

## Research Article

# Evaluation of Pharmaceutical and Microbial Qualities of Some Herbal Medicinal Products in South Western Nigeria

Adenike Okunlola, Babatunde A. Adewoyin and Oluwatoyin A. Odeku\*

Department of Pharmaceutics & Industrial Pharmacy, Faculty of Pharmacy, University of Ibadan, Ibadan, Nigeria.

## Abstract

**Purpose:** The aim of the present study was to investigate the pharmaceutical and microbial qualities of 21 different (of various dosage forms) Herbal Medicinal Products (HMPs) sourced from some traditional medicine sales outlets and retail pharmacy outlets in south western Nigeria.

**Method:** The pharmaceutical qualities evaluated include tablet crushing strength, friability, disintegration time; density of the solutions and suspensions; particle size and angle of repose of the powders. Phytochemical tests were carried out to assess the class of compounds present in the formulations and the microbial quality of the products was also evaluated.

**Results:** The results show that twelve (57.1%) of the products had their manufacturing and expiry dates stated, nine (42.9%) products have been registered by NAFDAC and ten (47.6%) did not have their content stated but had their therapeutic claims indicated on the container. The tablet formulation (Product A) showed acceptable crushing strength and friability but failed the test for disintegration time. The angle of repose of the powder dosage forms were considerably high showing that the powders were highly cohesive and not free flowing. The microbial load of the products varied considerably. Ten (47.6%) of the samples were contaminated by *E. coli*, seven (33%) were contaminated by *Salmonella*, fifteen (71.4%) were contaminated by *Staphylococcus aureus* and twelve (57.1%) were contaminated by fungi.

**Conclusion:** There is need for constant monitoring and control of the standards of herbal medicines available in the Nigerian market.

**Keywords:** Herbal medicinal products, microbial quality, pharmaceutical quality.

---

\*Correspondence: Email: [pejuodeku@yahoo.com](mailto:pejuodeku@yahoo.com) Tel 2348033235828; 23428106403

## INTRODUCTION

Herbal medicine, a form of complimentary and alternative medicine, is becoming increasingly popular in both developing and developed countries<sup>1</sup>. A World Health Organization (WHO) survey indicates that about 70-80% of the world population particularly in the developing countries rely on non-conventional medicines mainly of herbal sources in their primary healthcare<sup>2,3</sup>. WHO has described traditional medicine as one of the surest means to achieve total health care coverage of the world's population. In pursuance of its goal of providing accessible and culturally acceptable health care for the global population, WHO has encouraged the rational use of traditional plant based medicines by member states and has developed technical guidelines for the assessment of herbal medicine<sup>3,4</sup>.

In Nigeria, there appears to be an overwhelming increase in the public awareness and usage of herbal medicinal products in the treatment and/or prevention of diseases. This may not be unconnected to the active mass media advertisement embarked upon by the producers and marketers of the herbal medicinal products (HMPs) who have taken the advantage of the relatively high cost of the conventional pharmaceutical dosage forms, inaccessibility of the orthodox medical services to a vast majority of people particularly in the rural areas and the reservations by the public due to the prevalence of fake, substandard or counterfeit drugs in the market. These have placed the HMPs as a ready alternative to conventional dosage forms in the treatment of diseases. With this increased usage, the safety, efficacy and quality of these medicines have been an important concern for health authorities and health professionals. Although herbal remedies are often perceived as being natural and therefore safe, they are not free from adverse effects which may be due to factors such as adulteration, substitution, contamination, misidentification, lack of standardization, incorrect preparation and/or dosage, inappropriate labeling and/or advertisement<sup>5</sup>. In contrast to chemically defined

medicinal products, the biopharmaceutical quality and behavior of HMPs are often not well documented<sup>6</sup>. The WHO Good Manufacturing Practice Guidelines have provided technical guidelines to national regulatory authorities, scientific organizations, and manufacturers to undertake an assessment of the documentation/submission/dossiers in respect of herbal medicinal products. In Nigeria, the National Agency for Food Drug Administration and Control (NAFDAC) is responsible for drug administration and control of the quality of medicinal products including HMPs generally available in the market.

In an attempt to enhance the acceptability of the HMPs by consumers, many of the products have been formulated into conventional modern dosage forms such as tablets, capsules, suspensions, solutions and powders. With the prevalence in the Nigerian market of these products, it would be of great interest to evaluate the pharmaceutical qualities of these HMPs, irrespective of the medicinal content and therapeutic claims. Thus in the present study, the pharmaceutical and microbial qualities of 21 different brands of Herbal Medicinal Products (HMPs) sourced from various traditional medicine sales outlets and retail pharmacy outlets in south western Nigeria have been evaluated. Phytochemical tests were done to assess the class of compounds in the formulations and the microbial content was also evaluated.

## Materials and methods

### Materials

The following materials were used: Nutrient broth, Nutrient agar, MacConkey agar, Salt nutrient agar, Cetrimide Nutrient agar, Sabouraud dextrose agar were all obtained from Oxoid (Oxoid Products, Basingstoke, UK). Twenty one (21) different Herbal Medicinal Products (HMPs) were sourced from various traditional medicine sales outlets and retail pharmacy outlets in south western Nigeria. The type of dosage form, manufacture and expiry dates and the therapeutic indications are presented in Table 1.

**Table1:** Product code, Dosage form, Content, NAFDAC registration, Country of Origin, Manufacture and Expiry dates of 21 brands of HMPs from the Nigerian Market

Product code	Dosage form	Date of manufacture	Expiry Date	Country of Manu.	NAFDAC Reg.	Contents	Therapeutic claim
A	Tablet	Aug 2003	Aug 2007	US	No	Gingko biloba extract, Ganoderma lucidum, Schisandra chinensis	Brain fatigue, improvement of thought process.
B	Capsule	-	-	Nigeria	No	-	Hypertension, Diabetes, Yellow fever
C	Capsule	Jul 2004	Jun 2007	Nigeria	No	Strophanthus sarmentosus 75%, Pyrenanacantha 25%	Chronic fever, Cough
D	Capsule	Oct 2002	Oct 2005	Nigeria	Yes	Hibiscus sabdariffa 80mg, Sorghum bicolor 60mg, Gongronema latifolium 35mg	Energizer, Immune booster, Blood normalizer
E	Suspension	Oct 2002	Oct 2005	Nigeria	Yes	Hibiscus sabdariffa, Sorghum bicolor, Gongronema latifolium	Immune booster, Energizer, Blood normalizer
F	Solution	Aug 2004	Aug 2006	Nigeria	Yes	Enantia chloranthia, Mormodica charanthia, Citrus aurontifolia, Morinda lucida, Azaridachta indica, Cocos nucifera, Occimum gratissimum, Ananas comosus	Antimalarial.
G	Solution	Oct 2004	Oct 2006	Nigeria	Yes	Citrus aurantifolia, Lawsonia inermis, Azaridachta indica, Morinda lucida, Magnifea indica, Ficus capensis, Shonocentrum jollyanum, Theobroma cacao, Nuclea latifolia	Antimalarial
H	Solution	-	-	Nigeria	No	-	Antimalarial
I	Solution	-	-	Nigeria	No	-	Pruritis, Rashes, skin allergies.

Brand code	Dosage form	Date of manufacture	Expiry Date	Country of Manufacture	NAFDAC Registration	Contents	Therapeutic claim
J	Solution	-	-	Nigeria	No	-	Pile, dysentery, other stomach ailments
K	Solution	-	-	Nigeria	No	-	Eye ache, Stomach ulcer, waist pain, menstrual pain.
L	Solution	Jan 2004	Dec 2006	Nigeria	No	-	Heamorrhoids, pile.
M	Solution	Sept 2004	Sept 2007	Nigeria	Yes	Allium ascalonicum, Strophanthus hispidus, Chrysophyllum albidum, Nicotiana rustica, Pseuphatil kotsnyi	Gripe pain, teething problem, measles management
N	Solution	-	-	Nigeria	Yes	Allium sativa, Xylopia aromatica, Tetraptera tetraptera, Ficus carica, Nauclea latifolia, Combretum micrathum, Sterculia ureus.	Fever, measles, skin rashes and small pox.
O	Solution	Aug 2004	Jul 2007	Nigeria	Yes	Alstonia bonei, Anthoderta nobilis, Pandiaka involucrate, Citrus medica, Var acida, Phyllanthus reticulates, Lawsonia inermis, Ananaas comosus, Gossypium barbadense, Azaridachta indica, Zingiber officinale, Shaenocentrum jollyanum, Sacharinum officinum	Antimalarial
P	Solution	Aug 2004	Aug 2006	Nigeria	Yes	Xylopia aethopica, Citrus medica, Var acida, Cassia alata, Kigella Africana bark, Pterygota macrocana, Sacchatrium officinum	Menstrual disorder, pain and internal heat during menses
Q	Solution	Aug 2004	Aug 2006	Nigeria	Yes	Alstonia congensis, Opuntia spp, Daniella oleveri, Eugenia aromatica, Butyrospermum, Olax subscorpiodea, makamia tormentosa	Eczema, body rashes, nettlerashes, ringworm, acne.
R	Powder	-	-	Nigeria	No	-	Heamorrhoids
S	Powder	-	-	Nigeria	No	-	Typhoid fever
T	Powder	-	-	Nigeria	No	-	Diabetes
U	Powder	-	-	Nigeria	No	-	Pile

- = Not stated/available

#### **Determination of table/capsule properties:**

Twenty (20) tablets/capsules were weighed individually using an electronic balance (Mettler PE360 Deltarange, Switzerland) and the mean weight was calculated.

The crushing strength of the tablets was determined at room temperature using a Pfizer hardness tester (Pfizer Inc., Groton, CT). The mean crushing strength was calculated. The percent friability of the tablets was determined using a friabilator (Erweka Apparatebau, Germany) operated at 25rpm for 4 min.

The disintegration times, DT, of the tablets/capsules were determined in distilled water at  $37 \pm 0.5^\circ\text{C}$  using an Erweka disintegration tester (Erweka Apparatebau, Germany). All measurements were made in quadruplicate, and the results given are the means of four determinations.

#### **Determination of powder properties**

The particle size distribution and shape of the HMPs in powder dosage form were determined by optical microscopy on approximately 300 particles per sample.

The flow properties of the powders were evaluated by measuring the angle of repose. 5g of each powder was poured into a cylindrical glass fixed to a flat base of diameter 28mm. The cylinder was slowly pulled out vertically so as to form a cone of powder on the base. The height of the cone was measured and the angle of repose,  $\theta$ , was calculated using the equation:

$$\text{Tan } \theta = \frac{h}{r} \dots \dots \dots (1)$$

where **h** is the height of the conical powder heap, and **r** is the radius of the circular base. Determinations were done in triplicates.

#### **Determination of solutions properties**

The density of the solutions and suspension were determined by measuring the weight of 1ml of the sample. The density is taken as the mass per unit volume of the preparation.

#### **Phytochemical tests**

The following phytochemical tests were carried out on the products: Tests for alkaloids was performed using Wagner and Dragendoff's reagent<sup>7</sup>. 0.5 g of the product was added to 5ml of 1% aqueous HCl on a steam bath. This was filtered and 1 ml of the filtrate treated with a few drops of Dragendoff's reagent and another 1 ml portion treated similarly with Wagner's reagent. The formation of precipitates was an indication of the presence of alkaloids. The blood hemolysis test was used to tests for Saponins<sup>7</sup>. The test for anthraquinone was done by shaking 0.5 g of the product with 5 ml of chloroform for 5 min. The extract was filtered, and the filtrate shaken with an equal volume of 10% ammonia solution. A pink, violet or red colour in the ammoniacal layer (Lower layer) indicated the presence of free anthraquinones. The test for tannins was done using the ferric chloride test. A deep green colouration showed the presence of tannins<sup>8</sup>. The Keller- Kiliani test was used to test for the presence of cardenolides<sup>7,8</sup>. 0.5g of the product was dissolved in 2 ml of glacial acetic acid containing one drop of ferric chloride solution. This was then underlayered with 1 ml of concentrated sulphuric acid. A brown ring obtained at the interphase indicated the presence of a deoxy sugar typical of cardenolides<sup>8</sup>.

#### **Microbial content determination:**

For solid samples i.e. tablets and powders, 1g quantities were disintegrated in 9 mL of sterile distilled water while for the liquid formulations, 1ml quantities was dissolved/suspended in 9ml of sterile distilled water. Serial dilutions were made and viability assessed using the pour plate method. The plates were incubated at  $37^\circ\text{C}$  for 24h. The plate was placed on a colony counter and the number of colony forming units was taken. The microbial content was taken as the mean of duplicate determinations. The media utilized were Nutrient agar, Cetrimide Nutrient agar, Salt Nutrient agar, MacConkey agar. For detection of fungal growth in the samples, Sabouraud dextrose agar was poured into the plate and allowed to set and 1ml aliquot of each sample was spread on the surface and the plates were incubated at  $27^\circ\text{C}$  for 72 h.

**Table 2:** Physicochemical properties of the HMPs

Product code	Dosage form	Weight uniformity (mg)	Crushing Strength (N)	Friability (%)	Disintegration time (min)	Weight/ml (g/ml)	Mean particle size $\mu\text{m}$	Angle of repose	Bulk density ( $\text{g}/\text{cm}^3$ )
A	Tablet	548.0 $\pm$ 0.5	137.2 $\pm$ 19.6	0.8 $\pm$ 0.0	120.0 $\pm$ 3.0	-	-	-	-
B	Capsule	500.0 $\pm$ 0.5	-	-	30.0 $\pm$ 4.5	-	-	-	-
C	Capsule	380.0 $\pm$ 0.3	-	-	39.0 $\pm$ 2.5	-	-	-	-
D	Capsule	255.0 $\pm$ 0.3	-	-	54.0 $\pm$ 4.0	-	-	-	-
E	Suspension	-	-	-	-	1.256 $\pm$ 0.012	-	-	-
F	Solution	-	-	-	-	1.024 $\pm$ 0.023	-	-	-
G	Solution	-	-	-	-	1.010 $\pm$ 0.032	-	-	-
H	Solution	-	-	-	-	1.017 $\pm$ 0.046	-	-	-
I	Solution	-	-	-	-	1.019 $\pm$ 0.101	-	-	-
J	Solution	-	-	-	-	1.037 $\pm$ 0.024	-	-	-
K	Solution	-	-	-	-	1.006 $\pm$ 0.034	-	-	-
L	Solution	-	-	-	-	1.010 $\pm$ 0.058	-	-	-
M	Solution	-	-	-	-	1.012 $\pm$ 0.014	-	-	-
N	Solution	-	-	-	-	1.002 $\pm$ 0.008	-	-	-
O	Solution	-	-	-	-	1.039 $\pm$ 0.012	-	-	-
P	Solution	-	-	-	-	0.990 $\pm$ 0.009	-	-	-
Q	Solution	-	-	-	-	1.037 $\pm$ 0.056	-	-	-
R	Powder	-	-	-	-	-	215.56 $\pm$ 5.90	52.6 $\pm$ 1.2	0.232 $\pm$ 0.002
S	Powder	-	-	-	-	-	2.87 $\pm$ 0.98	61.9 $\pm$ 2.6	0.427 $\pm$ 0.026
T	Powder	-	-	-	-	-	98.70 $\pm$ 2.67	60.0 $\pm$ 0.9	0.368 $\pm$ 0.042
U	Powder	-	-	-	-	-	13.38 $\pm$ 1.12	66.5 $\pm$ 1.8	0.386 $\pm$ 0.008

The viable aerobic bacterial count and the viable count for moulds (dry surface method) and the absence (or presence) of *Escherichia coli* and *Staphylococcus aureus* were assessed using well established methods<sup>9,10</sup>.

### Results and discussion

The herbal medicinal products selected for this study consisted of one (1) tablet, one (1) suspension, three (3) capsule, four (4) powders and twelve (12) liquid dosage forms. All the samples, except for those samples without the manufacture or expiry dates indicated, were within their shelf life at the time of investigation. Twelve (57.1%) of the products had their manufacturing and expiry dates stated and only nine (42.9%) products have been registered by NAFDAC. This is contrary to the prohibition of the manufacture, advertisement, sale and distribution of herbal medicinal products in

Nigeria without proper registration by NAFDAC<sup>11</sup>. The European Agency for the Evaluation of Medicinal Products, EMEA and WHO have stated that the quantity of the herbal drug should be given as a range corresponding to a defined quantity of constituent with known therapeutic activity and if constituent(s) responsible for the therapeutic activity are unknown, the quantity of the whole herbal drug preparation should be given<sup>6,12</sup>. Furthermore, the dosage form, therapeutic indications and expiry dates should be stated. However, ten (47.6%) of the products did not have their content stated even though their therapeutic claims were indicated either on the container or in the leaflet insert.

The physicochemical properties of the product formulations are presented in Table 2. The European Pharmacopoeia have stated that

**Table 3:** Phytochemical analysis of the HMPs

Product code	Phytochemical tests				
	Alkaloids	Anthraquinones	Saponins	Tannins	cardenolides
A	+	-	-	+	-
B	-	+	+	+	+
C	+	-	+	+	-
D	-	-	+	-	-
E	+	-	+	-	-
F	-	-	+	+	+
G	-	-	+	+	-
H	-	-	+	+	-
I					
J	-	+	-	-	-
K	+	-	-	-	-
L	+	-	-	-	-
M	-	-	+	+	-
N	-	-	+	+	+
O	-	-	+	-	-
P	-	-	+	-	+
Q	-	-	+	-	+
R	+	-	-	+	-
S	-	-	+	+	-
T	+	-	+	+	+
U	+	+	+	+	-

Key: - = Absent  
+ = Present

tablets  $\geq 250\text{mg}$  may not deviate from the average weight by more than 5%w/w and capsules of  $\geq 300\text{mg}$  may not deviate by more than 7.5%w/w<sup>13</sup>. The results of the weight uniformity test for the tablets and capsules were within the acceptable pharmacopoeial limits. The tablet formulation (Product A) showed acceptable crushing strength and friability indicating the ability to resist chipping and abrasion or breakage under conditions of storage, transport and handling. However, the tablet had disintegration time of 120 min. This is considerably higher than the requirement for immediate release tablets (i.e. disintegration within 15minutes). This may result in delayed release of the active constituent and subsequent onset of action. In many cases, a complete and rapid disintegration/dissolution of the herbal drug preparation for example extracts, from the solid oral dosage formulation is a prerequisite for non-

problematic bioavailability and clinical efficacy of the HMPs. Some preparations such as lipid extracts and essential oils however are not easily soluble and do not dissolve completely from the pharmaceutical formulation<sup>6</sup>. Such HMPs may require other excipients such as disintegrants to aid the release of the active constituent from the dosage form. The disintegration times of the capsules (Products B – D) also varied considerably. Products C and D did not pass the pharmacopoeial test for disintegration of capsules (i.e. disintegration within 30minutes). The weight per mL of the solution was used as a means of assessing the uniformity of weight per dose of the product dispensed. The solutions (Products F - Q) generally contained about 1g per mL of the solution and the standard deviation showed that the variation in weight was generally  $\leq 5\%$ . The angle of repose of the powders (Products R - U)

**Table 4:** Microbial content of HMPs

Product code	Viable count (cfu/ml or g)					
	Aerobic organisms	<i>Staph. aureus</i>	<i>E.coli</i>	<i>Salmonella</i>	<i>Pseudomonads</i>	Fungi
A	-	-	-	-	-	4.0 x 10 <sup>3</sup>
B	5.0 x 10 <sup>3</sup>	1.4 x 10 <sup>4</sup>	7.0 x 10 <sup>3</sup>	-	-	-
C	7.5 x 10 <sup>3</sup>	2.0 x 10 <sup>3</sup>	3.0 x 10 <sup>3</sup>	-	-	-
D	3.0 x 10 <sup>4</sup>	7.0 x 10 <sup>3</sup>	13.0 x 10 <sup>3</sup>	3.0 x 10 <sup>3</sup>	-	-
E	2.0 x 10 <sup>4</sup>	3.0 x 10 <sup>4</sup>	7.0 x 10 <sup>3</sup>	1.0 x 10 <sup>3</sup>	-	3.0 x 10 <sup>4</sup>
F	1.0 x 10 <sup>3</sup>	1.0 x 10 <sup>3</sup>	-	-	-	-
G	5.0 x 10 <sup>2</sup>	-	-	-	-	-
H	5.0 x 10 <sup>3</sup>	1.0 x 10 <sup>3</sup>	-	-	-	-
I	5.0 x 10 <sup>3</sup>	2.0 x 10 <sup>3</sup>	1.5 x 10 <sup>2</sup>	1.0 x 10 <sup>3</sup>	-	2.0 x 10 <sup>3</sup>
J	1.5 x 10 <sup>2</sup>	5.0 x 10 <sup>2</sup>	-	-	-	-
K	7.5 x 10 <sup>3</sup>	1.0 x 10 <sup>3</sup>	-	-	-	-
L	5.0 x 10 <sup>3</sup>	-	-	-	-	-
M	-	-	-	-	-	3.0 x 10 <sup>3</sup>
N	2.0 x 10 <sup>4</sup>	4.0 x 10 <sup>3</sup>	1.5 x 10 <sup>4</sup>	-	-	1.2 x 10 <sup>4</sup>
O	-	-	-	-	-	3.5 x 10 <sup>3</sup>
P	-	-	-	-	-	3.0 x 10 <sup>3</sup>
Q	1.0 x 10 <sup>4</sup>	5.0 x 10 <sup>3</sup>	-	-	-	3.5 x 10 <sup>3</sup>
R	2.2 x 10 <sup>4</sup>	1.2 x 10 <sup>4</sup>	9.0 x 10 <sup>3</sup>	3.0 x 10 <sup>2</sup>	-	1.2 x 10 <sup>4</sup>
S	8.0 x 10 <sup>3</sup>	6.0 x 10 <sup>3</sup>	5.0 x 10 <sup>3</sup>	2.0 x 10 <sup>2</sup>	-	7.0 x 10 <sup>3</sup>
T	5.0 x 10 <sup>3</sup>	3.0 x 10 <sup>3</sup>	2.0 x 10 <sup>3</sup>	1.5 x 10 <sup>2</sup>	-	6.0 x 10 <sup>3</sup>
U	8.0 x 10 <sup>3</sup>	2.0 x 10 <sup>3</sup>	3.0 x 10 <sup>3</sup>	2.0 x 10 <sup>2</sup>	-	3.5 x 10 <sup>3</sup>

**Key:** - = No growth  
Cfu/ml = Colony forming unit per ml or g

was generally high showing that the powders were highly cohesive and are not free-flowing. This may result in the non-uniformity of dose dispensed from the container during use.

In contrast to orthodox medicinal products, the compositions of herbal drug preparations are determined by the practitioner, manufacturing process, and the intended dosage form herbal drug<sup>6</sup>. The results of the phytochemical tests are presented in Table 3. The results showed that the products contain alkaloids, anthraquinones, tannins, saponins and cardinolides. It is well known that HMPs usually consists of more than one plant or active constituents and their

therapeutic efficacy is not provided by a single group of compounds. Some of these compounds act synergistically to modify the bioavailability and efficacy of the active constituent. Furthermore, the constituent responsible for the claimed therapeutic effects are often unknown or only partly explained and thus precludes the level of control which could be routinely achieved with synthetic drug substances in conventional pharmaceuticals<sup>6</sup>. Moreover, the composition of an herbal drug preparation is determined by the practitioner who should be able to identify the correct plant species. Thus, misidentification of the plant is possible and the plants may contain potentially toxic constituents. As a result, there is



no guarantee of the authenticity and quantity of plant material used in the preparations and thus the quality of traditional medicines so produced varies widely and may not even be effective. Furthermore, herbal medicines, particularly those grown as cultivated crops have been shown to be contaminated by pesticides, fumigants, toxic metals and endotoxins<sup>13</sup>. Therefore, there is a need to select proper and appropriate technologies for the industrial production of traditional medicines such that the effectiveness of the preparation is ensured.

Herbal medicinal products usually contain bacteria and moulds from soil and atmosphere. The limits of microbial contamination are: total aerobic bacteria  $10^5$ cfu/g, yeasts and moulds  $10^3$  cfu/g, *Enterobacteria* and other Gram negative organisms  $10^3$ cfu/g and *E. coli* and *Salmonella* should be absent<sup>14</sup>. The microbial nature and content of the microbial contaminant in the HMPs are presented in Table 4. The results show that the microbial load of the products varied considerably. The samples were contaminated to varying degrees with bacteria and Fungi. Of concern is the level of contamination of the products by Gram negative organisms which are considered pathogenic. While none of the samples contained *Pseudomonas aeruginosa*, which is primarily a soil bacterium and the cause of various infections, e.g. of burns, and the urinary and respiratory tracts, ten (47.6%) of the samples were contaminated by *E.coli*, which is an intestinal bacterium and is an indicator for contamination by faeces and seven (33%) were contaminated with *Salmonella*. Fifteen (71.4%) of the products were contaminated by *Staphylococcus aureus* and twelve (57.1%) were contaminated by fungi. Soil, harvesting, drying, storage conditions and improper handling influence the microbiological quality of herbal drugs. The presence of microbial contaminant in non sterile pharmaceutical products can reduce or even inactivate the therapeutic activity of the products and has the potential to adversely affect patients taking the medicines<sup>15</sup>. Some infectious outbreaks have been associated with the use of heavily contaminated raw materials of natural origin<sup>16</sup>. The microbial quality of

pharmaceuticals is influenced by the environment and quality of the raw materials used during formulation. Thus manufacturers should ensure the lowest possible level of microorganisms in the raw material, finished dosage forms and the packaging components to maintain appropriate quality, safety and efficacy of the products.

The results of the present work show that Products A to D, which were among the products which did not show acceptable pharmaceutical and microbial qualities were expensive, popular, widely advertised and used in south western Nigeria for the treatment of various conditions. There is, therefore, the need for constant monitoring and quality control of herbal medicinal products manufactured, sold, advertised and used in Nigeria. The quality requirements for orthodox drug preparations are stringent in terms of content of active principles and toxic materials. Whereas the production of traditional medicines for local use does not require such stringent standards, HMPs are usually an improved version of the already produced medicines using traditional methods. As herbal medicinal products are complex mixtures which originate from biological sources, great efforts are necessary to guarantee a constant and adequate quality. By carefully selecting the plant material and a standardized manufacturing process, the pattern and concentration of constituents of herbal medicinal products should be kept as constant as possible as this is a prerequisite for reproducible therapeutic results<sup>6</sup>. Quality has to be built into the whole process beginning from the selection of propagation material to the final product reaching the consumer. Thus, there is need for constant monitoring and control of the standards of herbal medicines available in the market.

## REFERENCES

1. Eisenberg DM, Davis RB, Ettner SL, Appel S, Wilkey S, Van Rompay M, Kessler RC Trends in alternative medicine use in the United States, 1990-1997: Results of a follow-up national survey. *J. Amer. Med. Assoc.* 1998; 280: 1569-1575.

2. Akerele, O. Nature's medicinal bounty: don't throw it away. *World Health Forum*. 1993; 14: 390-395.
3. WHO, Regulatory situation of herbal medicine: A world wide review. World Health Organization, Geneva. 1998.
4. WHO, General guidelines for methodologies on research and evaluation of traditional medicines. World Health Organization, Geneva. 2000.
5. Lau A, Holmes MJ, Woo S and Koh H. Analysis of adulterants in a traditional herbal medicinal product using liquid chromatography-mass spectroscopy. *J. Pharm. Bio. Ana.* 2003; 31: 401-406.
6. The European Agency for the Evaluation of Medicinal Products, EMEA Working Party on Herbal Medicinal Products: Points to consider on the biopharmaceutical characterization of Herbal Medicinal Products. EMEA/HMPWP/344/03 (2003) <http://www.emea.eu.int/pdfs/human/hmpc/034403en.pdf> Last accessed on October 10, 2006.
7. Sofowora, E.A. *Medical Plant and Traditional Medicine in Africa*. University of Ife Press, Nigeria: 1994; pp. 1 – 23.
8. Evans WC: *Trease and Evans Pharmacognosy*. 14th edition. WB Saunders Ltd. London; 1996; pp.119-159.
9. Van Doorne H. and Claushaus EPM. The quantitative determination of Enterobacteriaceae in pharmaceutical preparations. *Int. J. Pharm.* 1979; 4: 119 - 125.
10. Waterman RF, Sumner ED, Baldwin JN and Warren FW. Survival of *Staphylococcus aureus* on pharmaceutical oral solid dosage forms. *J. Pharm. Sci.* 1973; 62: 1317 - 1320.
11. Herbal Medicines and Related Products (Registration) Regulations (2004) [www.nafdacnigeria.org/newregs/regulations.html](http://www.nafdacnigeria.org/newregs/regulations.html) Last accessed on October 10, 2006
12. WHO Expert Committee on Specifications for Pharmaceutical Preparations. Thirty-fourth report. Geneva, World Health Organization, 1996 (WHO Technical Report Series No. 863, thirty-fourth report, pp.178-184).
13. Chan K. Some aspects of toxic contaminants in herbal medicines. *Chemosphere* 2003; 52; 1361-1371.
14. European Pharmacopoeia: Directorate for the Quality of Medicines of the Council of Europe, 5<sup>th</sup> Edition. Strasbourg, France. (2007)
15. Nakajima K, Nonaka K, Yamamoto K, Yamaguchi N, Tani K and Nasu M. Rapid monitoring of microbial contamination on herbal medicines by fluorescent staining method. *Lett. Applied Microbiol.* 2005; 40 (2): 128-132
16. Kallings LO, Silver Stolpe L and Ernerfeldt F. Microbiological contamination of medical preparations. *Act. Pharm. Suec.* 1966; 3: 219 - 228.