5,7-Dimethoxycoumarin inhibits neuronal apoptosis by targeting caspase-3 expression in rats with acute cerebral infarction

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Abstract

Purpose: To investigate the effect of 5,7-dimethoxycoumarin on inhibition of neuronal apoptosis in a rat model of acute cerebral infarction.

Methods: Occlusion of middle cerebral artery was used for the preparation of a rat model of cerebral infarction. The rats in the treatment group were given 15 mg/kg dose of 5,7-dimethoxycoumarin daily for 30 days intraperitoneally. However, those in the normal control and model groups received the same volume of normal saline. The rats were sacrificed after anaesthetization to extract brain samples.

Results: Quantitative polymerase chain reaction (PCR) analysis showed a significant (p < 0.02) decrease in the level of caspase-3 mRNA by 5,7-dimethoxycoumarin compared to the model group. Western blot assay data revealed significant decrease in caspase-3 protein expression by 5,7-dimethoxycoumarin. The higher activity of caspase-3 in acute cerebral infarction rats was reduced significantly (p < 0.005) following treatment with 15 mg/kg dose of 5,7-dimethoxycoumarin for 30 days. Treatment of the acute cerebral infarction rats with 15 mg/kg dose of 5,7-dimethoxycoumarin daily for 30 days led to a greater decrease in apoptotic index than those of rats with model group (p < 0.005). 5,7-Dimethoxycoumarin treatment daily for 30 days at a dose of 15 mg/kg also caused a significant (p < 0.005) decrease in neurological impairment score compared to the model animal group.

Conclusion: 5,7-Dimethoxycoumarin successfully inhibits neuronal apoptosis via inhibition of the expression of caspase-3 in brain tissues and thus may be useful for cerebral infarction therapy.

Keywords: Cerebral infarction, 5,7-Dimethoxycoumarin, Neurological impairment, Apoptosis, Caspase-3

INTRODUCTION

Acute cerebral infarction is a cerebrovascular disease and has very high incidence and mortality rate throughout the world [1,2]. The disease is characterized by blockage of blood to brain tissues leading to ischemia and hypoxia followed by necrosis of cerebral tissues [3]. The advanced stage of the disease involves generation of reactive oxygen radicals inducing neuronal cell death and subsequent activity impairment [4,5]. It is reported that during cerebral infarction in animals the rate of neuronal apoptosis is very high [6]. One of the important factors expressed in higher level and associated with the induction of neuronal apoptosis is the caspases [7-9]. In the brain tissue samples of cerebral infarction patients the expression of caspase-3 has been found to be markedly higher [10]. Thus inhibition of caspase-3 expression in
patients with cerebral infarction can play a vital role in the treatment of the disease. Therefore, various studies have been performed to investigate the effect of anti-oxidants on the pathology of cerebral infarction [11].

Coumarins, isolated in abundance from plant source exhibit promising anti-oxidant property [12, 13]. Both natural as well as synthetic coumarins are in clinical trials for the treatment of various types of cancers because of their tendency to induce carcinoma cell apoptosis [14]. 5,7-Dimethoxycoumarin (Figure 1), a derivative of coumarin present in vegetables has shown potent anti-cancer activity through induction of DNA damage and inhibition of carcinoma cell proliferation [15,16]. The chemical structure of 5,7-dimethoxycoumarinindicates that it has potential to act as anti-oxidant agent. 5,7-dimethoxycoumarin also causes cell cycle arrest in melanoma cell lines resulting in inhibition of cell proliferation [17]. The current study was designed and performed to investigate the effect of 5,7-dimethoxycoumarin on inhibition of apoptosis induction in the rat model of acute cerebral infarction. The study demonstrated that 5,7-dimethoxycoumarin successfully inhibited neuronal apoptosis through targeting the expression of caspase-3 in brain tissues.

**Figure 1:** Chemical structure of 5,7-dimethoxycoumarin

**EXPERIMENTAL**

**Animals and ethical approval**

38-weeks old male Sprague-Dawley rats (~210 g) used in the study were purchased using Animal licence: SCXK (no. Hebei 2014/213/41/56) from the Experimental Animal Center of Hebei Medical University. Fifteen days before the start of actual experiment, the animals were acclimatized to the laboratory environment. All the rats were housed under 12 h light/dark cycle in groups of 2/cage. The room was maintained at a humidity of 50 – 60 % and temperature of 23 ± 1 °C. All the animals had free access to food and water. Approval for the study was taken from the Ethics Committee of the Xintai Municipal People’s Hospital (approval no. CK43140407). All experimental animal procedures were conducted in compliance with State and Federal laws, standards of the US Department of Health and Human Services, and guidelines established by Tulane University Animal Care and Use Committee, accredited by Association for the Assessment and Accreditation of Laboratory Animal Care [18]. Efforts were made to reduce animal suffering to the minimum possible.

**Preparation of animal model of acute cerebral infarction**

For the preparation of acute cerebral infarction, a rat model cerebral artery infarction method reported earlier was used [19]. The establishment of animal model was confirmed by carefully examining the rats after surgery. The rats were then divided into three groups of 10 each; normal control, model and the treatment groups. The rats in the treatment group were injected 15 mg/kg doses of 5,7-dimethoxycoumarin daily for 30 days whereas those in the model and normal control group received the same volume of normal saline.

**RT-PCR analysis**

Total RNA from the brain tissue samples was isolated using TRIzol (Invitrogen, Carlsbad, CA, USA) as per the instructions mentioned in the manual protocol. The extracted RNA (1 μg) samples were subjected to reverse transcription using M-MLV reverse transcriptase and the Random-pdN6 Primer (Invitrogen). Amplification of the signals was performed using polymerase chain reaction (PCR) as follows: in the 50 μL water was taken template cDNA (2 μL), dNTPs (0.2 mM), 1.25 U/μL Go Taq polymerase (Promega, Madison, WI, USA) along with the various primers (0.5 μM each). It was then denatured by heating at 95 °C for 30 s and subsequently annealed for 30 sec at melting temperature. Elongation over 30 cycles was performed at 72 °C for 1-min. The sequence of used caspase-3 primer was: forward, 5'-GGT ATT GAG ACA GAC AGT GG-3' and reverse, 5'-CAT GGG ATC TGT TTC TTT GC-3'. Electrophoresis of the PCR products was performed on 2.0 % agarose gel and photographed under UV.

**Western blot analysis**

The brain tumor samples from the rats were collected, washed with cold PBS and subsequently treated with lysis buffer. After treatment with ice-cold lysis buffer for 45 min, the lysates were subjected to centrifugation for 15 min at 4 °C at 12,000 x g. The supernatant was
then collected and the concentration of proteins in the samples was analysed using Bicinchoninic Acid Protein Assay kit (Pierce, Rockford, IL, USA). The 50 µg protein samples were subjected to electrophoresis over 12 % sodium dodecyl sulphate-polyacrylamide gel. The protein samples were transferred to polyvinylidene difluoride (PVDF) membranes using a semidry transfer system (Bio-Rad Laboratories, Hercules, CA, USA). The non-specific sites in the membranes were incubated with 5 % bovine serum albumin and Tris-buffered saline with Tween® 20 buffer for 3 h. The bands were incubated with a specific rabbit anti-human antibody against caspase-3 mRNA (Cell Signaling Technology, Beverly, MA, USA). The bands were normalized to β-actin concentration. The complex development was performed using an ECL detection kit as per the manufacturer’s instructions (ECL GST western blotting detection kit, Pierce Biotechnology, Inc., Waltham, MA, USA). For the purpose of imaging, GelGDoc2000 imaging system (Bio-Rad Laboratories GmbH, Munich, Germany) was employed.

Examination of neurological impairment

Analysis of the impairment in neurological activity was performed using the known scale developed by Bederson [20]. The score of neurological impairment after surgery on the scale was assigned as follows: No neurological impairment symptoms = 0 score; problem in front paw extension = 1 score; rolling of body to left side = 2; tilting to one side only = 3 and complete movement loss = 4.

Analysis of caspase-3 activity

The activity of caspase-3 was analysed using commercially available reagent for detection of caspase-3 protein action (Beyotime Institute of Biotechnology, Beijing, China).

Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay

Brain tissue sample apoptosis was analysed using the In Situ Cell Death Detection kit, POD (Roche). The brain tissue samples were embedded in paraffin after washing with PBS and cut into thin 2 µm sections. The sections were then de-paraffined in boiling xylene and treated with hydrogen peroxide. The sections were fixed with 4 % paraformaldehyde and then subjected to counter staining for 10 min with 4',6-diamidino-2-phenylindole under dark atmosphere. The sections were observed carefully for apoptotic cells (TUNEL-positive cells) under a fluorescence microscope (Olympus IX70; Olympus Corporation, Tokyo, Japan).

Statistical analysis

SPSS software, version 17.0 (SPSS, Inc., Chicago, IL, USA) was employed for the analysis of the data. The data are presented as mean ± standard deviation (n = 3). The variance analysis was used for the comparison of the data among the groups. The differences were taken as statistically significant at p < 0.05.

RESULTS

Effect of 5,7-dimethoxycoumarin on caspase-3 mRNA levels

Quantitative PCR analysis showed a significantly (p < 0.05) higher level of caspase-3 mRNA in the rats with acute cerebral infarction compared to the normal control group (Figure 2). Treatment of the rats with 5,7-dimethoxycoumarin caused a significant (p < 0.02) reduction in the level of caspase-3 mRNA than those in the model group (Figure 2). Analysis of the caspase-3 protein level in rats of the model group revealed significantly higher level than those of the normal control rats (Figure 2). 5,7-Dimethoxycoumarin treatment of the rats with acute cerebral infarction caused a significant decrease in the level of caspase-3 protein compared to the model animal group (Figure 2). The level of both mRNA and protein corresponding to caspase-3 was thus reduced in the rats with acute cerebral infarction on treatment with 15 mg/kg dose of 5,7-dimethoxycoumarin.

Effect of 5,7-dimethoxycoumarin on caspase-3 activity

Analysis of the activity of caspase-3 in the rats with acute cerebral infarction revealed significantly (p < 0.05) higher level than those of rats in the normal control group. The higher activity of caspase-3 in acute cerebral infarction rats was reduced significantly on treatment with 15 mg/kg dose of 5,7-dimethoxycoumarin for 30 days. Caspase-3 activity in the normal, control and treatment groups was 2, 15 and 3.6, respectively. On the day 30th of treatment, level of caspase-3 activity in the normal control and 5,7-dimethoxycoumarin treatment groups was found to be similar.

Effect of 5,7-dimethoxycoumarin on apoptotic index and infarction scope

The apoptotic index in rats with acute cerebral infarction was markedly higher compared to
**Figure 2:** 5,7-dimethoxy coumarin prevents increase in expression level of mRNA and protein corresponding to caspase-3 in the rats with acute cerebral infarction. Rats with acute cerebral infarction were treated for 30 days with 15 mg/kg dose of 5,7-dimethoxy coumarin daily. The expression level of caspase-3 mRNA and proteins in the rat brain samples was analysed using qPCR and western blot assay, respectively

normal control (Figure 3). However, treatment of the acute cerebral infarction rats with 15 mg/kg dose of 5,7-dimethoxy coumarin daily for 30 days led to a significant decrease in apoptotic index than those of rats in the model group (Figure 3).

**Effect of 5,7-dimethoxy coumarin on neurological impairment in rats with acute cerebral infarction**

In the rats with acute cerebral infarction, neurological impairment was found to be markedly higher compared to the normal control group (Figure 4). 5,7-dimethoxy coumarin treatment daily for 30 days at a dose of 15 mg/kg caused a significant decrease in the neurological impairment score compared to the model animal group.

**DISCUSSION**

Coumarins, natural phytochemical products obtained from plants possess promising anti-oxidant activity [12,13]. Many coumarin molecules are in clinical trials against various types of diseases including carcinoma [14]. Apoptosis, programmed death of cells is induced through the generation of reactive oxygen species in cells which damage DNA and affect cellular pathways. In brain tissue samples of rats and patients the rate of neuronal apoptosis has been found to be markedly high [21]. Therefore, in the current study effect of the anti-oxidant, 5,7-dimethoxy coumarin on prevention of apoptosis induction in neuronal cells of acute cerebral infarction rats was investigated. The main factor responsible for induction of cell apoptosis is a protease, caspase-3 which triggers cellular pathways involved in neuronal apoptosis [22]. In the current study, the expression of both mRNA and protein corresponding to caspase-3 in the brain samples of rats with acute cerebral infarction was markedly higher. However, 5,7-dimethoxy coumarin prevented the increase in the level of mRNA and protein corresponding to caspase-3 in the rats.

The higher activity of caspase-3 in acute cerebral infarction rats was reduced significantly on treatment with 5,7-dimethoxy coumarin for 30 days. Caspase-3 is considered to be the significant target for prevention of acute cerebral infarction because of its major role in ischemia and induction of apoptosis [23,24]. In the current
Figure 3: 5,7-Dimethoxycoumarin prevents increase in apoptosis index in the rats with acute cerebral infarction. The brain samples of rats with acute cerebral infarction after treatment with 5,7-dimethoxycoumarin were analysed for apoptotic index using TUNEL staining. Quantification of neuronal cell apoptosis was performed using TUNEL staining.

Figure 4: 5,7-Dimethoxycoumarin prevents the increase in neurological impairment score in rats with acute cerebral infarction. The rats in the treatment group were injected with 15 mg/kg dose of 5,7-dimethoxycoumarin daily for 30 days and then neurological impairment score was examined.

study, the apoptotic index of rats with acute cerebral infarction was markedly higher than for normal control. However, treatment of the acute cerebral infarction rats with 5,7-dimethoxycoumarin daily for 30 days led to a significant decrease in the apoptotic index than those of rats in the model group. In the rats with acute cerebral infarction, neurological impairment was markedly higher compared to the normal control group. 5,7-Dimethoxycoumarin treatment daily for 30 days caused a significant decrease in the neurological impairment score compared to the model animal group.

CONCLUSION

The findings of the present study demonstrate that 5,7-dimethoxycoumarin successfully inhibits neuronal apoptosis by inhibition of expression of caspase-3 in brain tissues. Therefore, 5,7-dimethoxycoumarin can potentially be used for the treatment of acute cerebral infarction in patients.
DECLARATIONS

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Conflict of Interest

No conflict of interest associated with this work.

Contribution of Authors

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

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