# Effects of diet supplementation with copper sulphate on growth performance and heamatological parameters of broiler chickens

<sup>1\*</sup>Unigwe, C.R., <sup>2</sup>Njoku, C. and <sup>3</sup>Orakwue, O.

<sup>1</sup>Department of Veterinary Biochemistry and Animal Production, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria

<sup>2</sup>Federal College of Agriculture, Ishiagu, Ebonyi State, Nigeria

<sup>3</sup>Federal College of Animal Health and Production Technology, Ibadan, Oyo State, Nigeria

\*Corresponding Author: robinsonunigwe@gmail.com Phone Number: +2348037707965

Target Audience: Animal scientists, Poultry farmers, Researchers and Veterinarians.

## Abstract

Diets of ninety-six day-old Abor-acre chicks were supplemented with different levels of copper sulphate  $(CuSO_4)$  to assess the growth performance and haematological parameters. The birds were conventionally brooded for two weeks after which they got allotted to;  $T_1$  (control),  $T_2$  (100 mg  $CuSO_4$  kg<sup>-1</sup>),  $T_3$  (200 mg  $CuSO_4$  kg<sup>-1</sup>), and  $T_4$  (300 mg  $CuSO_4$  kg<sup>-1</sup> of diet), in a completely randomized design. Each treatment had three replicates (n = 8). The supplements were given for 49 days during which feed intake and weekly weight gain were recorded. At the 49<sup>th</sup> day (63 day old), blood was aseptically collected via the wing vein using sterile syringe and needle for haematological studies. All data were subjected to analysis of variance and means separated using Duncan's New Multiple Range Test. The results showed that  $T_3$  had superior (p<0.05) weight gain and best FCR with enhanced feed intake similar (p>0.05) to  $T_1$  and  $T_2$  but differed (p<0.05) from  $T_4$  that had the worst growth performance. PCV, Hb and RBC of treated groups differed (p<0.05) from  $T_1$  that recorded the least haematological values. The WBC and its differentials did not statistically differ (p>0.05) except eosinophils where  $T_3$  spiked above others though statistically similar to  $T_1$  and  $T_2$  was the least. It could be concluded that  $CuSO_4$  supplementations at 100 and 200 mg kg<sup>-1</sup> diets had beneficial effects on growth performance whereas there was no detrimental effect of  $CuSO_4$  supplementations on all the haematological parameters of the broilers.

Keywords: Broiler chickens, Copper sulphate, growth, haematology

## **Description of Problem**

Food production in Nigeria has not kept pace with its population growth, because the population is growing at about 3.2% per annum while food production is at about 2.0% (1). The differences between the rate of food production and population growth has led to a food demand supply gap resulting in increased food importation and high rate of increase in food prices and ultimately wide spread hunger and malnutrition are evident in the country (2).

For many years, antibiotic growth promoters (AGP) have been incorporated into poultry diets because of their favorable effects on growth rate, and feed efficiency (3). However, there are increasing concerns that the frequent feeding of AGP is leading to the rise of antibiotic resistance in many pathogenic and opportunistic bacteria isolated from production animals as well as from humans (4). With the growing public concern of bacterial resistance to traditional antibiotics, poultry production industries need alternatives to orthodox antibiotics that could maintain intestinal health, have antimicrobial properties, and allow for optimal growth. Copper (Cu) has received great attention because of its antimicrobial properties that improve animal growth performance when fed over the minimum requirement (5; 6).

(7) recommended 8 mg Cu/kg of broiler diet as the minimum requirement and it has

been reported that its sulphate form ( $CuSO_4$ ) is more effective than other forms (8). Copper sulphate is the main Cu source in the diet of chickens and other animals; however, the inorganic salt shows poor bioavailability caused by the presence of ingredients that can inhibit absorption (9). However, the feed industry still prefers CuSO<sub>4</sub> for economic reasons. Cu is a key element required for animal growth and development of bones, connective tissue, the heart and several other organs (10). An excess of Cu may also have adverse effects on chicken performance (11). Others (11; 12) have also shown that supplemental copper has little or no effect on growth performance and feed utilization of boiler chickens. Copper is usually fed commercially at much higher pharmacological levels (100 to 300 mg/kg) because of its growth promoting properties, which is caused by its antibacterial properties (13).

Copper is needed for the development of antibodies and white blood cells, in addition to antioxidant enzyme production (14). Copper is also needed by animals to prevent microcytic hypochromic anaemia, thus, emphasizing its role in iron metabolism (15). In particular, feeding CuSO<sub>4</sub> at 200 mg/kg in white leghorn hens has shown a positive response, while levels of 400 mg/kg and above showed a progressively negative response (8). According to (16) broiler performance and intestinal physiology can be positively influenced by dietary Cu level (188 mg of Cu/kg of diet from CuSO<sub>4</sub> vs. no supplemental copper).

Many nutritionists and feed manufacturers are concerned about the contradictory reports on the growth promoting effect of different sources of supplemental copper. Therefore, the present study was carried out to evaluate the effects of supplemental  $CuSO_4$  on the growth performance and haematology of broiler chickens.

# Materials and Methods Site of the study and ethical consideration

The experiment was carried out at the Federal College of Animal Health and

Production Technology, Moor Plantation, Ibadan, Oyo State, Nigeria. Ibadan is located approximately on longitude 3° 5 to 4° 36' E and latitude 7° 23 to 7° 55' N (17). Ibadan has a tropical wet and dry climate, with a lengthy wet season. It has mean total rainfall of 9,233.60 mm, mean maximum and minimum temperatures of 39.82 °C and 22.5 °C respectively (18) and relative humidity of 74.55%. Ethical conditions governing the conduct of experiments with live animals were strictly observed as stipulated by (19). The experimental protocol was approved by the institutions ethical committee for the use of animals for experiment.

# Animals, feed, test ingredient, design and duration

Ninety-six (96) day-old Abor Acre broiler chicks were purchased from CHIMERO FARMS in Ibadan, for the experiment. Starter Top Feed® (3.200)Commercial ME/Kcal/Kg, CP = 22%), Finisher Top Feed<sup>®</sup> (2800 ME/Kcal/Kg, CP = 18%) and copper sulphate were also purchased from open market and used for the experiment. The treatments included  $T_1$  (control),  $T_2$  (100 mg CuSO<sub>4</sub>/kg of diet), T<sub>3</sub> (200 mg CuSO<sub>4</sub>/kg of diet) and  $T_4$  (300 mg CuSO<sub>4</sub>/kg of diet). Each treatment was replicated into three with 8 birds each. Birds were allotted to treatments using completely randomized design. The experiment lasted for 49 days.

# Management of the birds

On arrival, the day-old-chicks were provided with clean cool water mixed with multivitamins (Vitalyte<sup>®</sup>) and glucose to serve as anti-stress. Conventional brooding commenced during which they were served Starter plain feed (no CuSO<sub>4</sub>) and clean water ad libitum. Routine vaccinations (Newcastle disease [intra-ocular and lasota] and Infectious bursal disease vaccines) as prescribed by the Veterinarian were strictly followed. After two weeks of acclimatization. thev were weighed individually with sensitive scale (CAMRY,

### Unigwe et al

Model: EK5055, Max. 5kg, Min. 1g) and recorded and allotted to treatments and replicates as stated above. The Starter treatment diets (with CuSO<sub>4</sub>) commenced for another 2 weeks after which Finisher treatment diets (with CuSO<sub>4</sub>) were used for another 5 weeks. Therefore, they were on treatment diets for 49 days. The treatment diet and clean water ad libitum continued till the end of the study (at 63-day old). Standard management practices were followed throughout the experimental period.

#### **Data collection**

Feed intake and weekly weight gain were got by weigh-back mechanism. At the end of the study, blood was aspirated aseptically via the jugular vein into EDTA sample bottle for haematological analysis as described by (20).

#### Statistical analysis

All data obtained were subjected to analysis of variance (ANOVA) using a Statistical Package for Social Sciences (SPSS) version 20.0. Significantly different means were separated using Duncan's New Multiple Range Test (DNMRT) as described by (21).

Table 1: Growth	performance of b	oroilers fed	diets supplement	ed with copper sulphate

Parameters	T <sub>1 (control)</sub>	T <sub>2 (100mg)</sub>	T <sub>3 (200mg)</sub>	T <sub>4 (300mg)</sub>	±SEM
Initial weight (g)	233.33	231.20	232.40	232.77	0.67
Final weight (g)	2138.75 <sup>b</sup>	$2222.58^{ab}$	2309.17 <sup>a</sup>	1875.10 <sup>c</sup>	52.30
Total weight gain	1905.42 <sup>b</sup>	1991.38 <sup>ab</sup>	$2076.77^{a}$	1642.33 <sup>c</sup>	49.53
(g)					
Daily weight gain	38.89 <sup>b</sup>	$40.64^{ab}$	$42.38^{a}$	33.52 <sup>c</sup>	0.86
(g)				_	
Feed intake (g)	5344.91 <sup>a</sup>	5533.83 <sup>a</sup>	5476.13 <sup>a</sup>	4675 <sup>.</sup> 07 <sup>b</sup>	165.63
Daily feed intake	109.08 <sup>a</sup>	112.94 <sup>a</sup>	111.76 <sup>ª</sup>	95.41 <sup>b</sup>	2.25
(g)					
Feed conversion	2.81	2.78	2.64	2.85	0.05
ratio					

<sup>abc</sup>Means with different superscripts in the same row are significantly different (p<0.05)

## Results

# Growth performance and haematological parameters

Table 1 shows the growth performance of broilers fed diets supplemented with copper sulphate. There were significant effects (p<0.05) of CuSO<sub>4</sub> on growth performance indices (Final weight [FW], Daily weight gain [DWG], and Feed intake [FI]) except feed conversion ratio (FCR) of the chickens. The DWG of T<sub>3</sub> was superior and significantly different (p<0.05) from T<sub>1</sub> and T<sub>4</sub> but similar to T<sub>2</sub>. However, T<sub>1</sub> and T<sub>2</sub> equally outperformed (p<0.05) T<sub>4</sub>. With respect to FI, though T<sub>2</sub> consumed more than others, it did not statistically differ (p>0.05) from  $T_1$  and  $T_3$  but  $T_4$  (p<0.05). However, though there was no significant (p>0.05) effect of CuSO<sub>4</sub> on FCR of the chickens, numerically,  $T_3$  was the most efficient in converting feed to meat whereas  $T_4$  was the least.

Table 2 shows the haematological parameters of broilers fed diets supplemented with copper sulphate. Copper sulphate supplementation improved PCV, Hb and RBC significantly (p<0.05) when compared with the control group. There were no statistical differences (p>0.05) among the CuSO<sub>4</sub> treated groups in this regard. The WBC and the differentials had no significant difference

(p>0.05) among the treatments except for statistically differed (p<0.05) from other eosinophils that had T<sub>3</sub> as the highest and groups.

#### Unigwe et al

Table 2: Heamatological parameters of broilers fed diets supplemented with copper sulphate

Parameters	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	±SEM
PCV (%)	22.67 <sup>b</sup>	26.67ª	24.33 <sup>ab</sup>	26.00ª	0.56
Hb (g/dĹ)	7.23 <sup>b</sup>	8.63ª	8.27ª	8.67ª	0.19
RBC (x10 <sup>3</sup> /mm <sup>3</sup> )	1.84 <sup>b</sup>	2.87ª	2.11 <sup>ab</sup>	2.76ª	0.16
WBC (x10 <sup>3</sup> /mm <sup>3</sup> )	12.50	13.00	14.45	14.18	3.67
Platelet (x10 <sup>5</sup> /mm <sup>3</sup> )	967.00	1750.33	2010.00	1084.00	213.95
Lympho (x10 <sup>3</sup> /mm <sup>3</sup> )	67.67	63.67	62.00	67.67	1.54
Hetero (x10 <sup>3</sup> /mm <sup>3</sup> )	26.33	33.33	31.33	27.00	1.41
Mono (x10 <sup>3</sup> /mm <sup>3</sup> )	2.67	2.67	3.33	3.67	0.34
Eosino (x10 <sup>3</sup> /mm <sup>3</sup> )	2.67 <sup>ab</sup>	2.00 <sup>b</sup>	4.00ª	2.67 <sup>ab</sup>	0.32
Basophil (x10³/mḿ³)	0.33	0.67	0.33	0.33	0.15

<sup>ab</sup>Means with different superscripts in the same row are significantly different (p<0.05)

**Legend**: Lympho = lymphocyte, Hetero = heterocyte, Mono = monocyte, Eosino = eosinophil

## Discussion

Different sources such as Cu chloride, Cu oxide, Cu citrate, Cu sulphate and tribasic Cu chloride at different concentrations have been applied in poultry feed (22). However, the feed industry still prefers CuSO<sub>4</sub> for economic reasons. Cu is a key element required for animal growth and development of bones, connective tissue, the heart and several other organs (10). There was gradual improvement in growth indices as the CuSO<sub>4</sub> supplementation increased from 100 to 200 mg kg<sup>-1</sup> diet, but not with higher level (300 mg kg<sup>-1</sup> of diet). This result is in conformity with the reports of some authors (9; 23; 24; 25; 37) but contradicted the result of (26) probably because copper oxide nanoparticles (CuO-NPs) was used in lieu of CuSO<sub>4</sub>. (27) reported that Cu improved nitrogen retention whereas (9; 28) stated that the mechanism is by stimulating hormone and growth factors as well as increasing the availability of amino acids for absorption in the intestines. (7) recommended 8 mg/kg of Cu as the minimum requirement for broiler diets and it has been reported that its sulphate form is more effective than other forms (8). Copper (Cu) has antimicrobial

properties that improve animal growth performance and feed efficiency in broilers when provided at much higher pharmacological levels (5; 6; 29). One reason for this is that Cu plays an antioxidant role in the system of animals (11; 30) and has antibacterial properties (13; 31). In particular, feeding CuSO<sub>4</sub> at 200 mg/kg in white leghorn hens has shown a positive response, while levels of 400 mg/kg and above showed a progressively negative response (8). Addition of CuSO<sub>4</sub> could improve intestinal mucosal morphology, which may contribute in improving nutrient availability and is associated with increasing goblet cell numbers, total goblet cell area, goblet cell mean size, mucosal thickness and a greater number of segmented filamentous bacteria compared to control (32). (33) demonstrated that Cu might be involved in pituitary growth hormone gene expression and secretion of several neuropeptides in the hypothalamus. Supplementation of CuSO<sub>4</sub> to pig diets reduces the number of ureolytic bacteria as a group, of which Streptococci make up approximately 75%, inhibits the coliforms in the caecum and the colon, and reduces the number of Streptococci in fecal samples (22). The present results agree with the findings of (34), (35), (36) and (31) that reported that copper supplementation at the levels of 125 to 250 ppm improved growth and feed efficiency in broilers. However, contrary to the results of the present work, some authors (11; 32; 38; 39) have reported no positive effect of dietary copper

41, 42) reported that body weight gain were reduced significantly by the supplementation of Cu at 500 mg kg<sup>-1</sup> in the diet of chicken and pullets. demonstrated layer (43) that supranormal level of Cu gave different responses in different species. (44), (45) and (46) reported that Cu has antimicrobial actions thus has growth stimulating action, promotes hypothalamic appetite-regulating genes (47) and acts as a catalyst in enzyme systems within cells (48). It has also been demonstrated that intravenous injection of Cu stimulates growth of weanling pigs (33). Deficiency of Cu will certainly affect chicken growth, while an excess of Cu is not recommended because either it will be excreted or have an adverse effect on performance (9). Statistically, similar FCR in the present study agrees with the results of some authors (39; 49; 50) but contradicts those of others (51; 52). With 300 mg kg<sup>-1</sup> diet, the body weight gain and feed intake significantly decreased as well as higher FCR which is in tandem with the report of (53).

In supplemented birds, Cu plays a major role as cofactor in haematogenesis. Cu is one of the most critical trace elements in livestock because it is necessary for haemoglobin formation, iron absorption from GI-tract and iron mobilization from tissue stores (54). The birds in the present study did not show any haematological sign of excessiveness of Cu supplementation since even at 300 mg kg<sup>-1</sup> diet, the PCV, Hb and RBC increased beyond the control group. This is in consonance with the earlier reports of (24; 25; 55) where significant (p < 0.05) enhancement of PCV, Hb, and RBC counts were recorded in the

CuSO<sub>4</sub> group. Excess of dietary copper results in an accumulation of copper in the liver with decreased Hb concentration and packed cell volumes of blood (56). (57) reported similar finding in Wister albino rats. It can be inferred from the present study that Cu was not in excess in the diets with respect to heamatological parameters Additionally Unigwe et al

> or eryunocytes in the peripheral blood of the birds (37). The concentration of haemoglobin was increased, which allowed the birds to keep oxygen transport at an appropriate level (37; 58). The increased level of haemoglobin could be due to its continued synthesis by erythrocytes already circulating in the peripheral blood. It is also likely that the increased level of haemoglobin could be linked with the homeopathic function of copper. Cu to has been shown directly stimulate erythrocyte synthesis, as it determines iron absorption into the body and its incorporation in haemoglobin (59). With respect to WBC count, the marginal increase could be associated with the dietary CuSO<sub>4</sub> corroborating the findings of (27; 58) where there were increased counts of WBC after inovo injection of CuSO<sub>4</sub>. Monocyte count was enhanced in the present study which contradicted the results of (37) who reported reduced percentage of monocytes probably because they used heat stressed birds in their study. With the numerical enhancement of monocytes, it can be suggested that phagocytic and metabolic activities could be accelerated thereby conferring further protection upon the birds due to CuSO<sub>4</sub> supplementation.

# **Conclusion and Applications**

- 1. The results obtained in the present study revealed that supplementation of CuSO<sub>4</sub> at 100 and 200 mg kg<sup>-1</sup> of diets numerically increased feed intake and significantly enhanced growth performance of broiler chickens.
- 2. Contrary to the above, at a higher rate of 300 mg kg<sup>-1</sup> of diet, the performance

indices and feed intake declined significantly.

3. With respect to haematological parameters, even at 300 mg kg<sup>-1</sup> of diet, there was no deleterious effect on the parameters measured but rather significantly improved haemopoietic parameters of the birds

copper sulphate supplementation at the rates of 100 to 200 mg kg<sup>-1</sup> of diet improved the growth performance of broiler chickens and that the haematological parameters measured were enhanced even at 300 mg kg<sup>-1</sup> diet supplementation. Therefore, the use of CuSO<sub>4</sub> at 100 - 200 mg kg<sup>-1</sup> of diet is advocated.

## References

- 1. National Bureau of Statistics (NBS) (2011). Official website http://www. nigerianstat.gov.ng
- Ojo, S.O. (2003). Productivity and Technical Efficiency of Poultry Egg production in Nigeria. *International Journal of Poultry Science*, 2(6): 459-464.
- 3. Ghosh, T.K., Haldar, S., Bedford, M.R., Muthusami, N. and Samanta, I. (2012). Assessment of yeast cell wall as replacements for antibiotic growth promoters in broiler diets: effects on performance, intestinal histo-morphology and humoral immune responses. Journal Physiology of Animal and Animal Nutrition, 96: 275-284.
- 4. Kana, J.R., Teguia, A., Mungfu, B.M., Tchoumboue, J. (2011). Growth performance and carcass characteristics of broiler chickens fed diets supplemented with graded levels of charcoal from maize seed of Canarium cob or schweinfurthii Engl. Animal Tropical Health and Production, 43: 51-56.
- Lu, L. Wang, R.L., Zhang, Z.J., Steward, F.A., Luo, X. and Liu, B. (2010). Effect of dietary supplementation with copper sulphate or tribasic copper chloride on growth performance, liver copper

concentrations of broiler in floor pens, and stabilities of vitamin E and phytase in feeds. *Poultry Science*, 138: 181-189.

 Kim, G.B., Seo, Y.M., Shin, K.S., Rhee, A.R., Han, J. and Paik, I.K. (2011). Effects of supplemental copper-methionine chelate and copper-soy-proteinate on performance, blood parameters, liver

Unigwe et al

Poultry Research, 20: 21-32.

- NRC, (1994). Nutrient requirements of poultry, 9th Revise. Ed, National Academy Press, Washington, DC.
- Chiou, W.P., Chen, K,. Chiou, P.W.S., Chen, K.L. (1997). Toxicity, tissue accumulation and residue in egg and excreta of copper in laying hens. *Animal Feed Science and Technology*, 67: 49-60.
- Scott, A., Vadalasetty, K.P., Chwalibog, A. and Sawosz, E. (2018). "Copper nanoparticles as an alternative feed additive in poultry diet: a review". *Nanotechnology Reviews*, 7 (1): 69-93. https://doi.org/ 10.1515/ntrev-2017-0159
- 10. Hefnawy, A.E., and El-khaiat, H. (2015). Copper and animal health (importance, maternal fetal, immunity and DNA relationship, deficiency and toxicity). *International Journal for Agro Veterinary and Medical Sciences*, 9: 195– 211.
- 11. Karimi, A., Sadeghi, G. and Vaziry, A. (2011). The effect of copper in excess of the requirement during the starter period on subsequent performance of broilers. *Journal of Applied Poultry Research*, 20: 203-209.
- Xiang-Qi, Z. Zhang, K. Ding, X. and Boi, S. (2009). Effects of dietary supplementation with copper sulphate or tribasic copper chloride on carcass characteristics, tissular nutrients deposition and oxidation in broilers. *Pakistan Journal of Nutrition*, 8: 1114-1119.
- Payvastegan, S., Farhoomand, P., Shahrooze, R., Delfani, N. and Talatapeh, A. (2012). The effects of different levels

of canola meal and copper on performance, susceptibility to ascites and plasma enzyme activities in broiler chickens. *Annals of Biological Research*, 3: 5252-5258.

 Sharma, M.C., Joshi, C., Pathak, N.N. and Kaur, H. (2005). Copper status and enzyme hormone vitamin and immune Unigwe et al

*science*, *19*: 115–125. 10.1010/J.Ivsc. 2004.11.015

- Leeson, S. and Summers, J.D. (2001). Copper. In: Leeson S, Summers JD (eds.), Nutrition of the Chickens (4th ed.) University Books, P. O. Box 1326, Guelph, Ontario, Canada pp 394-397.
- Arias V.J., and Koutsos, E.A. (2006). Effects of copper source and level on intestinal physiology and growth of broiler chickens. *Poultry Science*, 85: 999-1007.
- 17. Oladele, B.M. and Oladimeji, B.H. (2011). Dynamics of urban land use changes with remote sensing: Case study of Ibadan, Nigeria. *Journal of Geography and Regional Planning*, 4 (11): 632-643.
- Egbinola, C.N. and Amobichukwu, A.C. (2013). Climate variation assessment based on rainfall and temperature in Ibadan, South-Western, Nigeria. *Journal of Environment and Earth Science*, 3 (11): 32-45.
- Ward, J.W. and Elsea, J.R. (1997). Animal case and use in drug fate and metabolism, methods and techniques. Vol. 2. New York: Marcel Deker; pp. 431.
- 20. Mafuvadze, B. and Erlwanger, K.H. (2007). The effect of EDTA, heparin and storage on the erythrocyte osmotic fragility, plasma osmolarity and haematocrit of adult ostriches (*Struthio camelus*). *Veterinarski Arhiv*, 77: 427-434.
- Obi, I.U. (2002). Statistical methods of detecting differences between treatment means and research methodology issues in laboratory and field experiments. AP Express Publishers, Limited, 3 Obollo road, Nsukka-Nigeria. Pp. 117.
- 22. Pang, Y., Patterson, J. A. and Applegate, T.J. (2009). The influence of copper

concentration and source on ileal microbiota. *Poultry Science*, 88: 586-592.

- 23. El-Ghalid, O.A.H., El-Ashry, G.M., Soliman, M.S. and Abd El-Hady, A.M. (2019). Effect of dietary sources and levels of copper supplementation on growth performance, blood parameters and slaughter traits of broiler chickens et al.
- 24. EI-Kazaz, S.E. and Hatez, M.H. (2020). Evaluation of copper nanoparticles and copper sulfate effect on immune status, behavior, and productive performance of broilers. *Journal of Advanced Veterinary and Animal Research*, 7(1): 16–25. doi: 10.5455/javar.2020.g388
- 25. El-Wardany I., Abdel-Hamid A.E., Morsi, A.M. and El-Naggar, A.S. (2020). Productive and immunological responses of broiler chicks to supplementation of different copper forms. *Arab University Journal of Agricultural Science*, 28(3): 928-948.
- Morsy, E.A., Hussien, A.M., Ibrahim, M.A., Farroh, K.Y. and Hassanen, E.I. (2021). Cytotoxicity and genotoxicity of copper oxide nanoparticles in chickens. *Biological Trace Element Research*, 199: 4731-4745.https://doi.org/10.1007/s12011-021-02595-4
- 27. Oguntoye, M.A., Akintunde, A.R., Ayoola, A.A., Ogundele, M.A. and Odetola, O.I. (2018). Haematological and biochemical responses of starter broiler chickens fed copper and probiotic supplemented diets. *Nigerian Journal of Animal Science*, 20 (1): 173-182.
- Braude, R. (1965). Copper as a growth stimulant in pigs. In Cuprum pro vita. Trans. Symp. Copper Development Association. London. pp. 55-57.
- Skrivan, M., Sevcikova, S., Tumova, E., Skrivanova, V. and Marounek, M. (2002). Effect of copper supplementation on performance of broiler chickens, cholesterol content and fatty acid profile of meat. *Czech Journal of Animal Science*, 47: 275-280.
- 30. Ajuwon, O.R., Idowu, O.M.O., Afolabi,

S.A., Kehinde, B.O., Oguntola, O.O., and Olatunbosun, K.O. (2011). The effects of dietary copper supplementation on oxidative and antioxidant systems in broiler chickens. *Archivos de Zootecnia*, 60: 275-282.

31. Pang, Y. and Applegate, T.J. (2007). Effects of dietary copper supplementation and copper source on digesta pH calcium 39 Unigwe et al

gasuonnesinai tract or oroner chicken. Poultry Science, 86: 531-537.

- 32. Payvastegan, S., Farhoomand, P. and Delfani, N. (2013). Growth performance, organ weights and, blood parameters of broilers fed diets containing graded levels of dietary canola meal and supplemental copper. *Journal of Poultry Science*, 50: 354-363.
- 33. Zhou, W., Kornegay, E.T., Lindemann, M.D., Swinkels, J.W., Welton, M.K. and Wong, E.A. (1994). Stimulation of growth by intravenous injection of copper in weanling pigs. *Journal of Animal Science*, 72: 2395-2403.
- 34. Choi, Y.J. and Paik, I.K. (1989). The Effect of Supplementing copper sulfate on the performance of broiler chicken. Korean. *Journal of Animal Nutrition and Feed*, 13: 193–200.
- 35. Baker, D.H., Odle, J., Funk, M.A. and Wieland, T.M. (1991). Bioavailability of copper in cupric oxide, cuprous and in a copper-lysine complex. *Poultry Science*, 70: 177–9.
- 36. Ward, T.L., Watkins, K.L., Southern, L.L. (1995). Interactive effects of dietary copper, water copper, and *Eimeria spp*. infection on growth, water intake, and plasma and liver copper concentrations of poults. *Poultry Science*, 74 (3): 502-509.
- 37. Nguyen, H.T.T., Morgan, N., Roberts, J.R., Swick, R.A. and Toghyani, M. (2020). Copper hydroxychloride is more efficacious than copper sulfate in chicken's improving broiler growth performance, both at nutritional and growth-promoting levels. Poultry Science, 99 (12): 6964-6973.

- 38. Torki, M., Kaviani, K and Ghasemi, H.A. (2014). Effects of diet supplementation by copper sulphate and ginger essential oil on growth performance and plasma biochemical parameters of broiler chickens under high environmental temperature conditions. *European Poultry Science*, 78. DOI: 10.1399/eps.2014.62
- 39 Vasanth S Dinu MT Mercy AD and al

Suprate versus Flavoniyem as a growin promoter in broiler chickens. *International Journal of Current Research*, 7 (5): 16536-16539.

- 40. Chen, K.L., Wu C.P. and Lu, J.J. (1996). Effects of dietary copper levels on performance, tissues and serochemistry value of Taiwan country chicken. *Journal* of Biomass Energy Soiety in China, 15: 70–5.
- 41. Chen, K.L., Chen, C.L., Lien, T.E. and Horng, Y.M. (1997a). Effect of dietary levels of copper on tissue residue and serochemistry value of growers. *Taiwan Journal of Veterinary Medicine and Animal Husbandry*, 67: 45–50.
- 42. Chen, K.L., Wu, C.P. and Chen, C.L. (1997b). Effects of dietary levels of copper on performance and intestinal structure in growers. *Journal of Chia-Yi Institute of Agricultural Technology*, 50: 31–9.
- 43. Paik, I.K., Seo, S.H., Um, J.S., Chang, M.B. and Lee, B.H. (1999). Effects of supplementary copper-chelate on the performance and cholesterol level in plasma and breast muscle of broiler chickens. *Asian-Australasian Journal of Animal Science*, 12: 794–8.
- 44. Vogt, H., Matthes, S. and Harnisch, S. (1981). Preservatives organic acids in broiler and laying rations. Conf. on Feed Additives, Budapest, Hungary.
- Burnell, T.W., Cromwell, G.L. and Stahly, T.S. (1988). Cited by J. Gohl, In: Bottom Line of Nutrition. Feed stuff, June 13, pp. 16–8.
- 46. Yang, W., Wang, J, and Liu, L. (2011). Effect of high dietary copper on somatostatin and growth hormonereleasing hormone levels in the

hypothalamic of growing pigs. *Biological Trace Element Research*, 143: 893–900. 10.1007/s12011-010-8904-x

- 47. Zhu, D., Yu, B., Ju, C., Mei, S. and Chen, D. (2011). Effect of high dietary copper on the expression of hypothalamic appetite regulators in weanling pigs. *Journal of Animal and Feed Sciences*, 20: 60–70. 10.22358/jafs/66158/2011
- 10 Durchanth I Vattanaani V Chittani D

review on role of essential trace elements in health and disease. *Journal of Dr. NTR University of Health Sciences*, 4: 75–85. 10.4103/2277-8632.158577

- 49. Ledoux, D.R., Miles, R.D., Ammerman, C.B. and Harms, R.H. (1987). Interaction of dietary nutrient concentration and supplemental copper on chick performance and tissue copper concentrations. *Poultry Science*, 66: 1379-1384.
- Wang, Z., Cerrate, S., Coto, C., Yan, F. and Waldroup, P.W. (2007). Evaluation of Mintrex copper as a source of copper in broiler diets. *International Journal of Poultry Science*, 6: 308-313.
- 51. Pesti, G.M., and Bakalli, R.I. (1996). Studies on the feeding of cupric sulfate pentahydrate and cupric citrate to broiler chickens. *Poultry Science*, 75: 1086–1091.
- 52. Samanta, B., Ghosh, P.R., Biswas, A. and Das, S.K. (2011). The effects of copper supplementation on the performance and hematological parameters of broiler chickens. Asian- Australian Journal of Animal Science, 7: 1001-1006.
- 53. Hashem, M.A., ABD El Hamied, S.S., Ahmed, E.M.A., Amer, S.A. and El-Sharnouby, M.E. (2021). Mitigating the Growth, Biochemical Changes, Genotoxic and Pathological Effects of Copper

Toxicity in Broiler Chickens by Supplementing Vitamins C and E. *Animals*, 11(6): 1811. https://doi.org/ 10.3390/ani11061811

54. Xia, M.S., Hu, C.H. and Xu, Z.R. (2004). Effect of copper bearing montmorillonite on growth performance, digestive enzyme activities and intestinal microflora and morphology of male broiler. *Poultry* Science, 82 (11): 1868–1875

Unigwe et al

U.O., Osinowo, O.A. and Ariyo, O.W. (2020). Haematological and serum biochemical indices of broiler chickens fed graded levels of inorganic and chelated trace minerals. *Nigerian Journal of Animal Production*, 47(2): 46 - 50.

- 56. Swensen, M.J. and Reece, W.O. (1996). In: Dukes Physiology of Domestic animal. 11<sup>th</sup> Edn. Panima Publishing Corporation, New Delhi, pp. 529-530.
- 57. Ozcelik, D., Toplan, S., Ozdemir, S. and Akyolan, M.C. (2002). Effects of excessive copper intake on haematological and haemorheological parameter. *Biological Trace Element Research*, 89 (8): 35-42.
- 58. Mroczek-Sosnowska, N., Batorska, M., Lukasiewicz, M., Wnuk, A., Sawosz, E., Jaworski, S. and Niemiec, J. (2013). Effect of nanoparticles of copper and copper sulfate administered in ovo on hematological and biochemical blood markers of broiler chickens. *Annals of Warsaw University of Life Sciences-SGGW, Animal Science*, 52: 141–149.
- Mullally, A.M., Vogelsang, G.B. and Moliterno, A.R. (2004). Warted sheep and premature infants the role of trace metals in hematopoiesis. *Blood Reviews*, 18: 227– 234. 10.1016/ S0268-960X(03)00067-5