

## Original Research Article

# Targeting GSK-3 $\beta$ enzyme by diazepino-quinolone derivatives

Lubna Swellmeen<sup>1\*</sup>, Haneen A Basheer<sup>2</sup>, Amal Uzrail<sup>3</sup>, Haneen Sallam<sup>2</sup>, Yusuf Al-Hiari<sup>4</sup>, Ahlam Alkilani<sup>2</sup>

<sup>1</sup>Department of Pharmaceutical Chemistry, Faculty of Pharmaceutical Sciences, Hashemite University, Zarqa, Jordan,

<sup>2</sup>Department of Pharmaceutical Sciences, Faculty of Pharmacy, Zarqa University, Zarqa 13132, <sup>3</sup>Department of Medical Analysis, Faculty of Sciences, Al-Albaysat University, <sup>4</sup>Faculty of Pharmacy, The University of Jordan, Amman, Jordan

\*For correspondence: **Email:** [lubnam@hu.edu.jo](mailto:lubnam@hu.edu.jo); **Tel:** +962 778004500, 39033335

Sent for review: 15 June 2022

Revised accepted: 24 September 2022

### Abstract

**Purpose:** To synthesize a heterocyclic system containing quinolone and diazepine scaffolds as GSK-3 $\beta$  inhibitor.

**Methods:** The diazepino-quinolone derivatives were synthesized starting from quinolone nucleus in a simple chemical reaction. The *in vitro* GSK-3 $\beta$  enzyme assay and MTT assay against cancer cell lines were carried out followed by Z'-LYTE GSK-3 $\beta$  assay. Anticancer activity was determined using U-87 glioma cell line.

**Results:** Diazepino-quinolone derivatives were obtained in a good yield, and compound 102 exhibited significant activity against *in vitro* GSK-3 $\beta$  (IC<sub>50</sub>: 0.114  $\mu$ M), and anticancer activity (IC<sub>50</sub>: 37  $\mu$ M) against U-87 glioma cell line.

**Conclusion:** The GSK-3 $\beta$  enzyme is a potential target to treat different diseases, and diazepines derivatives are a successful template for inhibitors design against GSK-3 $\beta$  enzyme with IC<sub>50</sub> in a micromolar range.

**Keywords:** GSK-3 $\beta$ , Heterocyclic compounds, Quinolone, Benzodiazepine nucleus

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, Web of Science, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

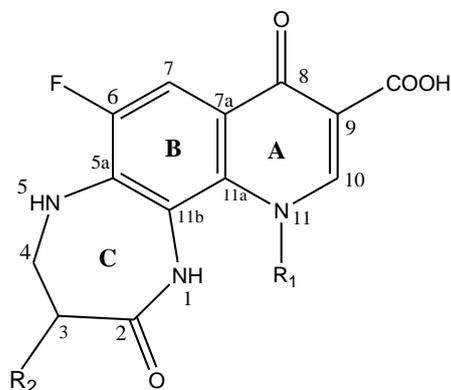
## INTRODUCTION

One of the most important serine/threonine kinase enzymes is the GSK-3 $\beta$  enzyme that is active, pervasive, and vital for life [1]. The GSK-3 $\beta$  enzyme is a key signaling molecule that is involved in insulin and glycogen metabolism [2] and inflammatory responses. It plays a significant role in viral infections [3,4] and in cancer [5]. Its expression in cancer has been associated with

survival, proliferation, and migration of cancer cells and their resistance to chemotherapy [6].

The GSK-3 $\beta$  enzyme inhibitors have gained a lot of interest as a potential treatment of neurodegenerative diseases such as Alzheimer's disease [7], diabetes [2] and malignancies [8,9]. The enzyme inhibitors were previously synthesized from heterocyclic compounds having a quinoxaline nucleus that showed good inhibitory activity against GSK-3 $\beta$  enzyme [10].

Diazepine derivatives exhibit many biological activities such as anti-leukemia and anti-platelet activities [11], anti-tumor [12], anti-malarial, anti-trypanosomal, anti-leishmanial agents [13], and anti-convulsant [14]. This study introduces a new hetero-tricyclic system that contains diazepine nucleus (Figure 1) as a potential GSK-3 $\beta$  enzyme inhibitor.



R<sub>1</sub>: Cyclopropyl, or p-fluorophenyl  
R<sub>2</sub>: H, or Methyl

**Figure 1:** General chemical structure of diazepine derivatives

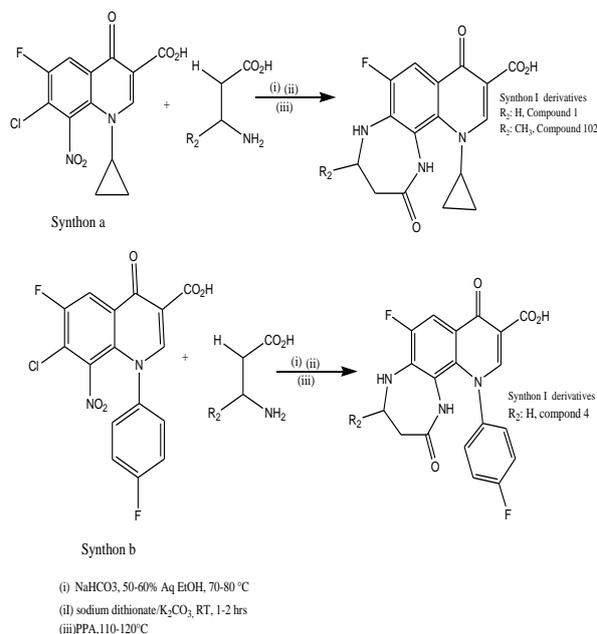
## EXPERIMENTAL

### Equipment and reagents

Melting point were measured by Stuart melting point apparatus, IR spectrum was recorded by Shimadzu 8400F FT-IR spectrophotometer. Bruker, Advance DPX-300 Nuclear magnetic resonance spectrometer was used to record (NMR) spectra. High-resolution mass spectrophotometer was used for mass instrument. Reagents and chemicals used are of synthetic grade and were bought from Sigma-Aldrich.

### Synthon I derivatives

The synthetic procedure used in scheme I is based on reported methods [10,15]. By applying three steps of reaction, synthon I was obtained: The first step was done by reacting synthon a, and b with different 3-aminopropanoic acid derivatives under conditions shown in Figure 2, and the reaction was done under reflux at 80 °C then solution of sodium dithionite (8.0 g, 46 mmol) was added. Finally, a reflux condition was used and polyphosphoric acid was added (10 mL) to get synthon I.



**Figure 2:** synthesis scheme for synthon I

### GSK 3 $\beta$ *in vitro* assay

The Z'-LYTE GSK-3 $\beta$  assay was used to make GSK-3 $\beta$  *in vitro* assay test (Z'-LYTETM Screening Protocol and Assay Conditions 2016); Z'-LYTE test is used to get the % Inhibition.

The compounds that have the highest percentage inhibition were tested for their IC<sub>50</sub> values. Stock solutions of 10 mM concentration of the tested compounds (1, 4, and 102) in dimethyl sulfoxide (DMSO) was prepared. Then each was tested at 10  $\mu$ M using the Select Screen Kinase Profiling Services (ThermoFisher Scientific, USA) [16].

### MTT assay

The cytotoxicity of GSK-3 $\beta$  inhibitors was determined by MTT assay protocol based on reported procedure carried out by Al-Sha'er *et al* [17]. The U-87 glioma cell line was used for the MTT assay and was provided by Dr. Ahmad Sharab, of the American University of Madaba while the chemicals were purchased from Sigma Aldrich.

### Molecular modeling studies

In order to support the *in vitro* assay results, a molecular docking procedure was carried out. The GSK-3 $\beta$  enzyme (PDB code: 3Q3B, resolution 2.7 Å) binding pocket were determined and used in the docking procedure and the most active compound was docked using the Dock Ligands (LibDock).

## RESULTS

### Spectral characteristics

Based on the synthetic approach in Figure 2, the following compounds were obtained:

#### 11-Cyclopropyl-2,8-dioxo-6-fluoro-2,3,4,5,8,11-hexahydro-1H-(1,4)diazepino(2,3-h)quinoline-9-carboxylic acid (1)

Brownish product was obtained. Yield 0.36 g (95 %); mp 346 - 347 ° [16].

#### 6-fluoro-11-(4-fluorophenyl)-2,8-dioxo-2,3,4,5,8,11-hexahydro-1H-(1,4)diazepino(2,3-h)quinoline-9-carboxylic acid (4)

Yellow powder, mp 299 - 302 °C. <sup>1</sup>H-NMR (300 MHz, DMSO, *d*<sub>6</sub>): δ 2.33 (m, 2H, CH<sub>2</sub>-3), 3.01 (m, 2H, CH<sub>2</sub>-4), 6.89 (t, 1H, NH-CH<sub>2</sub>), 7.30 (d, *J* = 11.4 Hz, 2H, H-3'/ H-5'), 7.64(d, *J* = 8.4 Hz, 2H, H-2'/ H-6'), 7.75 (d, <sup>3</sup>*J*<sub>H-F</sub> = 11.4Hz, 1H, H-7), 8.42 (s, 1H, H-10), 8.62 (s, 1H, N(1)-H), 14 (br s, 1H, C(9)-CO<sub>2</sub>H); HERMS (ESI, +ve): *m/z* (M<sup>+2</sup> + H) 387.10306 C<sub>19</sub>H<sub>15</sub>F<sub>2</sub>N<sub>3</sub>O<sub>4</sub> 387.1030.

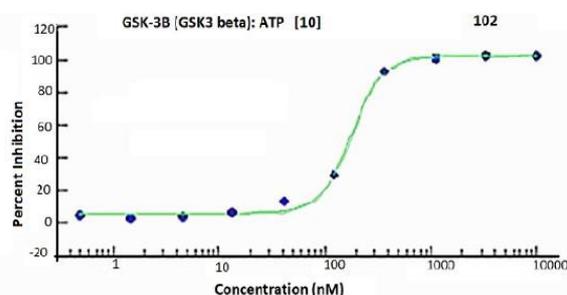
#### 11-cyclopropyl-6-fluoro-3-methyl-2,8-dioxo-2,3,4,5,8,11-hexahydro-1-H-(1,4)diazepino(2,3-h)quinoline-9-carboxylic acid (102)

Brownish powder, mp 309 - 315 °C; <sup>1</sup>H-NMR (300 MHz, DMSO- *d*<sub>6</sub>): δ 0.81 (m, 2H, H<sub>2</sub>-2'), 1.54 (d, *J* = 5.1 Hz, 3H, CH<sub>3</sub>), 3.24 - 3.66 (m, 3H; 1H, H-3 and 2H, H-4), 3.47 (d, d, *J* = 12, 10.5 Hz, 1H, H α-4), 3.66 (s, 1H, Hβ-4), 4.21 (m, 1H, H-1'), 6.88 (br s, 1H, N(5)-H), 7.71 (d, <sup>3</sup>*J*<sub>H-F</sub> = 11.1 Hz, 1H, H-7), 8.66 (s, 1H, H-10), 9.62 (s, 1H, N(1)-H), 16 (s, 1H, CO<sub>2</sub>H); IR (NaCl): ν 3500, 3250, 2999, 2355, 1710, 1659, 1550, 1250 cm<sup>-1</sup>.

### GSK-3β inhibitors activity

The synthon I derivatives were tested by using the *in vitro* human recombinant GSK-3β kinase (human recombinant) assay kit. The GSK-3β inhibitors was screened in this assay at 10 nM. Compounds that showed good percentage inhibition, namely, (1, 4, and 102) were screened at different concentrations to find their IC<sub>50</sub> values. Compounds (1, 4, and 102) activities

were measured against GSK 3 enzyme and the results are shown in Table 1. As presented in the Table; compound 1 showed 79 % inhibition while 4 showed 11 % inhibition and compound 102 showed 100 % inhibition the IC<sub>50</sub> was calculated for compounds 1 and 102 and it was 4.18 and 0.114 μM, respectively. Figure 3 illustrates the concentration (nM)/percent inhibition plots of the most active compound 102. Next step was to determine if these compounds have any pharmacological activities as potential anti-cancer treatments. The U-87 glioma cell line was used in the MTT assay as it is known to express GSK-3β [18]. The most active (compound 102) and the least active (compound 4) compounds from the enzyme assay were chosen to assess their anti-proliferative activity. The MTT results are in accordance with the enzyme assay results with IC<sub>50</sub> 37 μM and > 100 μM for compounds 102 and 4, respectively (Table 1). The results, confirm the importance of GSK-3β in cancer progression and open new avenues for the use of the synthon I derivatives GSK-3β inhibitors as anti-cancer drugs [18].



**Figure 3:** Plot of concentration (nM) vs. percent inhibition for compound (102)

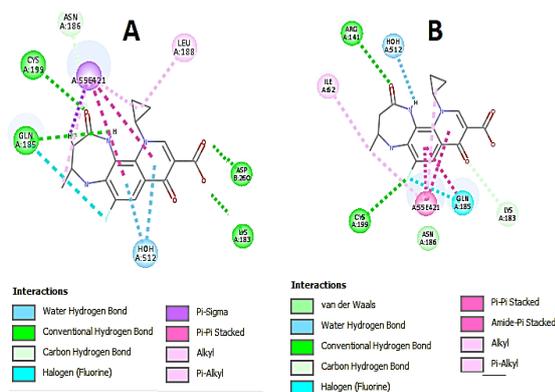
### Docking results

The software assists in identifying the binding site in the receptor of GSK-3β that binds to the synthesized compound and determined the conformations of the active compound 102 by docking procedure that allowed the types of interactions between the binding pocket and the active compound. Figure 4 shows the most active compound (102) with different poses that might be acquired inside the binding pocket of (3Q3B).

**Table 1:** The synthesized compounds tested against (1) GSK-3β enzyme presented with their % inhibition and IC<sub>50</sub> (μM). (2) U-87 glioma cell line in MTT assay presented with their % inhibition and IC<sub>50</sub> (μM)

Compound name	(ATP) Tested (μM)	% Inhibition mean	IC <sub>50</sub> (μM) GSK 3 enzyme	IC <sub>50</sub> (μM) MTT assay
102	Km app	100	0.114	37
1	Km app	79	4.180	NC
4	Km app	11	NC	>100

NC: not calculated



**Figure 4:** 2D diagram showing the interaction between docked compound 102;  $IC_{50}$  (0.114  $\mu$ M) and GSK-3 $\beta$  enzyme showing two different poses (in A and B) for the compound 102 in the binding pocket of GSK-3 $\beta$  enzyme

## DISCUSSION

Synthon I derivatives were obtained in a good yield and tested against GSK-3 $\beta$  enzyme with interesting inhibitory concentration as shown in Table 1. Compound 102 showed 100 % inhibition with  $IC_{50}$  (0.114  $\mu$ M), while compound 1, had 79 % inhibition with  $IC_{50}$  (4.180  $\mu$ M) while compound 4 had 11% inhibition so the  $IC_{50}$  was not measured and the  $IC_{50}$  can be explained by docking studies that revealed that there are many interactions for compound 1 GSK 3 $\beta$  inhibitor in the binding pocket of GSK-3 $\beta$  kinase (3Q3B), among these interactions, there is pi-pi stacking, pi-sigma, pi-alkyl, halogen, and H-bonding.

The results from this study, confirm the importance of GSK-3 $\beta$  in cancer progression and open new avenues for the use of the synthon I derivatives GSK-3 $\beta$  inhibitors as anti-cancer drugs [17]. The molecular docking studies also elucidated the binding modes of the compounds with the GSK-3 $\beta$  target

## CONCLUSION

Synthesis of novel GSK-3 $\beta$  enzyme inhibitors has been carried out using simple chemical reactions. Diazepine derivatives could be a successful template for the design of inhibitors against GSK-3 $\beta$  enzyme with  $IC_{50}$  in a micromolar range. Compound 102 shows potent inhibitory activity against both GSK-3 $\beta$  enzyme and U-87 glioma cell line in MTT assay. With these findings, efforts are expanding for further optimization of diazepine derivatives in order to develop potent inhibitors with potential therapeutic activity against GSK-3 $\beta$ .

## DECLARATIONS

### Acknowledgements

The authors thank the Deanship of Pharmaceutical Science at Hashemite University and Deanship of Faculty of Pharmacy at Zarqa University, Jordan.

### Funding

None provided.

### Ethical approval

None provided.

### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

### Conflict of Interest

No conflict of interest associated with this work.

### Contribution of Authors

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

### Open Access

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

## REFERENCES

- Woodgett JR. Molecular cloning and expression of glycogen synthase kinase-3/factor A. *EMBO J* 1990; 9: 2431–2438.
- Henriksen EJ, Dokken BB. Role of glycogen synthase kinase-3 in insulin resistance and type 2 diabetes. *Curr Drug Targets* 2006; 7: 1435-1441.
- Alfhili M, Al sughayir J, Mc Cubrey J, Mc Cubrey J, Akula Sh. GSK-3-associated signaling is crucial to virus

- infection of cells. *Biochim Biophys Acta Mol Cell Res* 2020; 1867(10): 118-767
4. Wu CH. Glycogen synthase kinase-3 regulates the phosphorylation of severe acute respiratory syndrome coronavirus nucleocapsid protein and viral replication. *J Biol Chem* 2009; 284(8): 5229-5239
  5. Luo J. Glycogen synthase kinase 3beta (GSK3beta) in tumorigenesis and cancer chemotherapy. *Cancer lett* 2009; 273: 194-200.
  6. Augello G, Emma MR, Cusimano A, Azzolina A, Montalto G, Mc Cubrey JA, Cervello M. The Role of GSK-3 in Cancer Immunotherapy: GSK-3 Inhibitors as a New Frontier in Cancer Treatment. *Cells* 2020; 9: 1427.
  7. Martinez A, Gi C, Perez DI. Glycogen synthase kinase 3 inhibitors in the next horizon for Alzheimer's disease treatment. *Int J Alzheimers Dis* 2011; 280502.
  8. Luo J. Glycogen synthase kinase 3beta (GSK3beta) in tumorigenesis and cancer chemotherapy. *Cancer lett* 2009; 273: 194-200.
  9. Zhang P, Min Z, Gao Y, Gao Y, Bian J, Lin X, He J, yong D, Li Y, Peng C, Cheng Y, Chu Y. Discovery of novel benzothiazepinones as irreversible covalent glycogen synthase kinase 3 $\beta$  inhibitors for the treatment of acute promyelocytic leukemia. *J Med Chem* 2021; 64: 7341–7358.
  10. Swellmeen L, Uzrail A, Shaheen R, AL-Hiari Y. Molecular modelling studies and synthesis of novel quinoxaline derivatives with potential inhibitory effect on GSK-3 $\beta$ . *Trop J Pharm Res* 2021; 20: 599-604
  11. Ramajayam R, Giridhar R, Yadav RR. Balaraman, Djaballah H, Shum D, Radu C. Synthesis, antileukemic and antiplatelet activities of 2,3-diaryl-6,7-dihydro-5H-1,4-diazepines. *Eur J Med Chem* 2008; 43: 2004-2010.
  12. Insuasty B, Orozco F, Lizarazo C, Quiroga J, Abonia R, Hursthouse M, Noguerras M, Cobo J. Synthesis of new indeno(1,2-e)pyrimido(4,5-b)(1,4)diazepine-5,11-diones as potential antitumor agents. *Bioorg Med Chem* 2008; 16: 8492-8500.
  13. Insuasty B, Ramírez J, Becerra D, Echeverry C, Quiroga J, Abonia R, Robledo S, Vélez I, Upegui Y, Muñoz J, et al. An efficient synthesis of new caffeine-based chalcones, pyrazolines and pyrazolo(3,4-b)(1,4)diazepines as potential antimalarial, anti-trypanosomal and antileishmanial agents. *Euro J Med Chem* 2015; 93: 401-413.
  14. El-Subbagh HS, Hassan G, El-Azab A, Abdel-Aziz A, Kadi A, Al-Obaid A, Al-Shabanah O, Sayed-Ahmed M. Synthesis and anticonvulsant activity of some new thiazolo(3,2-a)(1,3)diazepine, benzo(d)thiazolo(5,2-a)(12,6)diazepine and benzo(d)oxazolo(5,2-a)(12,6)diazepine analogues. *Euro J Med Chem* 2011; 46: 5567-5572.
  15. Al-Hiari Y, Abu-Dahab R, El-Abadelah M. Heterocycles {h}-Fused Onto 4-Oxoquinoline-3-Carboxylic Acid, Part VIII {1}. Convenient synthesis and antimicrobial properties of substituted hexahydro(1,4)diazepino(2,3-h)quinoline-9-carboxylic acid and Its Tetrahydroquino (7,8-b)benzodiazepine Analog. *Molecules* 2008; 13: 2880-2893.
  16. Select Screen TM biochemical kinase profiling service, Z\_LYTETM screening protocol, and assay conditions 2016 ®, USA. <https://www.thermofisher.com/jo/en/home/products-and-services/services/custom-services/screening-and-profiling-services/selectscreen-profiling-service/selectscreen-kinase-profiling-service.html>.
  17. A. Al Sha'er M, A. Basheer H, Taha O. Discovery of new PKN2 inhibitory chemotypes via QSAR guided selection of docking-based pharmacophores. *Mol Divers* 2022; 26: 11030-10434.
  18. Li Y, Lu H, Li G, Yan G. Glycogen synthase kinase-3B regulates astrocytic differentiation of U87-MG human glioblastoma cells. *Acta Pharmacol Sin* 2010; 31: 355-360.