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Original Research Article

Isolation of cytotoxic triterpenes from the mangrove plant, *Scyphiphora hydrophyllacea* C.F.Gaertn (Rubiaceae)

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Abstract

Purpose: To isolate active cytotoxic compounds from the hexane and chloroform extracts of the leaves of the mangrove plant, Scyphiphora hydrophyllacea C.F. Gaertn (Rubiaceae), grown in Sri Lanka. **Methods:** Dried pulverized leaves of S. hydrophyllacea were extracted with hexane and chloroform. Vacuum liquid chromatography (VLC), column chromatography (size exclusion chromatography, Sephadex LH-20) and reversed phase preparative recycling high performance liquid chromatography (HPLC) techniques were used to isolate three compounds (compounds **1**, **2** and **3**). The structures of the isolated compounds were established with the aid of ¹H, ¹³C and two-dimensional nuclear magnetic resonance (2D-NMR) and electron ionization-mass spectrometry (EI-MS) techniques. 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay was used to evaluate the cytotoxic effects of the compounds on oestrogen receptor positive breast (MCF-7) and non-small cell lung (NCI-H-292) cancer cells.

Results: The isolated compounds were identified as oleanolic acid (1), ursolic acid (2) and eichlerianic acid (3). Ursolic acid and eichlerianic acid showed strong cytotoxic effects { IC_{50} - ursolic acid: 8.47 µg/mL (24 h, MCF-7), 7.78 µg/mL (24 h, NCI-H292) and eichlerianic acid: 8.86 µg/mL (24 h, MCF-7), 10.15 µg/mL (24 h, NCI-H292)} in MCF-7 and NCI-H292 cancer cells at 24, 48 and 72 h post-incubation periods.

Conclusion: Hexane and chloroform extracts of the leaves of S. hydrophyllacea yielded three compounds namely oleanolic acid, eichlerianic acid and ursolic acid. Ursolic acid and eichlerianic acid have been isolated for the first time from the leaves of S. hydrophyllacea grown in Sri Lanka and demonstrate in-vitro cytotoxic effects in oestrogen receptor positive (MCF-7) and non-small lung cancer (NCI-H-292) cells.

Keywords: Scyphiphora hydrophyllacea, eichlerianic acid, ursolic acid, oleanolic acid, MCF-7, NCI-H-292

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INTRODUCTION

Cancer is considered a major cause of death in the world [1]. Lung and breast cancers are the most commonly diagnosed cancer among men and women respectively. Despite the availability of modern cancer treatment options, there is no permanent cure for cancer [2,3]. Radiotherapy, chemotherapy, hormone therapy and surgery are the main available treatment options for cancer [4.5]. Chemotherapy and radiotherapy will be the ultimate treatment option for metastatic cancer when surgery causes some functional defects in the organs and systems of the patient. However, chemotherapy and radiotherapy cause severe side effects [6]. Therefore identification of alternative treatment options for cancer with fewer side effects is of paramount importance. Phytochemicals have gained much attention in the development of cancer drugs and several plant derived natural compounds have been developed as anti-cancer drugs [7].

Mangroves are considered as one of the main productive coastal eco-systems. Mangroves have widely been used in traditional medicine to treat cancers and a number of natural compounds with different bio-activities have been reported from several mangrove plants [8]. S. hydrophyllacea (Family: Rubiaceae) is one of the common manarove plants found mainly in South Asia including Sri Lanka [8, 9]. Several flavonoids, terpenoids and iridoids have been isolated from S. hydrophyllacea by previous Cytotoxic and researchers [9]. apoptotic properties of leaf and bark extracts of S. hydrophyllacea and isolation of hopenone-I from the hexane leaf extract of S. hydrophyllacea have reported in our previous studies [10,11]. The present study was aimed at to isolate cytotoxic compound/s in the active hexane and chloroform extracts of S. hydrophyllacea leaves and to evaluate their cytotoxic effects in nonsmall cell lung (NCI-H292) and breast cancer (MCF-7) cells.

EXPERIMENTAL

General

Cell lines and cell culture reagentsused in this studuy were purchased from the ATCC (American Type Culture Collection, Manassas, USA) and all the chemicals in the study were purchased from the Sigma-Aldrich (St Louis, MO, USA) unless otherwise stated. Bruker AV-500 and AV-600 NMR instruments (Fällanden, Switzerland) were used to record NMR spectra. Synergy™ HT Multi-Mode micro plate reader (Bio- Tek, Winooski, VT) was used to measure absorbance. All the solvents used in this study were of analytical grade.

Collection of plant material

Leaves of *S. hydrophyllacea* were collected from the same plant used for our previous studies [10, 11]. Leaves of the plant were collected from the National Aquatic Resources Research and Development Agency (NARA) Negombo, Sri Lanka and the plant material was identified by Mr. W.A. Sumanadasa, Research Officer at NARA. Voucher specimen (no. S-11) of the plant was deposited at the herbarium of IBMBB, University of Colombo, Sri Lanka.

Extraction and isolation of compounds

Air-dried pulverized leaves of S. hydrophyllacea (1.5 kg) were extracted with hexane (3×1.75 L) and chloroform (3 × 1.75 L) by sonication at room temperature. Resulting chloroform extract was concentrated (32 g) and subjected to vacuum liquid chromatography (VLC) with a gradient solvent system composed of hexane: ethyl acetate (100:0 to 0:100) to yield 10 fractions. Fraction 9 (7.6 g), which was found to be cytotoxic, was seperated in a Sephadex LH-20 column (120 × 3.5 cm) with 50 % (v/v) dichloromethane and methanol to yield 17 fractions. The collected fractions were combined according to TLC profiles to make 4 fractions which were then subjected for cytotoxic evaluations. Most active fraction (fraction 3, 30 mg) was then subjected to preparative recycling HPLC in a normal-phase column with isocratic elution [5% (v/v) MeOH in CHCl₃; flow rate: 5 mL/min]. Peaks were detected using refractive index detector (RI detector) at a sensitivity of 20nm/RIU. Two recycled base line separated peaks were collected to afford compounds 1 (7 mg) and 2 (12 mg).

Hexane crude extract (12 g) was subjected to vacuum liquid chromatography (VLC) with hexane : ethyl acetate (100:0 to 0:100) to yield 10 fractions. Fraction 6, which showed the most cytotoxic activity, was chromatographed on a normal phase silica column with hexane: ethyl acetate (100:0-0:100) to yield 20 fractions. As the fraction 14 (200 mg) exerted most cytotoxic effects it was separated in a Sephadex LH-20 column (120 × 3.5 cm) with 50 % (v/v) dichloromethane and methanol to yield compound **3** (120 mg).

Structure elucidation

Structures of the isolated compounds were established through of UV and IR spectroscopy,



Figure 1: Structures of isolated compounds 1-3

¹H-NMR, ¹³C-NMR, 2D-NMR (COSY, HSQC, and HMBC) and ESI-MS techniques.

Cell culture

Oestrogen receptor positive breast cancer (MCF-7) and non-small cell lung (NCI-H292) cancer cells were plated in 96-well cell culture plates in Dulbecco's Modified Eagle's Medium (DMEM) at 37 °C in 95 % air and 5 % CO_2 atmosphere with 95 % humidity.

Cytotoxic assay

MTT assay was carried out as previously described methods [12,13]. Cancer cells plated in 96 well plates were treated with different concentrations (6.25, 12.5, 25, 50 and 100 μ g/mL) of compounds and incubated for 24, 48 and 72 h. After incubation, MTT reagent (20 μ L of 1 mg/mLstock solution) was added to each well and plates were incubated for 4 h at 37 °C. Following incubation, content was removed from each well and 100 μ L of isopropanol/HCl mixture was added. Absorbance of treated and untreated wells were measured at 570 nm in a micro plate reader. Paclitaxel was used as the positive control.

Morphological examination

Morphological changes of breast (MCF-7) and lung cancer (NCI-H292) cells exposed to three compounds were examined under phase contrast microscope (OPTIKA, XDS 3, Italy).

Statistical analysis

Data are shown as mean \pm SD (n = 3). Statistical differences as well as half-maximal inhibitory concentration (IC₅₀) values of the three compounds in two different cancer cells were analyzed using GraphPad Prism 5 (San Diego, California, USA) software package. *P* < 0.05 was taken as statistically significant.

RESULTS

Structure of isolated compounds

Structure of compound 1 and 2 were confirmed

as oleanolic acid and ursolic acid by comparing NMR and mass spectroscopic data of these two compounds with reported data (Table 1 and 2) in literature [14,15]. Compound **3** was obtained as a white powder. EI-MS spectrum of compound **3** displayed pseudo molecular ion peaks at *m*/z 475 [M⁺ + H] and 492 [M⁺ + NH₄]. EI-MS spectrum displayed peaks at *m*/z 459 (M⁺- Me), 441 ((M⁺- Me-H₂O), 415 (M⁺- Me-COOH), 397, 143 and 125.¹H- and ¹³C-NMR chemical shift values of compound **3** found to be similar with reported compound eichlerianic acid [14].

Comparative NMR chemical shift values of compound **3** are given in Table 3. Structure of compound **3** was further confirmed from 2D-NMR spectroscopic techniques (HSQC, COSY, HMBC and NOESY). Structures of the isolated compounds are shown in Figure 1.

Cytotoxic effects of isolated compounds

Cytotoxic effects of isolated compounds are shown in the Table 4. All three compounds were cytotoxic to cell lines tested. Compared to compound 1 (oleanolic acid), compound 2 (ursolic acid) and compound 3 (eichlerianic acid) were highly cytotoxic to both cancer cell lines tested. However compound 3 appeared to exert a higher effect in breast cancer cells than in lung cancer cells.

Table 4: IC_{50} values (in µg/mL) of oleanolic acid (compound 1), ursolic acid (compound 2) and eichlerianic acid (compound 3) in MCF-7 and NCI-H292 cells at 24, 48 and 72 h incubation periods

Compound	MCF-7			Ν	CI -H29	2
	24 h	48 h	72 h	24 h	48 h	72 h
1	208.4	141.6	118.9	102.5	80.96	62.84
2	8.47	7.06	4.36	7.78	6.26	5.72
3	8.86	7.22	5.00	10.15	9.77	8.35
Paclitaxel	4.25	3.18	1.94	10.2	3.25	1.25

Morphological observations

Changes in cell structure such as membrane blebbing and apoptotic body formation changers in cell shape and decrease in cell number were evident in lung and breast cancer cells after

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Position	Report	ported data [15] Observed data		served data
	δC	δ H (J Hz)	δС	δ Η (J Hz)
1	38.5	1.63 (m)	38.5	1.63 (m)
2	28.1	1.60 (m)	28.1	1.59 (m)
3	79.1	3.23 (dd; <i>J</i> = 10.7; 4.7 Hz	79.1	3.20 (dd; <i>J</i> = 4.5; 10.0 Hz)
4	38.8	-	38.8	,
5	55.3	0.74 (m	55.2	0.71 (m)
6	18.8	1.54 (m)	18.2	1.54 (m)
7	32.7	1.49 (m)	32.9	1.48 (m)
8	39.3	-	39.4	
9	47.6	1.54 (m	47.5	1.54 (m)
10	37.0	-	37.0	
11	23.8	0.94 (m)	23.5	0.94 (m)
12	122.8	5.31 (dd; <i>J</i> = 3.6; 3.5 Hz)	125.8	5.23 (d; <i>J</i> = 22.5 Hz)
13	143.5	-	137.9	
14	41.5	-	41.9	
15	27.7	1.60 (m)	27.9	1.61 (m)
16	23.7	0.94 (m)	24.1	0.96 (m)
17	46.7	-	47.8	
18	42.1	2.82 (m)	41.9	
19	46.0	2.87 (m)	47.5	
20	31.0	-	30.6	
21	33.9	1.62 (m)	38.75	1.63 (m)
22	33.2	1.30 (m)	36.98	1.31 (m)
23	28.0	1.00 (s)	28.12	0.96 (s)
24	16.8	0.79 (s)	16.9	0.75 (s)
25	15.3	0.93 (s)	15.4	0.91 (s)
26	17.1	0.79 (s)	17.0	0.76 (s)
27	26.0	1.16 (s)	27.2	1.06 (s)
28	180.0	-	181.2	
29	33.1	0.92 (s)	36.68	0.84 (s)
30	23.7	0.94 (s)	23.2	0.93 (s)

Table 1: ¹³ C NMR (150MHz) and Image: control of the second	¹ H NMR data (600 MHz) of oleanolic acid (1) (CDCl ₃)
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treatment with the three compounds at 24, 48 and 72 h incubations (Figure 2 and Figure 3).

DISCUSSION

Scyphiphora hydrophyllacea is a mangrove plant of the family Rubiaceae which is widely distributed in Asia and Western Pacific [9]. Presence of flavonoids, terpinoids and iridoids in the mangrove plant S. Hydrophyllacea has already been reported [9]. Structures of three compounds isolated from the hexane and chloroform extracts of leaves of S. Hydrophyllacea were elucidated by means of 1 H- and ¹³C-NMR data and the compounds were identified as oleanolic acid (compound 1), ursolic acid (compound 2) and eichlerianic acid (compound 3). Although isolation and structure elucidation of ursolic acid and oleanolic acid from several other mangrove varieties has been reported [16,17], this is the first report on the

isolation eichlerianic acid from *S. Hydrophyllacea* leaves.

Cytotoxicity results from the cytotoxic assays demonstrated that, among the three compounds ursolic acid and eichlerianic acid exerted strong cytotoxic effects on non-small lung cancer cells (NCI-H292) and oestrogen receptor positive (MCF-7) breast cancer cells at 24, 48 and 72 h post incubation periods. Morphological changes (changes in cell shape, volume and decrease in cell number) observed in ursolic acid and eichlerianic acid-treated cancer cells also confirmed cancer cell growth inhibitory effects of these two compounds. Oleanolic acid showed less cytotoxic effects in both the cancer cell lines tested.

Several researchers have also evaluated the cytotoxic effects of ursolic acid and eichlerianic acid [18-21] isolated from other plant species in several cancer cell lines including MCF-7 breast cancer cells [22,23] and those results are some

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Position	Reported data [15]		Observed data		
	δC	δ Η (J Hz)	δС	δ Η (J Hz)	
1	38.6	1.72 (m)	38.8	1.74 (m)	
2	28.2	1.60 (m)	28.0	1.60 (m)	
3	78.7	3.22 (dd; J = 10.8; 4.9 Hz)	78.9	3.16 (dd; <i>J</i> = 4.5, 9.0 Hz)	
4	38.5		38.5		
5	55.2	1.34 (m)	55.1		
6	18.3	1.60 (m)	18.2	1.60 (m)	
7	32.9	1.72 (m)	32.9	1.74 (m)	
8	39.5		39.0		
9	47.3	1.60 (m)	47.4	1.60 (m)	
10	37.0		36.7		
11	23.7	1.91 (m)	23.4	1.91 (m)	
12	125.9	5.27 (dd; J = 3.6; 3.5 Hz)	125.0	5.18 (dd; ; <i>J</i> = 2.8; 5.7 Hz)	
13	137.9		138.0		
14	42.0		42.0		
15	28.1	1.60 (m)	28.0	1.60 (m)	
16	25.0	1.34 (m)	24.1	1.33 (m)	
17	48.1		48.1		
18	53.8	2.2 (m)	52.72	2.10 (m)	
19	38.5	1.00 (m)	38.8	1.04 (m)	
20	38.5	0.95 (m)	38.8	0.98 (m)	
21	30.3	1.27 (m)	30.6	1.27 (m)	
22	37.4	1.72 (m)	37.4	1.74 (m)	
23	28.9	1.00 (s)	32.9	0.94 (s)	
24	15.6	0.79 (s)	15.5	0.72 (s)	
25	15.4	0.94 (s)	15.3	0.86 (s)	
26	17.1	0.82 (s)	16.9	0.94 (s)	
27	23.5	1.10 (s)	23.5	0.75 (s)	
28	179.6		180.2	1.03 (s)	
29	17.0	0.87 (dd; <i>J</i> = 6.4 Hz)	16.8	0.67 (d; <i>J</i> = 10.0 Hz)	
30	21.4	0.97 (dd; <i>J</i> = 6.3 Hz)	21.1	0.80 (d; <i>J</i> = 5.5 Hz)	

Table 2:¹³C NMR (150MHz) and ¹H NMR data (600 MHz) of ursolic acid (2) (CDCI₃)



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Figure 2: Morphological changes in NCI-H292 lung cancer cells after exposure to oleanolic acid (compound 1), ursolic acid (compound 2) and eichlerianic acid (compound 3)

Figure 3: Morphological changes in MCF-7 breast cancer cells after exposure to oleanolic acid (compound 1), ursolic acid (compound 2) and eichlerianic acid (compound 3)

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Position	Reported data [14]		Observed data	
	δС	δH (JHz)	δС	δ Η (J Hz)
1		1.75 (m)	34.2	1.84 (m)
	34.3	1.53 (m)		1.60 (m)
2	28.5	2.14 (m)	28.1	2.18
		2.32 (m)		2.38
3	179.5	-	179.0	-
4	147.5	-	147.4	-
5	50.8	-	49.7	-
6	24.6	-	24.6	-
7	33.9	1.15 (m)	33.8	1.07 (m)
8	40.0	-	40.0	-
9	41.2	1.43 (m)	41.1	1.48 (m)
10	39.0	-	39.0	-
11	22.3	1.35 (m)	22.3	1.39 (m)
12	26.9		26.8	-
13	42.9	1.60 (m)	42.8	1.65 (m)
14	50.3	-	50.3	-
15	31.4	1.40 (m)	31.4	1.45 (m)
16	25.8	1.75 (m)	25.8	1.75 (m)
17	49.7	1.80 (m)	49.7	1.84 (m)
18	16.3	0.82 (s)	16.3	0.87 s
19	20.2	0.79 (s)	20.2	0.84 s
20	86.6	-	86.5	-
21	27.1	1.08 (s)	27.1	1.13 s
22	34.7	1.57 (m)	34.7	1.60 (m)
23	26.3	1.80 (m)	26.3	1.83 (m)
24	86.3	3.57 (dd, <i>J</i> = 10.3, 5.0	86.3	3.62 (dd, <i>J</i> =
		Hz)		5.5,10.0 Hz)
25	70.3		70.2	-
26	27.8	1.13 (s)	27.8	1.17 s
27	23.2	1.05 (s)	23.2	1.09 s
28		4.60 (brs)	113.4	4.64 brs
	113.4	4.78 (brs)		4.83 brs
29	24.0	1.67 (s)	24.0	1.71 s
30	15.3	0.99 (s)	15.3	0.99 s

Table 3:13C NMR (150MHz) and	¹ H NMR data (600 MHz	c) of compound 3 eichlerianic acid ((CDCl ₃)
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what in accordance with the results obtained in the present study. Results of the present study illustrate cytotoxic effects of ursolic acid and eichlerianic acid isolated first time from the leaves of *S. hydrophyllacea*.

CONCLUSION

The findings of the present study indicate that ursolic acid and eichlerianic acid isolated from the leaves of *S. hydrophyllacea* grown in Sri Lanka have potentials for the treatment of breast and lung cancer. Detailed molecular studies are necessary to elucidate the anti-cancer mechanisms of these two compounds.

DECLARATIONS

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Conflict of interest

No conflict of interest is associated with this work.

Author contributions

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Author SRS was involved in designing the study. Authors SRS, MKE, LW, NF, LT and PP were involved in conducting the experiment, AA was involved in structure elucidation of compounds. MKE drafted the manuscript and SRS, MKE, PP and KHT were involved in analyzing data. Manuscript was revised by authors SRS, MKE, AA and KHT. All authors read and approved the final manuscript.

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