Evaluation of Antioxidant and Anti-amylase Activities of Sukhasarak Churna, an Ayurvedic Formulation

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INTRODUCTION

The use of Ayurvedic formulations has led to the sudden increase in the number of Ayurvedic drug manufactures due to the toxicity and side effects of allopathic medicines. Sukhasarak churna, an ayurvedic formulation, contains sonamukhi (Cassia angustifolia), ginger (Zingiber officinale), Haritaki (Emblica officinalis), Vidanga (Emblica ribes), Amla (Phyllanthus emblica) ajowan (Trachyspermum ammi) and liquorice (Glycyrrhiza glabra). Sukhasarak churna traditionally used as laxative, digestive, blood purifier and liver disorder.

Antioxidants are the substance that reduce oxidation and so counteract the reactive species. Reactive oxygen species (ROS) are major free radicals generated in many redox processes, which may induce oxidative damage to biomolecules, including carbohydrates, proteins, lipids, and DNA. Reactive oxygen species affect living cells, which mediate the pathogenesis of many chronic diseases, such as atherosclerosis, Parkinson’s disease, Alzheimer’s disease, stroke, arthritis, chronic inflammatory diseases, cancers, and other degenerative diseases (Mc Dermott, 2000). The action of ROS is opposed by a balanced system of antioxidant compounds produced in vivo (Halliwell and Gutteridge,1998). Endogenous antioxidants are insufficient, and dietary antioxidants are required to countermeasure excess ROS (Lim and Murtijaya, 2007).

Amylase inhibitors are also known as starch blockers because they contain substances that prevent dietary starches from being absorbed by the body. Starches are complex carbohydrates that cannot be absorbed unless they are first broken down by the digestive enzyme amylase and other secondary enzymes (Arribio Valenica et al., 2000).

More recent research utilizing purified amylase inhibitors have demonstrated that these antinutrients can rapidly inactivate amylase in human intestinal lumen in a dose dependent manner and post prandial rises in glucose and insulin (Marshall and Lauda, 1975). Essentially, it allows the carbohydrates to pass through the system possibly with less caloric intake.

Although the acute effects of α-amylase inhibitors may appear to have therapeutic benefit in patients suffering from diabetes mellitus, obesity and other diseases of insulin resistance, chronic administration in animal models has been shown to induce adverse effects including deleterious histological changes to the pancreas. The main of this study is to evaluate antioxidant and α-Amylase inhibition activities of Sukhasarak churna.

MATERIALS AND METHODS

Preparation of Churna and Extraction

Sukhasarak churna contents: Cassia angustifolia-40 gm, Zingiber officinale-20 gm, Emblica officinalis-20 gm, Emblica ribes -20 gm, Phyllanthus emblica-100 gm, Trachyspermum ammi -20 gm and Glycyrrhiza glabra-10 gm. were cleaned and dried material was made to fine
powder and passed through sieve no.100 and mixed geometrically and extracted with water and dried.

**Antioxidant Activity**

**DPPH Radical Scavenging Activity**

The DPPH assay measured hydrogen atom donating activity and hence provided an evaluation of antioxidant activity due to free radical scavenging. DPPH, a purple-colored stable free radical, was reduced into the yellow-colored diphenylpicryl hydrazine; the color intensity of the reaction mixture was measured spectrophotometrically at 510 nm (Layer et al., 1986). Briefly, 2.8 ml of DPPH (24 mg in 100 ml of ethanol) solution was added to various concentration of aqueous extract of leaves of Sukhasarak churna and incubated at 25 °C for 20 min. After incubation, the absorbance of the reaction mixture was read at 510 nm. Similarly, a control reaction was carried out without the Sukhasarak churna. The DPPH radical-scavenging activity was expressed as percentage inhibition and calculated according to the following equation:

% of Inhibition = \( \frac{(A_0 - A_1)}{A_0} \times 100 \)

Where \( A_0 \) was the absorbance of the control (without extract) and \( A_1 \) was the absorbance in the presence of the extract.

**Nitric Oxide Scavenging Activity**

In nitric oxide (NO) scavenging activity, the nitric oxide was liberated from sodium nitroprusside at physiological pH. This nitric oxide gets converted to nitrous acid and further forms nitrite ions (NO\(^2\)-), which may be quantified by the Griess reagent. 20 Sodium nitroprusside (10 mM, 50 µl) in phosphate buffer saline was incubated with 50 µl of various concentrations of aqueous extract of leaves of Sukhasarak churna at room temperature for 15 min. After incubation, 125 µl of Griess reagent was added and the reaction mixture was incubated for 10 min at room temperature. The color developed was measured at 546 nm (Vani et al., 1997). The nitric oxide radical scavenging activity was calculated according to the following equation:

% of Inhibition = \( \frac{(A_0 - A_1)}{A_0} \times 100 \)

**Ferric Reducing Power**

The ferric reducing power of the aqueous extract of Sukhasarak churna was determined by using potassium ferricyanide–ferric chloride method (Sreejayan and Rao, 1997). Different concentrations (100 –1000 µg/ml) of extracts were added to 2.5 ml 0.2 M phosphate buffer (pH 6.6) and 2.5 ml potassium ferricyanide (1%). The mixtures were incubated at 50°C for 20 min, after which 2.5 ml trichloroacetic acid (10%) was added. Two and one half milliliters of the mixture was taken and mixed with 2.5 ml water and 0.5 ml 1% FeCl\(_3\). The absorbance at 700 nm was measured after allowing the solution to stand for 30 min. A graph of absorbance vs. extract concentration was plotted to observe the reducing power.

**In Vitro Alpha Amylase Inhibition Assay**

The inhibition assay was performed according to Miller (1986) using DNS (3, 5-dinitrosalicylic acid method) method. Aqueous extract of leaves of Sukhasarak churna of varied concentrations in 500 µl were added to 500 µl of 0.02 M sodium phosphate buffer (pH 6.9 containing 6 mM sodium chloride) containing 0.04 units of alpha amylase solution and were incubated at 37°C for 10 min, followed by addition of 500 µl of a 1% starch solution in 0.02 M sodium phosphate buffer (pH6.9) all the test tubes. The reaction was stopped with 1.0 ml of 3, 5 DNSA reagent. The test tubes were then incubated in a boiling bath for 5 min, cooled to room temperature. The reaction mixture was then diluted after adding 10 ml distilled water and absorbance was measured at 540 nm. The control samples were also prepared accordingly without any plant extracts and were compared with the test samples containing various concentrations of the plant extracts prepared with different solvents. The results were expressed as % inhibition calculated using the formula:

% Inhibition activity = \( \frac{\text{Abs (Control)} - \text{Abs (Extract)}}{\text{Abs (Control)}} \times 100 \)

**RESULTS**

DPPH, which is a stable free radical with purple color, the intensity was measured at 510 nm spectrophotometrically to determine DPPH radical scavenging activity. The aqueous extract of Sukhasarak churna reduced DPPH in to 1,1-diphenyl-2-picryl hydrazine, a colorless compound with IC\(_{50}\) value of 521.31±2.3 µg/ml. Sodium nitroprusside in aqueous solution at physiological pH spontaneously generates nitric oxide, which interact with oxygen to produce nitrite ions. The aqueous extract of Sukhasarak churna had significantly scavenged the generated nitric oxide radicals with an IC\(_{50}\) value of 490.76±1.8 µg/ml (Table 1) Figure 1 showed the ferric reducing power of aqueous extract of Sukhasarak churna. Aqueous extract of Sukhasarak churna potent α-amylase inhibition and IC\(_{50}\) value was found to be 570.95±1.2 µg/ml.

**Table 1: Antioxidant activity aqueous extract of Sukhasarak churna (IC\(_{50}\) Value)**

<table>
<thead>
<tr>
<th>Treatment</th>
<th>DPPH Radical Scavenging assay</th>
<th>NO Radical Scavenging assay</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aqueous extract of Sukhasarak churna</td>
<td>521.31±2.3 µg/ml</td>
<td>490.76±1.8 µg/ml</td>
</tr>
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</table>

**Figure 1:** Ferric reducing power assay at various concentration of aqueous extract of Sukhasarak churna

**Table 2: In vitro α- Amylase inhibition of Sukhasarak churna**

<table>
<thead>
<tr>
<th>Treatment</th>
<th>IC(_{50}) value (µg/ml)</th>
</tr>
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DISCUSSION

In living systems, free radicals are constantly generated and they can cause extensive damage to tissue and biomolecules leading to various disease conditions, especially degenerative diseases, and extensive lysis (Oyaizu, 1986). Many synthetic drugs protect against oxidative damage but they have adverse side effects. An alternative solution to the problem is to consume natural antioxidants from food supplements and traditional medicines (Halliwell and Gutteridge, 1998; Yazdanparast and Ardestani, 2007). Recently, many natural antioxidants have been isolated from different plants materials (Yazdanparast et al., 2008; Packer and Ong, 2008).

The effect of antioxidants on DPPH is thought to be due to their hydrogen donating ability (Jovanovic and Simic, 2000). Free radicals are known to be a major factor in biological damages and DPPH has been used to evaluate the free radical scavenging activity of natural antioxidants (Baumann et al., 1979). DPPH, which is a radical itself with a purple color, changes in to a stable compound with a yellow color by reacting with an antioxidant and the extent of the reaction depends on the hydrogen donating ability of the antioxidant. The reduction capability of the DPPH radical is determined by the decrease in its absorbance at 510 nm, induced by antioxidants. In order to evaluate the antioxidant potency through free radical scavenging by the test sample, the change of optical density of DPPH radicals was monitored. Table No.1 shows the decrease in absorbance of DPPH radical due to the scavenging ability of aqueous extract of Sukhasarak churna. IC\textsubscript{50} value of aqueous extract on DPPH radical was 521.31±2.3.

It is well known that nitric oxide plays an important role in various inflammatory processes. Sustained levels of production of this radical are directly toxic to tissues and contribute to the vascular collapse associated with septic shock, whereas chronic expression of nitric oxide radical is associated with various carcinoma and inflammatory conditions including juvenile diabetes, multiple sclerosis, arthritis and ulcerative colitis (Zhu et al., 2001). The toxicity of NO increases greatly when it reacts with superoxide radical forming the highly reactive peroxynitrite anion (ONOO\textsuperscript{-}). The nitric oxide generated from sodium nitroprusside reacts with oxygen to form nitrite. The extract inhibits nitrite formation by directly competing with oxygen in the reaction with nitric oxide. IC\textsubscript{50} value of aqueous extract on nitric oxide scavenging was 490.76±1.8 (Table 1).

Ferric reduction is frequently used as a marker of electron-donating activity and considered as a significant mechanism of phenolic antioxidant action (Hatano et al., 2010). In the ferric reducing antioxidant power (FRAP) assay, the antioxidants in the extracts, if any, would result in reducing Fe\textsuperscript{3+} to Fe\textsuperscript{2+} by donating an electron. Amount of this Fe\textsuperscript{2+} complex can be determined at 700 nm by evaluating the formation of Perl's Prussian blue. The reductive ability is directly proportional with the absorbance of the APC at 700 nm (Nabavi et al., 2008). Figure 1 showed the ferric reducing power of aqueous extract of Sukhasarak churna.

Postprandial hyperglycemia is one of the risk factors associated with type 2 diabetes mellitus. Digestion of dietary starch α-amylase plays significant role in elevated blood glucose thus inhibition of amylase enzyme is very useful tool in management of hyperglycemia (Ebrahimzadeh et al., 2010). Drugs that inhibit carbohydrate hydrolyzing enzymes have been demonstrated to decrease postprandial hyperglycemia and improve impaired glucose metabolism without promoting insulin secretion of NIDDM patients. Sukhasarak churna showed potent α-Amylase inhibition 64.04 and 71.09 at 800 and 1000 µg/ml (Table 2). Natural health products of vegetable origin were clearly indicated as a promising avenue for the prevention of chronic diseases (Kwon et al., 2006).

Sukhasarak churna ingredients posses antidiabetic properties: Cassia angustifolia (Punitha and Manohar, 2006), Zingiber officinale (Yadav et al., 2010), Emblica officinalis (Ali et al., 2008), Ipomoea turpentine (Khan, 2009), Emblica ribes (Shankaraiah et al., 2012), Curcuma longa (Suresh Babu and Srinivas, 1997) and Glycyrrhiza glabra (Koga et al., 2004). Our study supports the antidiabetic activity of Sukhasarak churna.

CONCLUSION

Sukhasarak Churna, a well known ayurvedic formulation, exhibits antioxidant activity and α-Amylase inhibition activity under in vitro conditions. The studies are of great significance as the demand for herbal products as antioxidants and alpha amylase inhibitor is increasing constantly.

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Conflict of Interest

Authors declared no conflict of interest

REFERENCES


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