Tropical Journal of Pharmaceutical Research December 2018; 17 (12): 2445-2448

ISSN: 1596-5996 (print); 1596-9827 (electronic)

© Pharmacotherapy Group, Faculty of Pharmacy, University of Benin, Benin City, 300001 Nigeria.

Available online at http://www.tjpr.org http://dx.doi.org/10.4314/tjpr.v17i12.20

Original Research Article

In-vitro studies on acetylcholinesterase and butyrylcholinesterase inhibitory potentials of aerial parts of Vernonia oligocephala (Asteraceae)

Waqas Mahmood¹, Hammad Saleem^{2,3*}, Irshad Ahmad¹, M Ashraf⁴, M. Shoaib Ali Gill², Hafiz Muhammad Ahsan^{5,6}, Kashif-ur-Rehman Khan², Sadia Chaman², Seema Abbas², Anam Mubashar², Shafi Ullah Khan³, Nafees Ahemad³

¹Department of Pharmacy, The Islamia University of Bahawalpur, Bahawalpur, 63100, Pakistan, ² Institute of Pharmaceutical

¹Department of Pharmacy, The Islamia University of Bahawalpur, Bahawalpur, 63100, Pakistan, ²Institute of Pharmaceutical Sciences (IPS), University of Veterinary and Animal Sciences (UVAS), Lahore, 54000, Pakistan, ³School of Pharmacy, Monash University, Jalan Lagoon Selatan, 47500 Bandar Sunway, Selangor Darul Ehsan, Malaysia, ⁴Department of Chemistry, The Islamia University of Bahawalpur, Bahawalpur, 63100, Pakistan, ⁵School of Pharmacy, Jiangsu University, Zhenjiang, Jiangsu, P.R. China 212013. ⁶Faculty of Pharmacy, University of Central Punjab, 54000, Lahore, Pakistan

*For correspondence: Email: hammad.saleem@monash.edu_hammad.saleem@uvas.edu.pk; Tel: +92-3357885316

Revised accepted: 8 October 2018

Abstract

Purpose: To evaluate the cholinesterases (acetylcholinestrase and butyrylcholinestrase) enzyme inhibitory potential of aerial parts Vernonia oligocephala (V. oligocephala) as a potential remedy for the management of neurodegenerative disorders.

Methods: Crude methanol extract of V. oligocephala leaves and its fractions including n-hexane, dichloromethane (DCM), ethyl acetate and n-butanol were investigated for their inhibitory potentials against cholinesterases enzymes employing standard ELISA microtiter plate reader assay.

Results: All the extracts were moderately active against both the tested enzymes. The ethyl acetate and n-hexane fractions showed highest inhibition against acetylcholinesterase enzyme with IC_{50} values of $128.81\pm0.44~\mu g/mL$ and $200.51\pm0.22~\mu g/mL$, respectively. In case of butrylcholinesterase, ethyl acetate (IC_{50} ; $145.71\pm0.19\mu g/mL$) and DCM (IC_{50} ; $269.31\pm0.45~\mu g/mL$) fractions were the most active. All other fractions including crude methanol extract exhibited least inhibition. Overall, ethyl acetate fraction was the most active against both enzymes.

Conclusion: Results indicate the promising potential of V.oligocephala as source of new potential compounds for management of Alzheimer's disease. However, further studies to isolate and identify the potential bioactive components are needed.

Keywords: Vernonia oligocephala, Acetylcholinesterase, Butyrylcholinesterase, Alzheimer's disease

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, International Pharmaceutical Abstract, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

INTRODUCTION

Alzheimer's disease is linked with the loss of cholinergic neurons in different parts of the brain and it is the common cause of dementia in older people [1]. Cortical cholinergic projections arising from the basal forebrain play critical role in the memory component of both Alzheimer and Parkinson types of dementia [2]. Alzheimer's disease originally posit a connection between disturbances in cerebral cholinergic neurotransmission and some of the cognitive impairments especially memory [3]. Symptoms of the disease can be modified by most of the cholinomimetic drugs [4]. Acetylcholine (ACh) and butyrylcholine (BCh) inhibitors are the approved agents Food by and Drua Administration (FDA) for the treatment of Alzheimer's disease (AD) [5]. The serious side effects of synthetic licensed drugs which are used to treat AD have forced the researchers to investigate safer AChE or BChE inhibitors from natural sources [6]. In traditional practices of medicine, numerous plants and their constituents are reported to enhance cognitive functions and to alleviate other symptoms of AD [7]. The discovery of AChE or BChE enzymes inhibitors from the medicinal plants have attracted the attention of the scientists and open a new way in research field [8].

The genus Vernonia is one of the major genus of family Asteraceae consisting of about 650 species [9]. The chemical constituents of this family mainly include volatile oils, terpenoids, phenolic compounds and tannins [10]. V. oligocephalais one of the important perennial herb of this genus traditionally used in treating asthma, colics, inflammation, ophthalmology, pruritus, and to prevent gonorrhea, tetanus and urinary tract infections [11]. The purpose of this study was to evaluate the possible inhibitory effects of the extracts against AChE or BChE.

EXPERIMENTAL

Chemicals and Standards

All enzymes were purchased from Sigma-aldrich and used without further purification. These are AChE Electric eel (Sigma-Aldrich GmbH, USA), BChE equine serum lyophilized (Sigma-Aldrich GmbH, USA), substrates: acetylthiocholine iodide (Sigma-Aldrich, U.K.), butyrylthiocholineiodide (Sigma-Aldrich, Switzerland), DTNB (5,5dithio-bis-nitrobenzoic acid) (Sigma-Aldrich, Germany) andeserine Sp. (Sigma-Aldrich, dipotassium France). Chemicals including $(K_2HPO_4),$ potassium hydrogen phosphate dihydrogen phosphate (KH₂PO₄), potassium hydroxide were of extra pure analytical grade from Merck Group (Darmstadt, Germany) were used.

Plant material and extraction

Aerial parts of *V. Oligocephala* were collected from the peripheral areas of Bahawalpur and identified by Dr. Muhammad Arshad Chaudhry (late); Director, Cholistan Institute of Desert Studies (CIDS), The Islamia University of Bahawalpur and a voucher specimen number was also deposited for future reference.

The shade dried leaves of *V. oligocephala* were macerated in 80% w/v methanol for 7 days with periodic shaking, filtered and then concentrated using a rotary evaporator at 37°C. Concentrated methanol crude extract was further subjected to fractionation with *n*-hexane, dichloromethane (DCM), ethyl acetate and *n*-butanol [thrice with each solvent]. The filtrates of the respective fractions thus obtained were again concentrated by rotary evaporator. The respective concentrates, VOAC (V. oligocephala aerial crude methanol extract), VOAD (V. oligocephala aerial DCM extract), VOAE (V. oligocephala VOAH (V. aerial ethyl acetate extract), oligocephala aerial n-hexane extract) and VOAB (V. oligocephala aerial n-butanol extract), were weighed and stored in labelled airtight vials in refrigerator.

Acetylcholinesterase assay

The AChE inhibition activity was performed a slight modification of the protocol previously reported [12]. Total volume of the reaction mixture was 100 µL. It contained 60 µL Na₂H PO₄ buffer with concentration of 50 mM having pH 7.7. 10 µL test compound (0.5 mM/well) was added, followed by the addition of 10 μ L (0.005 unit/well) enzyme. The contents were mixed and pre-read at 405 nm. Then, contents were preincubated for 10 min at 37°C. The reaction was initiated by the addition of 10 µL of 0.5 mM/well substrate (acetylthiocholine iodide), followed by the addition of 10 µL DTNB (0.5 mM/well). After 30 min of incubation at 37°C, absorbance was measured at 405 nm using 96-well plate reader (Synergy HT, Biotek, USA). All experiments were carried out with their respective controls in triplicate. Eserine (0.5 mM/well) was used as a positive control. Percentage inhibition was calculated as follows:

Inhibition
$$\cdot$$
 (%) = $\frac{Control - Test}{Control} x \cdot 100$ (1)

where Control is the total enzyme activity without inhibitor and Test is the enzyme activity in the presence of test compound.

IC₅₀ values (concentration at which there is 50 % enzyme catalyzed reaction) were calculated using EZ–Fit Enzyme kinetics software (Perrella Scientific Inc. Amherst, USA).

Butyrylcholinesterase assay

The BChE inhibition activity was performed using a slight modification of the method previously reported [12]. Total volume of the reaction mixture was 100 μL containing 60 μL, Na₂HPO₄ buffer, and 50mM having pH 7.7. 10 µL test compound 0.5 mM/well was added followed by the addition of 10 µL (0.5 unit/well) BChE. The contents were mixed and pre-read at 405 nm and then pre-incubated for 10 min at 37°C. The reaction was initiated by the addition of 10 µL of mM/well substrate (butyrylthiocholine chloride), followed by the addition of 10 µL DTNB, 0.5 mM/well. After 30 min of incubation at 37°C, absorbance was measured at 405 nm using 96-well plate reader (Synergy HT, Biotek, USA). All experiments were carried out with their respective controls in triplicate. Eserine (0.5 mM/well) was used as positive control. Inhibition (%) was calculated using above Eq 1.

Statistical analysis

The results obtained were expressed as mean value and standard deviation. All the tests were performed for three times and the data analysis was carried out in triplicates. To calculate IC_{50} , graph Pad Prism software (Version 6.03) was used One-way analysis of variance (ANOVA) and Tukey's significant difference post hoc test (p<0.05) were used to calculated differences. SPSS v22.0 software was used to carry out all experimental analysis.

RESULTS

The results of percentage AChE and BChE enzyme inhibition (at the concentration of 0.5 mg/mL) and half maximal concentration (IC $_{50}$) values of crude methanol extract and different fractions of V. oligocephala leaves were calculated. The results of AChE inhibition are presented in Table 1.

The effect of the *V. oligocephala* extract and its fractions on the inhibitory activity against BchE are depicted in Table 2.

Table 1: Percentage enzyme inhibition and IC_{50} values of V. oligocephala extract and fractions against acetylcholinesterase

Sample code	AChE inhibition (%)	AChEIC ₅₀ (μg/mL)
VOAC	52.41±0.39	**>500
VOAD	23.31±0.34	**>500
VOAE	73.79±0.11	128.81±0.44
VOAH	68.01±0.41	200.51±0.22
VOAB	43.01±0.21	**>500
Eserine	91.29±1.17	0.04±0.0001

Values are expressed as means ± SD of three replicates; ** IC₅₀ value was higher than 500 μg/mL, AChE= acetylcholinesterase. Eserine was used as positive control.

Table 2: Percentage enzyme inhibition and IC₅₀ values of *V. oligocephala* extract and fractions against butyrylcholinesterase

Sample code	BChE inhibition (%)	BChEIC ₅₀ (μg/mL)
VOAC	29.44±0.55	**>500
VOAD	60.78±0.11	269.31±0.45
VOAE	74.25±0.44	145.71±0.19
VOAH	44.01±0.41	**>500
VOAB	34.63±0.34	**>500
Eserine	82.82+1.09	0.85+0.0001

Values are expressed as means \pm S.D. of three replicates; **IC $_{50}$ value was higher than 500 μ g/mL, BChE= butyrylcholinesterase. Eserine was used as positive control.

DISCUSSION

This study has demonstrated that the aerial parts of *Vernonia oligocephala* has moderate inhibitory activity against cholinesterases (acetylcholinestrase and butyrylcholinestrase) enzymes with IC $_{50}$ values in the range of 128.81 \pm 0.44 - 269.31 \pm 0.45 µg/mL. The methanol crude extracts of the plant did not produce any significant inhibition of cholinesterases unlike the n-hexane and ethyl acetate extracts.

The aerial part of V. oligocephala is known to possess sesquiterpene lactones, triterpene (3βacetoxyneohop-13(18)-ene) along sitosterol, 3β-hydroxylup-20,29-ene, 3β-hydroxyolean-12en-28-oic acid and β-sitosterol-3-O-β-Dglucopyranoside [13]. The AChE inhibitory potential of V. oligocephala extracts can be occurrence attributed to the sesquiterpenes in this plant [14-16]. Similarly, as alkaloids, such galantamine, physostigmine [17], and other compounds like glycosides, terpenoids, flavonoids [18], lignans[19], coumarins [20], ursolic acid [21] terpenoid, flavonoids and phenolic compounds are known potent cholinesterases inhibitors [2224]. Several *venonia* species have been found to contain phenolic and flavonoids like naringenin, apigenin, protocatechuic acid and luteolincatechin [25]. These compounds could also have contributed to the observed bioactive potential of *V. oligocephala* reported in this study.

CONCLUSION

The leaves of *V. oligocepha* has inhibitory activity against both acetylcholinesterase and butyrylcholinesterase enzymes. Although further studies are required to isolate the compound responsible for the observed bio-activity, the plant could be considered for further studies in the treatment of neurodegenerative disorders.

DECLARATIONS

Conflict of interest

The authors declare that they have no conflict of interest with regard to this work.

Authors' contribution

We declare that this work was performed by the authors named in this article, and all liabilities pertaining to claims relating to the content of this article will be borne by the authors.

REFERENCES

- Nordberg A, Svensson AL. Cholinesterase inhibitors in the treatment of Alzheimer's disease. Drug safety. 1998; 19: 465-80.
- 2. Perry EK. The cholinergic hypothesis—ten years on. British Medical Bulletin. 1986; 42:63-9.
- Wallin A, Blennow K, Andreasen N, Minthon L. CSF biomarkers for Alzheimer's Disease: levels of βamyloid, tau, phosphorylated tau relate to clinical symptoms and survival. Dement Geriatric Cogn Dis. 2006; 21: 131-8.
- Birks J. Cholinesterase inhibitors for Alzheimer's disease. Cochrane Database Syst Rev. 2006; 1: CD005593.
- Akhondzadeh S, Noroozian M, Mohammadi M, Ohadinia S, Jamshidi A, Khani M. Salvia officinalis extract in the treatment of patients with mild to moderate Alzheimer's disease: a double blind, randomized and placebo-controlled trial. J. Clin. Pharm. Tther. 2003; 28:53-9
- Saleem H, Ahmad I, Shahid MN, Gill M, Nadeem MF, Mahmood W. In vitro acetylcholinesterase and butyrylcholinesterase inhibitory potentials of Jatropha gossypifolia plant extracts. Acta Pol. Pharm. 2016; 73: 419-23.
- 7. Talic S, Dragicevic I, Corajevic L, Martinovic Bevanda A. Acetylcholinesterase and butyrylcholinesterase inhibitory activity of extracts from medicinal plants. Bull Chem Technol Bosnia Herzegovina. 2014; 43: 11-4.
- 8. Asanuma M, Miyazaki I, Ogawa N. Dopamine-or L-

- DOPA-induced neurotoxicity: the role of dopamine quinone formation and tyrosinase in a model of Parkinson's disease. Neurotox Res. 2003; 5: 165-76.
- Kirtikar KR, Basu BD. Kirtikar and Basu's Illustrated Indian Medicinal Plants: Their Usage in Ayurveda and Unani Medicines: Sri Satguru Publications, a division of Indian books centre; 2000.
- 10. Evans WC. Trease and Evans' Pharmacognosy E-Book: Elsevier Health Sciences: 2009.
- 11. Duke JA. Duke's handbook of medicinal plants of Latin America: CRC Press; 2008.
- Ellman GL, Courtney KD, Andres Jr V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem Pharmacol. 1961; 7: 88-95.
- Riaz N, Feroze K, Saleem M, Musaddiq S, Ashraf M, Alam U, Mustafa R, Jabeen B, Ahmad I, Jabbar A. Oligocephlate, a new α-glucosidase inhibitory neohopane triterpene from Vernonia oligocephala._J Chem Soc Pak. 2013:35(3): 972-975
- Owokotomo IA, Ekundayo O, Abayomi TG, Chukwuka AV. In-vitro anti-cholinesterase activity of essential oil from four tropical medicinal plants. Toxi Reports. 2015;2:850-857
- Loizzo MR, Tundis R, Conforti F, Menichini F, Bonesi M, Nadjafi F, Frega N, Menichini F. Salvia leriifolia Benth (Lamiaceae) extract demonstrates in vitro antioxidant properties and cholinesterase inhibitory activity. Nutr Res. 2010; 30:823-830
- Patel MB, Amin D. Sphaeranthus indicus flower derived constituents exhibits synergistic effect against acetylcholinesterase and possess potential antiamnestic activity. J Complement Integr Med. 2012; 9(1) 10.1515/1553-3840.1618
- 17. Mukherjee PK, Kumar V, Mal M, Houghton PJ. Acetylcholinesterase inhibitors from plants. Phytomedicine. 2007; 14:289–300.
- Oinonen PP, Jokela JK, Hatakka AI, Vuorela PM. Linarin, a selective acetylcholinesterase inhibitor from Mentha arvensis. Fitoterapia. 2006;77:429–34.
- 19. El-Hassan A, El-Sayed M, Hamed Al, Rhee IK, Ahmed AA, Zeller KP, et al. Bioactive constituents of Leptadeniaarborea. Fitoterapia. 2003;74: 184–7.
- Rollinger JM, Hornick A, Langer T, Stuppner H, Prast H. Acetylcholinesterase inhibitory activity of scopolin and scopoletin discovered by virtual screening of natural products. J Med Chem. 2004;47:6248–54.
- 21. Chung YK, Heo HJ, Kim EK, Kim HK, Huh TL, Lim Y, et al. Inhibitory effect of ursolic acid purified from Origanum majoranaL on the acetylcholinesterase. Mol Cells. 2001;11: 137–43.
- Uysal S, Zengin G, Locatelli M, Bahadori MB, Mocan A, Bellagamba G. Cytotoxic and enzyme inhibitory potential of two Potentilla species (P. speciosa L. and P. reptans Willd.) and their chemical composition. Front. Pharmacol. 2017; 8: 290.
- Stasiuk M, Bartosiewicz D, Kozubek A. Inhibitory effect of some natural and semisynthetic phenolic lipids upon acetylcholinesterase activity. Food Chem. 2008; 108: 996-1001.
- Bahadori MB, Dinparast L, Valizadeh H, Farimani MM, Ebrahimi SN. Bioactive constituents from roots of Salvia syriaca L.: acetylcholinesterase inhibitory activity and molecular docking studies. S Afr J Bot. 2016; 106: 1-1
- 25. Silvada LAL, Faqueti LG, Reginatto FH, Santosdos ADC, Barison A, Biavatti MW. Phytochemical analysis of Vernonanthura tweedieana and a validated UPLC-PDA method for the quantification of eriodictyol. Revista Brasileira de Farmacognosia. 2015; 25: 375-381..