Formulation and characterization of norethisterone transdermal patch as an alternative route to oral administration

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Abstract

Purpose: To design a transdermal norethisterone (NE) patch for controlled drug release

Methods: Polymers, viz, sodium salt of carboxymethyl cellulose (CMCNa) and hydroxyethyl cellulose (HEC), as well as a plasticizer (propylene glycole, PG) were used at various ratios to prepare NE films. The drug was dispersed in water and mixed with the polymer gel/plasticizer mixture prior to film casting. Weight variation, thickness, tensile strength, and the interaction between drug and film excipient were evaluated. The films were also characterized by Fourier transform infrared spectroscopy (FT–IR). In vitro drug dissolution and release from the transdermal patches (coded C-L, C-T, H-L and H-T) were evaluated.

Results: There was significant difference between the batches in terms of in vitro drug release (p > 0.05). Dissolved drug was significantly higher in H-T film when compared to C-L film (p < 0.05). In addition, drug release was significantly higher from H-T patch than from C-L patch (p < 0.05).

Conclusion: NE transdermal patches containing HEC and Transcutol® have been successfully formulated. They can potentially be used to deliver NE via transdermal route to decrease the side effects associated with the oral route of administration of the hormone.

Keywords: Norethisterone, Transdermal delivery, Controlled release, Hormone therapy

INTRODUCTION

Norethisterone (NE) is a potent synthetic progestogen which is used for the treatment of dysfunctional bleeding by shifting the time of menstruation [1,2]. NE is absorbed rapidly and completely following oral administration, and it reaches maximal plasma levels within 1 1/2 h. Oral bioavailability of NE is about 64 % due to the first pass metabolic effect. The half-life of NE is 8.0 ± 3.3 h. Being insoluble in water, NE is suitable for transdermal use because of its lipophilic nature [3-5].

Transdermal therapeutic systems (TTS) are developed for administration of drugs that have short plasma half-lives through the skin, so as to achieve controlled release. These systems minimize systemic side effects such as gastrointestinal (GI) side effects of drugs when...
taken through the oral route. In addition, drugs can be avoided from the first pass metabolism, thereby increasing their bioavailability [6,7]. Another advantage is that transdermal drugs can be easily removed by the patient when necessary. TTS, can be prepared using either natural or synthetic polymers in membrane or matrix systems [8,9].

To ease the penetration of drugs through the skin, penetration enhancers or fasteners may be added to the formulation [10]. TTS consists of non-permeable backing layer, drug reservoir, skin contact adhesive layer and a protective membrane [11].

This study was aimed at preparing transdermal patches of NE for the first time, in order to overcome the side effects associated with oral administration of the drug.

EXPERIMENTAL

Materials

Norethisterone was purchased from Bayer (Turkey). Nitrocellulose membranes were from Millipore, while 3M 9773 Foam Tape, CoTran 9719 Backing and Scotchpak 1020 Release Liner were products of Drug Delivery Systems, Germany. Bio-Psa AC7-4201 was procured from Dow Corning Corp, Turkey. Carboxymethyl cellulose sodium, hydroxymethylcellulose and limonene were products of Merck (Turkey). Propylene glycol was bought from Doğa, Turkey, while Transcutol® was product of Gattefosse (France).

Preparation of NE transdermal films and patches

NE films were prepared with CMCNa and HEC (Table 1). Aqueous polymer was prepared under ultrasonic bath (Bersonic Turkey) and kept at room temperature for 24 hours. Then PG was added to the polymer gel and mixed using HS-100D propeller mixer (Daian Scientific, Korea) at 500 rpm. NE was dispersed in water (1:5) and mixed with gel - plasticizer mixture, with blending by propeller mixer at 500 rpm. The mixture was homogenously poured into a 9 cm-diameter petri dish and dried inside a WiseVen Fuzzy Control System at 40 ± 2 °C (Daian Scientific, Korea). Limonene and Transcutol® were added to the liquid phase and films were prepared by the same method. NE transdermal patches (C-L, C-T, H-L, H-T) were prepared by using appropriate films. Films chosen were cut into 3.15 cm² surface area pieces Transdermal patches were made by 3M 9773 Foam Tape, CoTran 9719 Backing, film, silicone skin contact adhesive (0.01 ml) (Bio-Psa AC7-4201) and Scotchpak 1020 Release Liner, as transdermal patch components (Table 1).

Table 1: Composition (%) of NE transdermal films

<table>
<thead>
<tr>
<th>Composition</th>
<th>C</th>
<th>C-L</th>
<th>C-T</th>
<th>H</th>
<th>H-L</th>
<th>H-T</th>
</tr>
</thead>
<tbody>
<tr>
<td>CMC Na</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>HEC</td>
<td>-</td>
<td>-</td>
<td>3</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>PG</td>
<td>7.5</td>
<td>7.5</td>
<td>7.5</td>
<td>10</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>Transcutol®</td>
<td>-</td>
<td>-</td>
<td>5</td>
<td>-</td>
<td>-</td>
<td>5</td>
</tr>
<tr>
<td>Limonen</td>
<td>-</td>
<td>5</td>
<td>-</td>
<td>-</td>
<td>5</td>
<td>-</td>
</tr>
<tr>
<td>NE</td>
<td>0.75</td>
<td>0.75</td>
<td>0.75</td>
<td>0.75</td>
<td>0.75</td>
<td>0.75</td>
</tr>
</tbody>
</table>

Assessment of weight variation, thickness and tensile strength of films

Six different pieces (1 cm²) were carefully sliced from films and weighed. Their thickness were measured by a micrometer (Mitutoyo, Japan), and their tensile strengths were measured with Texture analyzer (TA – TX Texture Apparatus, U.K.). Determination was carried out under 200 g force with 5 kg loading chamber for 2s. Films of size 10 x 80 mm were cut, with film length fixed at 40 mm. Rotation speed of the probe were 1 mm/s and 10 mm/s at pre- and post-measurement phases respectively. The test stage speed was 0.5 mm/s.

Fourier transform infrared spectroscopy (FT – IR) analysis

FT - IR (Perkin Elmer 100 FT – IR instrument, U.K.) was performed to determine the interaction between drug and transdermal film components. Adequate amounts of pure NE and 0.5cm diameter film pieces were searched on a wave number between of 4000 to 650 cm⁻¹ at resolution of 4 cm⁻¹ in a FT-IR apparatus, at 100 N force.

Determination of NE content of films

The amounts of NE in the films were determined spectrophotometrically (Shimadzu UV - Visible Spectrophotometer UV - 1601, Japan) at 250 nm.

Circular pieces (1 cm²) sliced films were dissolved in 10 ml phosphate buffer solution (pH 7.4) containing 10 % ethanol (dissolution medium) in the ultrasonic bath for 30 min.
Samples were diluted to 25 ml and then filtered using blue ribbon filter S&S papers.

Validation of method of NE determination

The spectrophotometric method was validated for linearity, intra-day and inter-day precision, accuracy, recovery and specificity. Absorption spectra of NE showed λ max at 250 nm in dissolution medium. For this, 0.5 - 3 μg/mL standard NE solutions were prepared and assayed by suitable dilutions of 100 μg/mL stock NE solutions. Each study was replicated 6 times for linearity [12,13]. Precision of the method was shown as the relative standard deviation (RSD, %). Recovery (%) was calculated based on the amount of drug found. Stability study was done to confirm variation of amount of NE with time.

In vitro drug dissolution from films

In vitro dissolution test for transdermal patches was carried out in accordance with the procedure reported in European Pharmacopoeia in 2008. The test was performed on 4 cm pieces cut from films. Dissolution apparatus (Sotax AT – 7 Switzerland) was used with 900 mL of dissolution medium at 32 ± 0.5 °C and rotation speed of 50 rpm. Samples were taken at pre-determined time intervals, diluted and assayed spectrophotometrically to determine their NE content. Release profiles were kinetically assessed by using zero order, first order, and Higuchi and Korsmeyer - Peppas models [14-16].

Table 2: Physical properties and NE contents of films

<table>
<thead>
<tr>
<th>Formulation</th>
<th>Weight (mg)</th>
<th>Thickness (mm)</th>
<th>Tensile strength (kg/s)</th>
<th>NE (mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C</td>
<td>65.90±3.26</td>
<td>0.33±0.002</td>
<td>0.310±0.010</td>
<td>1.10±0.01</td>
</tr>
<tr>
<td>C-L</td>
<td>63.70±2.89</td>
<td>0.32±0.002</td>
<td>0.345±0.015</td>
<td>1.11±0.01</td>
</tr>
<tr>
<td>C-T</td>
<td>73.90±1.93</td>
<td>0.43±0.002</td>
<td>0.359±0.021</td>
<td>1.19±0.01</td>
</tr>
<tr>
<td>H</td>
<td>64.41±3.03</td>
<td>0.34±0.003</td>
<td>0.318±0.010</td>
<td>1.13±0.01</td>
</tr>
<tr>
<td>H-L</td>
<td>69.28±2.92</td>
<td>0.37±0.004</td>
<td>0.350±0.011</td>
<td>1.15±0.02</td>
</tr>
<tr>
<td>H-T</td>
<td>75.10±3.56</td>
<td>0.47±0.004</td>
<td>0.363±0.013</td>
<td>1.23±0.02</td>
</tr>
</tbody>
</table>

Statistical analysis

In vitro drug dissolution and release data were evaluated using GraphPad Prism 5 software to determine differences for profiles of all formulations at 0.05 as the level of significance.

RESULTS

Weight variation, thickness, tensile strength and drug composition of films

Results obtained from characterization studies, pharmaceutical quality of films and drug contents are shown in Table 2.

FT-IR spectra

FT-IR spectra of pure NE and its transdermal films are shown in Figures 1 and 2, respectively.

Characteristic bands of NE were detected at 3325.99 cm⁻¹ (O – H stretching band), 1646.53 cm⁻¹ (C = O carbonyl stretching band), 1614.60 cm⁻¹ (C = C double bound stretching band), 2975.84 – 2833.94 cm⁻¹ (C - H aliphatic stretching band), and 3271.08 cm⁻¹ (C = CH triple bond acetylene stretching band). All the peaks between 1500 - 400 cm⁻¹ are considered to be the finger print area of NE in Figure 1.
Figure 1: FT-IR spectra of pure NE

In the spectrum of C-T film, the main bands of NE were detected with lower intensities as seen in Figure 2 (b). Bands related to the structure of PG in the IR spectrum lost their sharpness. At 3351.18 cm⁻¹ (O-H stretching band), there was a broader PG peak. A broad peak (O-H stretching band) was formed by Transcutol®. The C-H aliphatic stretching band (2974.61 – 2877.59 cm⁻¹) peak was not so sharp due to the effect of PG. The C=C double bond stretching band was observed at 1647.93 cm⁻¹. Peaks belonging to NE skeletal structure were observable in the fingerprint area.

In the H-T film spectrum (Figure 2 (c)), the peaks of bands related to the structure of PG lost their sharpness. At 3345.23 cm⁻¹ (O-H stretching band), there was a broad PG peak along with another peak close to it. Thus, the C=CH triple bond acetylene stretching band was within the band. Here, the O-H stretching band of Transcutol® was distinguishable. Compared with the other bands, the O-H band was broader. Due to the influence of PG, 3007.59 - 2876.50 cm⁻¹ band (C-H aliphatic stretching band) was reduced in sharpness. The C=C stretching band (1648.43 cm⁻¹) was observable, but the C=O stretching band was not observable because of the broadening of the other peak. The C-O stretching band (1537.35 cm⁻¹), which comes from Transcutol® was visible. Peaks belonging to NE skeletal structure were seen in the fingerprint area.

Validation of spectrophotometric assay method

In this method, the regression equation was as in Eq 2.

\[ y = 0.0652x + 0.0016 \quad \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots (2) \]

where \( x \) is the concentration (μg/ml), and \( y \) is the absorbance. The assay coefficient was \( r^2 = 0.9992 \). The R.S.D. values gained from intra-day and inter-day precision of the analytical method were between 0.5 - 1.74. R.S.D. and values from accuracy analysis were between 1.37 and 1.71. The recovery of NE was between 98.45 - 99.69 %. Reagents such as the polymers and plasticizer did not absorb at 250 nm, the wavelength of maximum absorbance for NE. From the calculations, it was found that concentration range of 0.98 - 1.01 μg / ml, and R.S.D. values of 0.22 - 1.32 % were adequate for the stability of NE.

In vitro drug dissolution from films

Films were integrated in the medium during the test period. The profiles obtained from the dissolution study are shown in Figure 3.
amounts of drug dissolved at 8 hr for C-T, C-L and C were 42.79 %, 36.18 % and 35.12 %, respectively; and 60.04 %, 55.43 % and 54.18 %, respectively at 24 hr. There were no statistically significant difference between the dissolution rates of the formulations within the same time frame (p > 0.05). For the films, the highests amounts of drug dissolved at 8 th hour were 62.07 %, 52.07 % and 35.51 %; but at 24 hr, the amounts were 76.50 %, 67.29 % and
47.05 % for H - T, H - L and H, respectively. H - T had higher dissolution rate than H (p < 0.05), while H - T had higher dissolution rate than C and C - L (p < 0.05). Comparison of the drug release of the films showed that HEC (with 10 % PG), had a higher drug release after 24 hr than CMCNa with 7.5 % PGs. All films showed Higuchi kinetics pointing out matrix formation. When Korsmeyer Peppas kinetics was evaluated, all the patches showed non-Fickian kinetics, except C - L (Table 3). Therefore, H - L, H - T, C - L, and C - T were more suitable for incorporation into transdermal patches.

**In vitro drug release**

The result of transdermal patch drug release study from nitrocellulose membranes and the release profiles obtained are given in Figure 4. From this data, the amount of NE that diffused out from C - T was higher than that from C - L. For C - T and C - L patches, the calculated amount of NE released were 0.075 mg / cm² and 0.052 mg / cm² at the 8th hour; and 0.136 mg / cm² and 0.112 mg / cm² at the 24th hour, respectively. Among the CMCNa patches, there were no statistically significant difference between the drug released (p > 0.05). The amount of NE that diffused out from H - T was higher than the amount that diffused out from H - L.

For H - T and H - L patches, NE release were 0.084 mg / cm² and 0.069 mg / cm² at 8 hr; and 0.140 mg / cm² and 0.089 mg / cm² at the 24 hr, respectively. However, in HEC patches, there were no statistically significant differences between the released amounts of drug (p > 0.05). Released drug was highest in H - T followed by C - T when compared to C - L patch (p < 0.05).

All patches displayed Higuchi kinetic model. When Korsmeyer Peppas kinetics was evaluated, all the patches displayed non-Fickian model except H - L (Table 3).

**Table 3:** Kinetic modeling of drug release from films and patches

<table>
<thead>
<tr>
<th>Formulation</th>
<th>Zero Order r²</th>
<th>First order r²</th>
<th>Higuchi model r</th>
<th>Exponential (Korsmeyer–Peppas model) r²</th>
<th>Order of release (Korsmeyer–Peppas model) n</th>
</tr>
</thead>
<tbody>
<tr>
<td>Films</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C</td>
<td>0.9258</td>
<td>0.8105</td>
<td>0.9778</td>
<td>0.9593</td>
<td>0.9476</td>
</tr>
<tr>
<td>C - L</td>
<td>0.9155</td>
<td>0.6378</td>
<td>0.9811</td>
<td>0.8938</td>
<td>0.9231</td>
</tr>
<tr>
<td>C - T</td>
<td>0.8794</td>
<td>0.6662</td>
<td>0.9714</td>
<td>0.9599</td>
<td>0.7166</td>
</tr>
<tr>
<td>H</td>
<td>0.8842</td>
<td>0.7639</td>
<td>0.9747</td>
<td>0.9825</td>
<td>0.4052</td>
</tr>
<tr>
<td>H - L</td>
<td>0.8256</td>
<td>0.6724</td>
<td>0.9420</td>
<td>0.9603</td>
<td>0.5764</td>
</tr>
<tr>
<td>H - T</td>
<td>0.8220</td>
<td>0.6771</td>
<td>0.9388</td>
<td>0.9604</td>
<td>0.5464</td>
</tr>
<tr>
<td>Patches</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C - L</td>
<td>0.9370</td>
<td>0.8241</td>
<td>0.9885</td>
<td>0.9866</td>
<td>0.7539</td>
</tr>
<tr>
<td>C - T</td>
<td>0.9516</td>
<td>0.7313</td>
<td>0.9966</td>
<td>0.9855</td>
<td>0.8442</td>
</tr>
<tr>
<td>H - L</td>
<td>0.8897</td>
<td>0.7580</td>
<td>0.9713</td>
<td>0.9854</td>
<td>0.4492</td>
</tr>
<tr>
<td>H - T</td>
<td>0.9235</td>
<td>0.7605</td>
<td>0.9888</td>
<td>0.9876</td>
<td>0.5085</td>
</tr>
</tbody>
</table>

*Note: r² : determination coefficient; n : release exponent; n/a : not applicable*
The NE release rate from patches was highest with H - T (0.0058 mg /cm²/h), followed by C – T, C - L and H - L with release rates of 0.0056 mg /cm²/h, 0.0046 mg /cm²/h and 0.0037 mg /cm²/h, respectively. In addition, permeability coefficients of the patches was highest for H - T (4.55 x 10⁻³), followed by C – T, C - L and H - L with values of 4.02 x 10⁻³, 3.65 x 10⁻³ and 2.50 x 10⁻³, respectively.

DISCUSSION

The coefficient of the analytical method was $r^2 > 0.999$, which shows the linearity of the method. Precision was established by the R.S.D values in intra – day and inter - day NE analyses, with values lower than 2 % in line with literature requirements [12]. The recovery values of NE were very good and appropriate at between 98.45 - 99.69 %. Moreover, there were no interferences in absorbance at 250 nm by the polymers and other formulation components used. This is proof of the specificity of the method. NE was stable during the solubility and drug delivery studies, as shown by the R.S.D values which were lower than 2 %. The results obtained in tests on the properties and pharmaceutical qualities of the NE films had low standard deviation values which satisfy the requirements for quality. The higher drug contents of films containing Transcutol® indicate that NE release was slower when the drug was formulated with this agent.

The strength of transdermal films is crucial in transdermal formulations. The physical strength and resistance of the films were acceptable, due to the fact that the qualities of PG [18] resulted in production of more flexible films. From the FT - IR graphs, it was clear that the drug and film excipients were chemically compatible. Peaks belonging to NE skeletal structure were observed in fingerprint zones in all films [12]. Thus any changes in peaks must have occurred as a result of mixing of components and not due to any chemical interactions.

In this study, limonene and Transcutol® were used to ease drug release from the formulations. These compounds are generally recognized as safe (GRAS) by the FDA. Limonene and Transcutol® not only form direct structural change on the skin, but also ease drug release from formulations [19]. Results of in vitro dissolution studies show that HEC films were of higher profiles during the first 24 hours than CMCMNa films. H, which did not contain penetration enhancers, showed lower dissolution results. The higher dissolution profiles of films containing penetration enhancers than films without them can be explained with the characteristics of Transcutol® and limonen. Increases in the amount of PG in formulations resulted in higher drug release. This can be attributed to the synergistic action of PG with penetration enhancers. In addition, the solubility of NE was 45.326 ± 1.09 µg / ml in dissolution medium. This value indicates that the water-insoluble NE can be dissolved sufficiently in the dissolution medium. In vitro drug release was higher in NE patches containing Transcutol® when compared to patches with limonene, indicating that Transcutol® was more effective than limonene. Drug release from all films and patches displayed Higuchi kinetic model [15]. This strongly suggests that diffusion may be referred to drug release mechanism. The Fickian kinetic model indicates that a drug is released through diffusion mechanism, while the non-Fickian kinetic model indicates that drug release occurs through a hybrid mechanism of diffusion / erosion [16,17].

CONCLUSION

NE transdermal patches containing HEC and 5 % Transcutol® may be good candidates for enhanced skin penetration and delivery of NE. Thus, these patches can be further developed as an alternative to the oral route of administration of this drug to improve patient compliance.

DECLARATIONS

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Conflict of interest

No conflict of interest is associated with this work.

Contribution of Authors

The authors declare that this work was done by themselves and all liabilities pertaining to claims relating to content of this article will be borne by them.

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