle Disease*. Kluwer Academic Publishers, Boston, MA. pp. 247-255.
- Spradbrow, P.B., (1992). A review of the use of food carriers for the delivery of oral Newcastle disease vaccine. In: *Spradbrow, P.B. ed. Proceedings of an International Workshop, Kuala Lumpur, 6-10 October, 1991*. Canberra, ACAIR Proceedings No. **39**, 18-20.
- Spradbrow, P.B., (1997). Policy framework for smallholder rural poultry development In: *Proceedings of International Workshop on Sustainable Poultry Production in Africa, Addis Ababa, Ethiopia, 1995*. Pp. 30-39.
- Steel, R.G.; Torrie, H.H., (1980). *Principles and Procedures of Statistics, 2nd ed.* New York, NY, USA. McGraw Hill Inc.
- Tu, T.D., Phuc, K.V., Dinh, N.T.K., Quoc, D.N. and Spradbrow, P.B., (1998). Vietnamese trials with a thermostable Newcastle disease vaccine (strain 1-2) in experimental and village chickens. *Prev. Vet. Med.*, **34**:205-214.
- Usman, M., (2002). Effects of vaccination of chickens against Newcastle disease with thermostable V4 and Lasota vaccines using different grains and their brans as vehicles. M.Sc. Thesis, Department of Veterinary Surgery and Medicine, Ahmadu Bello University, Zaria, Nigeria.
- Westbury, H.A. (1984). Comparison of the immunogenicity of NDV strain V4, B1 and LaSota in chickens. I Tests in susceptible chickens. *Aust Vet J*, **61**(2):2-9.