Bone morphogenetic protein 2/7 promotes repair of bone defect via induction of endochondral ossification

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Abstract

Purpose: To study the influence of bone morphogenetic protein 2/7 (BMP2/7) on repair of bone defect, and the mechanism involved.

Methods: Bone marrow stromal cells (BMSCs) were randomly assigned to 2 groups: control and transfection groups. The cells were transfected with rBMP2/7 recombinant adenosivirus. Cell growth and alkaline phosphatase (ALP) activity were determined in both groups. Rabbit model of femoral bone defect was prepared using standard methods. Male New Zealand white rabbits were randomly assigned to 3 groups, each of which had 10 rabbits: control, gelatin and BMP2/7 groups. Histopathological and x-ray examinations, and three-point bending flexural test were used to compare the potential of gelatin and BMP2/7 to repair bone defects.

Results: Transfection of BMSCs with rBMP2/7 recombinant adenosivirus significantly enhanced their growth (p < 0.05). Alkaline phosphatase (ALP) level was also markedly and time-dependently higher in transfection group than in control group (p < 0.05). Rabbits with grade 4 bone healing or above were more in BMP2/7-treated category than in control and gelatin groups. New bone hyperplasia with typical lamellar bone structure, irregular medullary cavity, as well as transition from osteoblast to osteocyte were observed in BMP2/7 group. Moreover, maximum flexural strength and repair were significantly higher in BMP2/7-transfected group than in control.

Conclusion: These findings indicate that BMP2/7 promotes the repair of bone defect via induction of endochondral ossification in rabbits. Thus, this protein may be useful for the repair of bone defects in humans.

Keywords: Alkaline phosphatase, Bone lesions, BMP2/7, Endochondral ossification

INTRODUCTION

Bone defects are caused by trauma, bone disease or tumor, congenital abnormalities, infection (osteomyelitis) and failed arthroplasty. Bone defect often leads to bone disconnection, delayed union, fracture, as well as local dysfunction. The maintenance of bone length and union remains a major challenge to orthopedic surgeons. This has necessitated the search for novel and effective bone regeneration therapy. Techniques for reconstruction of bone defects include grafting, distraction osteogenesis, biomaterial and prosthetic implants.
Bone grafting, a commonly used method for repairing bone defects, comprises autogenous bone graft, allogeneic bone graft and artificial bone material filling [1]. Often regarded as the gold standard for bone graft, autogenous bone graft has advantage of being osteoconductive, osteo-inductive and osteogenic. However, it is limited by restricted donor sites and possible harvesting morbidity, unpredictable resorption, and small bone volume [2]. Autogenous graft is superior to allograft, since remodeling and bone healing take place more slowly in allografts, relative to autografts.

In recent years, tissue engineering has made it possible to carry out in vitro construction and in vivo recombination. Bone marrow mesenchymal stem cells (BMSCs) have been used as seed cells in the implantation of scaffold materials. However, the process of osteoblast induction is too complex, requiring the induction of appropriate growth factors [3,4].

A member of the transforming growth factor (TGF) β superfamily, BMG stimulates BMSCs to differentiate into osteoblasts and chondrocytes. It also induces ectopic bone formation during healing and repair of bone defects [5]. At present, BMP-2 and BMP-7 are most frequently used. The present investigation focused on the influence of BMP2/7 on repair of bone defect, and the mechanism involved.

EXPERIMENTAL

Materials

Sodium pentobarbital injection was bought from Shanghai New Asia Pharmaceutical Co. Ltd. Epinephrine hydrochloride was a product of Tianjin King York Amino Acid Co. Ltd. Penicillin G sodium was obtained from Shanghai Baoman Biotechnology Co. Ltd. Atropine sulfate was purchased from Tianjin Pharmaceutical Group Xinzheng Co. Ltd. Collagen sponge was bought from Hangzhou Lianke Biotechnology Co. Ltd. Pure titanium implant was a product of Zhejiang Guangci Medical Device Factory, while purified recombinant human BMP2/7 was obtained from R & D Corporation (USA).

Double-double display constant temperature water bath was bought from Shanghai Chenlian Biotechnology Development Co. Ltd. Vacuum freeze dryer was purchased from Shanghai Jingxin Industrial Development Co. Ltd. Precision electronic universal material testing machine (AGS-H) was bought from Shimadzu Corporation (Japan). Scanning electron microscope was a product of Shanghai Jingke Chemical Technology Company, while x-ray machine was purchased from Beijing Huarisen Technology Development Co. Ltd.

Cell preparation and culture

Bone marrow extracted from New Zealand white male rabbits was dispersed into individual cells by subjecting it to density gradient centrifugation at 1500 rpm for 15 min at 4 °C using lymphocyte separation solution (a mixture of Ficoll and sodium diatrizoate) as the density gradient solution. The bone marrow stromal cells (BMSCs) were subsequently re-suspended in M199 medium containing 10 ng/mL vascular endothelial growth factor (VEGF), 10 % FBS, 5 ng/mL fibroblast growth factor (FGF) and 5 ng/mL epidermal growth factor (EGF). The resultant cell suspension was seeded in culture plates pre-coated with human fibronectin, and cultured in a mixture of DMEM and Ham's F-12 medium (DMEM-F12) containing calcium sulfate at 37 °C for 72 h in a humidified atmosphere of 5 % CO2 and 95 % air. The medium was changed every 2 days. The cells were observed under an inverted microscope, and those in logarithmic growth phase were selected and randomly assigned to 2 groups: control and transfection groups. The cells were transfected with rBMP2/7 recombinant adenovirus.

Determination of ALP activity

After incubation for 9 days, ALP activity of cells on the composite scaffold was determined using enzyme-linked immunosorbent assay (ELISA).

Rabbits

New Zealand white male rabbits aged 16 to 20 weeks and weighing 1.8 - 3.3 kg (mean age = 18 ± 2 weeks; mean weight = 2.55 ± 0.75 kg) were obtained from Beijing Vital River Lab. Animal Tech. Co. Ltd. The rabbits were randomly assigned to 3 groups (10 rabbits/group): control, gelatin and BMP2/7 groups. The BMP2/7 (0.05 µg/L) was prepared in 4 mmoles/L HCl containing 0.1 % FBS. Collagen sponges were cut into 15 × 4 × 2.5 mm fragments and sterilized. Each gelatin sponge was made to absorb 100 µL of BMP2/7 solution via aspiration, and allowed to dry under aseptic conditions.

This research was approved by the Animal Ethical Committee of Fuzhou Second Hospital Affiliated to Xiamen University (approval no. 2019826638), and was performed according to the guidelines of Principles of Laboratory Animal Care [6].
Establishment of rabbit model of femoral bone defect

Each rabbit was anesthetized via intravenous injection of 3 % pentobarbital sodium (25 mg/kg body weight) at the edge of the ear. After aseptic preparation of the surgical site (left side of the rabbit’s femur), the skin and subcutaneous tissue were cut open and an anterolateral longitudinal incision was made to expose the femur. Bone defects of 10 × 4 × 3 mm (groove-shaped) were then made in the middle of the femur. In the control group, the bone defect area was filled with collagen sponge containing BMP. Rabbits in gelatin group were implanted with gelatin sponge of equivalent size, while BMP2/7 group rabbits were implanted with collagen sponge containing BMP2/7. Penicillin (40 U) was administered to each rabbit twice daily for three days.

Assessment of fracture healing

Computed radiography (CR) image of each rabbit femur was taken on the day of operation and 12 weeks post-operation. The images were analyzed using Image-Pro Plus analysis software (5.0). Healing of bone defect was assessed via X-ray examination, and was graded using standard grading system: 0 = absence of new bone formation; 1 = formation of new bone ≤ 25 %; 2 = formation of new bone ≥ 25 %; 3 = formation of new bone ≥ 30 %; 4 = formation of new bone ≥ 75 %; and 5 = complete replacement of bone defect area by new bone.

Histological examination of bone tissue

Histological examination of the fractured area was performed using hematoxylin and eosin (H & E) staining 12 weeks after operation. Histopathological changes were assessed based on the degree of inflammatory cell infiltration, osteogenesis, damage, underlying bone destruction and articular cartilage damage. The implanted material was taken out together with near bone segment and fixed in 10 % formalin solution. It was subsequently treated with 10 % EDTA (pH = 7.2) for 72 h to allow for decalcification. ImageJ analysis software was employed to determine the formation of new bone around the affected area.

Three-point bending flexural test

Precision electronic universal material testing machine (AGS-H) was used to carry out three-point bending flexural test. The test was performed in triplicate at 10 min intervals, and the maximum bending strength of new bone was compared amongst the groups. The extent of bone repair (B) was calculated as shown in Eq 1.

\[ B = \frac{St}{Sr} \times 100 \]  \hspace{1cm} (1)

where St and Sr are the maximum bending strengths of new bone and reference control, respectively.

Statistics

Measurement data are presented as mean ± SD. Student’s t-test was used for comparing two groups was performed with. Counting data are expressed as n (%), and χ² test was applied for comparing data amongst groups. All statistical analyses were done with SPSS (21.0). Values of \( p < 0.05 \) were taken as indicative of statistically significant differences.

RESULTS

Cell growth on scaffold material

Transfection of BMSCs with rBMP2/7 recombinant adenovirus significantly enhanced their growth (\( p < 0.05 \); Figure 1).

Activity of ALP on scaffold material

As shown in Table 1, ALP activity was significantly and time-dependently higher in transfected cells than in control.

<table>
<thead>
<tr>
<th>Group</th>
<th>Day 3</th>
<th>Day 7</th>
<th>( t )</th>
<th>( P )-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Transfection (U)</td>
<td>0.67 ± 0.02</td>
<td>1.09 ± 0.04</td>
<td>29.699</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Control (U)</td>
<td>0.33 ± 0.05</td>
<td>0.65 ± 0.03</td>
<td>17.354</td>
<td>&lt; 0.001</td>
</tr>
</tbody>
</table>

\( T = 19.966 \), \( P \)-value = 0.001

Data are presented as mean ± SD
Table 2: Grades of healing of bone defect in each group of rabbits

<table>
<thead>
<tr>
<th>Group</th>
<th>0 (Grade 0)</th>
<th>1 (Grade 1)</th>
<th>2 (Grade 2)</th>
<th>3 (Grade 3)</th>
<th>4 (Grade 4)</th>
<th>5 (Grade 5)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>6 (60.00%)</td>
<td>4 (40.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
</tr>
<tr>
<td>Gelatin</td>
<td>10 (100.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
</tr>
<tr>
<td>BMP2/7</td>
<td>0 (0.00%)</td>
<td>0 (0.00%)</td>
<td>1 (10.00%)</td>
<td>2 (20.00%)</td>
<td>5 (50.00%)</td>
<td>2 (20.00%)</td>
</tr>
</tbody>
</table>

Results are presented as n (%)

Healing of bone defect

At 12 weeks post-operation, CR images of femurs of rabbits in the control group showed that bone callus was formed in a small amount, while the absorption of the broken end appeared thinner. Bone defect area of rabbits in gelatin group revealed non-union image with medullary cavity atresia. However, CR images of femurs of rabbits in BMP2/7 group revealed transformation of callus into woven bone with clear cortical structure (linkage of new bone with broken bone). Moreover, the defect area was full of fibrous connective tissue, inactive bone hyperplasia, and closed medullary cavity. In addition, rabbits with grade 4 bone healing or above were more in BMP2/7-treated rabbits than in gelatin and control rabbits (Table 2).

Effect of BMP2/7 on bone histology

In the control group, new bone hyperplasia, bone trabecula and lamellar bone were observed at the junction of the implant area with host bone. Most of the materials in the center of the implant area were absorbed, and a large amount of fibrous tissue and chondrocytes were also visible. The bone defect area of rabbits in gelatin group was filled with fibrous connective tissue containing inactive bone end hyperplasia, and closed medullary cavity. New bone hyperplasia with typical lamellar bone structure, irregular medullary cavity, as well as transition from osteoblasts to osteocytes were observed in BMP2/7 group. These results are shown in Figure 2.

Three-point bending test data

As shown in Table 3, maximum flexural strength and percentage repair were markedly higher in BMP2/7 group than in control (p < 0.05).

<table>
<thead>
<tr>
<th>Group</th>
<th>Maximum flexural strength (N)</th>
<th>Bone repair (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>38.27 ± 8.10</td>
<td>27.02</td>
</tr>
<tr>
<td>Normal bone control of the same group</td>
<td>141.64 ± 15.23</td>
<td></td>
</tr>
<tr>
<td>BMP2/7</td>
<td>121.03 ± 12.72*</td>
<td>86.03</td>
</tr>
<tr>
<td>Normal bone control of the same group</td>
<td>140.68 ± 12.07</td>
<td></td>
</tr>
</tbody>
</table>

*P < 0.05, compared with control group

DISCUSSION

The bone plays a vital function in maintaining structural integrity, mineral storage, erythropoiesis, blood clotting and immune response. It is a highly vascularized tissue with inherent healing capacity. Bone defect refers to the absence of bone where it should normally occur. Surgical reconstruction of bone defects can present a significant challenge to orthopedic surgeons. In the treatment of bone defect, autologous/allogeneic bone and other synthetic materials are used to fill the bone gaps and promote fracture healing. However, these strategies generally have limitations such as excessive blood loss in the donor site, small bone volume, as well as complications and aggravation of pain in patients. Advances in bone tissue engineering have made it possible for artificial bone substitutes to be used in clinical practice [7,8].

Seed cells, signal factors and scaffold materials are essential elements needed for tissue engineering.

Bone tissue engineering promotes production of new and viable bone via synergistic interaction amongst biomaterials, cells, and factor therapy. This is viewed as a potential replacement for traditional usage of bone grafts due to its ready availability and zero risk of contracting diseases. At present, autografts are the gold standard for bone grafts due to their being histocompatible and non-immunogenic, while having all characteristics of bone graft substance. Scaffold-based gene therapy is a tissue engineering technique used for transfection of cells to facilitate production of target proteins, or
for silencing of genes linked to lesions of bone and joint.

Bone morphogenetic protein (BMP) has become the most studied target gene and the most frequently applied in tissue engineering due to its high osteogenic capacity [9,10]. It is known that BMP is the only local growth factor that can independently induce bone tissue formation, thereby promoting bone and cartilage repair. Bone morphogenetic proteins (BMPs) 2, 4, 5, 6, 7 and 9 possess high biological properties and strong osteogenic effects. Bone morphogenetic protein (BMP)-2 and 7 are used in spinal fusion and repair of long bone defects [11, 12]. Application of BMP in bone microenvironment at doses higher than physiological dose enhances osteogenesis and osteoclastogenesis [13]. High concentration of BMP2 in cancellous bone promotes osteoclast activity around the implant [14]. Studies have shown that BMP heterodimers possess strong bone-inducing activity. Low-dose concentration of BMP2/7 heterodimer have been demonstrated to effectively improve the function of osteoclasts, and to induce and reshape new bone at an early stage [15]. The high osteo-inductive activity of BMP2/7 heterodimer is thought to be due to its increased affinity for BMP receptors [16].

The present study was focused on the influence of BMP2/7 on repair of bone defect, and the mechanism involved. The results showed that BMSCs transfected with rBMP2/7 effectively adhered to scaffold material and grew uniformly. The activity of ALP activity was significantly and time-dependently higher in transfection group than in control group, an indication that transfection of BMSCs with rBMP2/7 may promote osteogenesis in vivo. The results of histological examination showed that at 12 weeks post-operation, femurs of rabbits in BMP2/7 group had new bone hyperplasia with typical lamellar bone structure. Thus, it is likely that BMP2/7 participates in intramembranous osteogenesis (direct conversion of mesenchymal tissue into bone). The results of three-point bending test suggest that the strength of femur after repair of defect reached that of normal bone.

CONCLUSION

The results of this study indicate that BMP2/7 promotes the repair of bone defect via induction of endochondral ossification in rabbits. Thus, the protein has a potential for use in the repair of bone defects.

DECLARATIONS

Acknowledgement

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Conflict of interest

No conflict of interest is associated with this work.

Authors’ contributions

This study was done by the authors named in this article, and the authors accept all liabilities resulting from claims which relate to this article and its contents. The study was conceived and designed by Jianjun Wu. Jianjun Wu, Changkun Zheng, Guiqing Liang collected and analyzed the data. Jianjun Wu wrote the text. All authors read and approved the manuscript for publication.

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