### Tropical Journal of Pharmaceutical Research June 2016; 15 (6): 1225-1233

**ISSN:** 1596-5996 (print); 1596-9827 (electronic)

© Pharmacotherapy Group, Faculty of Pharmacy, University of Benin, Benin City, 300001 Nigeria.

All rights reserved.

Available online at http://www.tjpr.org http://dx.doi.org/10.4314/tjpr.v15i6.15

# **Original Research Article**

# Dose-dependent protection of reseveratrol against spinal cord ischemic-reperfusion injury in rats

## Meng-gin Su\*, Wei Zhong and Su-en Ren

Department of Anesthesiology, The Chest Hospital of Henan Province, Zhengzhou, Henan 450008, China

\*For correspondence: Email: sumengqinss@hotmail.com; Tel: 0086-371-65662780

Received: 4 November 2015 Revised accepted: 11 May 2016

#### Abstract

Purpose: To examine the protective effects of resveratrol (RESV) against spinal cord ischemic reperfusion (SCIR) injury.

**Methods:** Forty-eight male rats were divided into six groups: sham-operated (control-I), SCIR-treated (SCIR-II), rats receiving 20 mg/kg of RESV with SCIR (RESV 20+SCIR-III), rats receiving 40 mg/kg of RESV with SCIR (RESV 40+SCIR-IV), rats receiving 60 mg/kg of RESV with SCIR (RESV 60+SCIR-V), and rats receiving 50 mg/kg of methylprednisolone (MP) with SCIR (MP + SCIR-VI), for 7 days prior to IR (pre-treatment) and 7 days after IR (post-treatment).

**Results:** The levels of oxidative markers (TBARS, MPO) and inflammatory markers (IL-1β, IL-6, TNF-α, and NF-p65) were concomitantly suppressed in RESV-treated rats, which showed improved locomotor function. A pronounced increase in the activities of antioxidant enzymes (SOD, CAT and GSH) was noted in the RESV group compared with the MP and SCIR groups. RESV and MP supplementation increased neuronal count with decreased nuclear degeneration. RESV (40 mg) exhibited greater protective effect than 20 mg and 60 mg of RESV and 50 mg of MP.

**Conclusion:** The results show the neurotherapeutic potential of RESV (40 mg) to attenuate oxidative stress and the inflammatory response to SCIR injury.

**Keywords:** Spinal cord ischemia reperfusion, Resveratrol, Locomotor function, Antioxidant, Inflammatory markers

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, International Pharmaceutical Abstract, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

#### INTRODUCTION

Spinal cord ischemic reperfusion (SCIR) injury is a fatal postoperative complication of thoracic-abdominal aortic aneurysm surgery that contributes to paraplegia in about 10 % to 40 % of cases [1]. Paraplegia is a medical condition in which the sensory or motor part of the lower extremities is impaired due to spinal injury, resulting in long-term disability [2]. Numerous methods or techniques are recommended to treat paraplegic conditions, such as draining the cerebrospinal fluid, placement of shunts or performing a partial cardiopulmonary bypass,

hypothermia, and administration of pharmacological products such as steroids and vasodilators [3]. However, these protective effects do not completely revert the paraplegic condition. Instead, these treatment regimens lead to various adverse events that may worsen the paraplegic condition.

Many researchers have focused on various natural products that may suppress spinal cord ischemic injury (SCII) and subsequently prevent the paraplegic condition. Several pathophysiological events, including excitetoxicity (glutamate-mediated), energy failure,

inflammation, apoptosis, and oxidative stress, have been implicated in SCIR injury. Among these events, oxidative stress and inflammation are the major contributors and initiators of SCIR injury [4]. Therefore, choosing an antioxidant and an anti-inflammatory agent that alleviate SCIR injury would be a promising strategy. Recent studies have shown the protective effects of various natural antioxidant products against SCIR injury [5]. Methylprednisolone (MP) is a neuroprotective agent that possesses antioxidant and anti-inflammatory properties, and is recommended for treating spinal cord injuries [6]; thus, MP was used as a standard in the present study.

Resveratrol (RESV; 3,4',5-tri-hydroxy stilbene) is a phenolic compound commonly found in red wine/red grapes, soybeans and pomegranates [7]. RESV has two aromatic rings, with the free hydroxyl group contributing to its antioxidant and free radical scavenging activities [8]. RESV has been used as a potential drug for treating cardiovascular disease, diabetes, cancer, and various neurological disorders [9]. RESV exhibits neuroprotective activities due to its antioxidant, anti-inflammatory and anti-apoptotic properties [10].

RESV induces nitric oxide (NO) production, thereby acting as a vasodilatory agent [11]. Additionally, RESV readily crosses the bloodbrain barrier, which favors its neuroprotective effects in various ischemic models. Several studies have shown the neuroprotective effects of RESV against SCII in various animal models using different treatment regimens [12]. However, to date, no studies have explored the long-term, dose-dependent protective effects of RESV against SCIR injury. Moreover, a treatment regimen using RESV (combination of both pre- and post-treatment) remains to be established. Therefore, in this study, we explored the effective dose of RESV against SCIR injury based on a combination of both pre- and posttreatment patterns (pre- and post-ischemic conditions) by assessing locomotor capacity, neuronal viability, and oxidative and inflammatory markers in a rat model.

## **EXPERIMENTAL**

### **Drugs and reagents**

Resveratrol (RESV), Hematoxylin and eosin (H & E) and Methylprednisolone (MP) were procured from Sigma Aldrich (St. Louis, MO, USA). Paraformaldehyde, physiological saline (0.89 %), glycerol, sodium hydroxide, hydrogen peroxide were supplied by Lingjin Co. Ltd, Shanghai,

China. Phosphate buffered saline (PBS) readymade buffer Xylene was bought from Beijing Zhongshan Golden Bridge Biotechnology Co. Ltd., Beijing, China.

#### **Experimental animals**

Healthy, adult male Sprague-Dawley (SD) rats weighing 300–320 g were used for the experiment. Rats were housed in a steel cage under a 12/12-h light/dark cycle. Food and water were given ad libitum. The Ethics Committee of China approved the animal experiments for the present study (protocol ref no. 02315), which were conducted according to the guidelines from the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

SCIR injury was performed based on the method previously reported by Hwang [13] with slight modifications. Briefly, all rats were fasted overnight (12 h) and anesthetized with ketamine (50 mg/kg) injected intramuscularly. Body temperature was maintained at 37 °C using warming or heating pads and heating lamps. The hair around the neck region (carotid-inquinal) was shaved by placing the rats in the supine position. A polyethylene catheter (PE-50) was inserted into the tail artery to inject heparin and monitor the mean distal arterial pressure (MDAP). Thereafter, a Fogarty balloon catheter (Edwards Life Science, Shanghai, China) was inserted via the left femoral artery into the proximal descending thoracic aorta (approximately 11 cm from the insertion site), and the balloon was inflated with saline to induce spinal cord ischemia.

Proximal artery pressure was maintained at 80 mm Hg during aortic occlusion by draining the blood from the carotid artery into the external blood reservoir filled with heparinized saline. Immediate loss of pulse with decreased MDAP confirmed aortic occlusion. The balloon was deflated after 25 min of ischemia to initiate reperfusion by restoring blood flow. Then, the catheters were withdrawn and the rats were allowed to recover from anesthesia. All procedures were performed in sham-operated control rats, which served as negative controls, except for aortic occlusion. Mean arterial pressure, body temperature and heart beat (vitals) were continuously monitored before ischemic reperfusion.

### **Experimental protocol**

Forty-eight male SD rats were divided into six treatment groups with eight rats per group. Rats received saline (sham-operated control-I), rats

subjected to SCIR insult (SCIR-II), rats receiving 20 mg/kg b.wt of RESV dissolved in 2 % DMSO saline via i.p. (RESV 20+SCIR-III), rats receiving 40 mg/kg b. wt of RESV dissolved in 2 % DMSO saline via i.p. (RESV 40+SCIR-IV), rats receiving 60 mg/kg b. wt of RESV dissolved in 2 % DMSO saline via i.p. (RESV 60+SCIR-V) and rats receiving 50 mg/kg b. wt MP dissolved in saline via i.p. (MP + SCIR-VI) for 7 days prior IR (pre-treatment) and 7 days after IR (post-treatment).

### **Neurological assessment**

Locomotor function (hind limb movement) was tested by an independent observer blinded to the grouping and experimental design on days 1, 3, 5 and 7 after SCIR injury using the Basso Beattie Bresnahan (BBB) motor rating scale. The rating scale has 0 - 21 levels, where the value 0 (no detectable hind limb movement), 1 - 8 (rated for slight or higher movements of hindlimb joints), 9 (rated for dorsal stepping), 10 - 20 (rated for progressive improved walking ability) and 21 (normal movement).

After the last neurological assessment (on day 7), the rats in each group were subdivided into two subgroups for histological (n = 4) and biochemical analysis (n = 4). On the day 7, all rats were euthanized by i.p. pentobarbital sodium injection and spinal tissue were removed immediately and fixed in 4 % paraformaldehyde for morphological studies. Spinal tissue homogenate (10 %) was prepared in Tris-HCl buffer, centrifuged at 2500 g for 20 min and used for biochemical analysis.

#### **Evaluation of edema (water content)**

Edema (E) was detected using procedures previously published by Mdzinarishvili with a few modifications [14]. The wet and dry weights of spinal samples were measured to calculate the E (water content). E was calculated as the % difference between the two weights.

# Lipid peroxidation and antioxidant enzyme assays

The levels of superoxide dismutase (SOD), catalase (CAT) and reduced glutathione (GSH), as well as the lipid peroxidation product (malondialdehyde, MDA), were measured in spinal tissue using commercial kits based on the manufacturer's protocols (Nanjing Jiancheng Bioengineering Institute, Nanjing, China). Bradford method was used to assess protein concentration.

### Assay of myeloperoxidase activity

Myeloperoxidase (MPO) levels in spinal tissue were quantified by using MPO assay kit (Nanjing Jiancheng Bioengineering Institute, Nanjing, China). One unit of MPO activity was defined as the amount of enzyme degrading 1 mmol of peroxidase/min at 25 °C and expressed as Unit per gram (U/g) weight of wet tissue.

# Assay of TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and NF- $\kappa$ B p65 subunit

The concentration of various proinflammatory cytokines such as TNF-α, IL-1β, and IL-6 in spinal homogenate were assessed commercially available ELISA kits (sandwich enzyme immunoassay) based on manufacturer's protocol (Thermo Fisher Scientific Inc, MA, USA). Similarly, the levels of the NF-κB free p65 subunit in the nuclear fraction (CellBiolabs Nuclear/Cytosolic Fractionation Kit) of the spinal tissue was determined by ActivELISA (Imgenex, San Diego, USA) kit.

#### Histopathological examination

Four rats in each group were euthanized and the spinal cord (tissue) were removed, fixed in 10 % buffered formalin for 24 h and embedded in paraffin. Transverse sections were cut at a thickness of 4  $\mu$ m and stained with H & E (100x). Images were captured with a high-resolution digital camera attached to a Nikon Eclipse 80i microscope (Nikon Co., Tokyo, Japan) and analyzed by Image Pro Plus software.

#### Statistical analysis

All values are explicated: mean  $\pm$  standard deviation (SD). Variation between each experimental groups were measured by one way analysis of variance (ANOVA) using the software SPSS 21 version (SPSS Inc., Chicago, USA) and least significant difference (LSD) was determined using post-hoc multi-comparison test. P < 0.05 was deemed statistically significant.

#### RESULTS

#### Effect of RESV on locomotive function

Movement (motor function) of hind limbs were normal in the sham-operated control rats (Table 1). However, the SCIR treated animals showed a significant reduction in hind limb movement, which depicts the loss of several neurons. After supplementation with RESV and MP, hind limb movement was substantially improved, thus lowering the incidence of neurological deficits.

Table 1: Effect of RESV on spinal locomotive function (BBB rating) in sham control and experimental rats

Group	1 <sup>st</sup> day	3 <sup>rα</sup> day	5 <sup>™</sup> day	7 <sup>™</sup> day
Sham (control)	20.00±0.57	20.50±0.87	21.00±0.50	21.00±1.00
SCIR	3.24±0.13 a**	7.42±0.19 a**	10.25±0.25 a**	12.53±0.54 a**
RESV 20+ SCIR	7.02±0.22 b*	11.50±0.59 b*	13.05±0.82 b*	15.64±0.90 b*
RESV 40+ SCIR	8.36±0.31 b*	13.45±0.88b*	15.23±0.78 b*	18.35±0.88b**
RESV 60+ SCIR	8.58±0.39 b*	11.24±0.79 b*	14.56±0.60 b*	16.09±0.98 b**
MP 50+ SCIR	8.16±0.35b*	12.56±0.88 b*	14.78±1.35 b*	17.05±1.08 b**

Values were expressed as mean  $\pm$  S.D for 8 rats in each group; \*\*p < 0.01, \*p < 0.05 (a) SCIR group compared with the sham control group; (b) Treatment groups (RESV 20, 40, 60 and MP 50) compared with the SCIR injured group

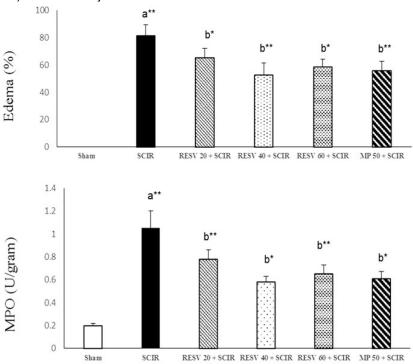
# Effect of RESV on spinal edema

As shown in Fig 1A, spinal E values were dramatically higher (81.35 %) in the SCIR group compared with the sham-operated control group. RESV (20, 40 and 60 mg/kg) groups showed lower spinal E values (65.12, 52.56 and 58.34 %, respectively) than SCII group; MP treatment resulted in an E value of 55.82 %, similar to that of RESV 40 group.

# Effect of antioxidant and lipid peroxidation products (MDA)

The levels of antioxidant and lipid peroxidative products (MDA) in SCIR-injured rats were

determine whether **RESV** examined to administration protected spinal tissue against oxidative stress (Table 2). The spinal SOD, CAT, and GSH activities were is substantially lower (p < 0.05) in the SCIR group than in the shamoperated control group, whereas the MDA levels were markedly elevated (p < 0.05). Treatment with RESV significantly improved (p < 0.05) SOD, CAT and GSH activities as well as concomitantly attenuated the MDA level in spinal tissue. However, the MP group showed a slight improvement in antioxidant activity and partially reverted the MDA levels. RESV 40 (p < 0.01) showed higher antioxidant activity than other RESV doses or MP groups.



**Figure 1:** The effect of RESV on spinal edema (1A) and MPO levels (1B)in sham control and experimental rats.\*\*p < 0.01, \*p < 0.05; (a) SCIR group compared with the sham control group; (b) treatment groups (RESV 20, 40, 60 and MP 50) compared with the SCIR injured group

**Table 2:** Effect of RESV on the activities of spinal antioxidants and lipid peroxidation in sham control and experimental rats.

Group	SOD (U/mg ptn)	CAT (U/mg ptn)	GSH (μg/mg ptn)	MDA (nmols/mg ptn)
Control	3.79±0.45	65.80±6.15	9.03±1.41	0.67±0.08
SCIR	2.21±0.21 a**	54.29±4.78 a**	5.83±0.72 a**	1.44±0.10 a**
RESV 20 + SCIR	3.02±0.18 b*	58.56±5.06 b*	6.98±0.97 b*	0.94±0.07 b*
RESV 40 + SCIR	3.38±0.25 b*	62.99±7.62 b*	8.06±0.81 b**	0.72±0.07 b**
RESV 60 + SCIR	3.14±0.23 b*	60.73±5.83 b*	7.84±0.58 b*	0.89±0.03 b*
MP 50 + SCIR	3.08±0.22 b*	58.74±8.34 b*	7.21±0.62 b*	0.84±0.07 b*

Data are expressed as mean  $\pm$  S.D for 8 rats in each group; \*\*p < 0.01, \*p < 0.05; (a) SCIR group compared with the control group, (b) Treatment groups (RESV 20, 40, 60 and MP 50) compared with the SCIR injured group; ptn = protein. One unit (U) of SOD activity is equal to the amount of enzyme that inhibits the auto-oxidation reaction by 50%; one U of CAT activity was defined as 1 mg of tissue protein that consumed 1  $\mu$ mol of H<sub>2</sub>O<sub>2</sub> at 405 nm for 1 second

# Effect of RESV on neutrophil infiltration (MPO)

To evaluate the effect of RESV on neutrophil activation and infiltration, MPO activity was determined (Fig. 1B). Compared with the SCIR group, the sham-operated group showed a decline (p < 0.05) in MPO activity. Supplementation with RESV and MP markedly attenuated the MPO activity in a dose-dependent fashion.

# Effect of RESV on activities of inflammatory markers

The levels of these inflammatory markers in the spinal tissue of SCIR induced rats were higher (p < 0.01) as compared with sham control rats. Equivalence with SCIR induced rats, RESV and MP treatment (p < 0.01) lowered the concentrations of these pro-inflammatory

cytokines, thus suggesting its anti-inflammatory potential.

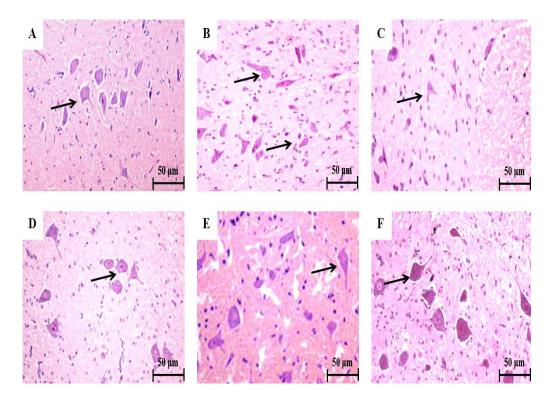
# Effect of RESV on spinal section with hematoxylin and eosin staining

Control showed normal architecture of neuron with a prominent nucleus (Plate 2A). The spinal section of SCIR treated rat portrait a higher degenerated neurons (hemorrhage congestion) with widespread edema, increased cytoplasmic vacuolization and disintegrated nucleus (2B). However, RESV-administered rats (2C - E) showed fewer degenerated neurons (hemorrhage and congestion) compared with SCIR-injured animals, confirming neuroprotective effects of RESV. Similarly, MP treatment (2F) also showed a significant restoration in neuronal structure, but still mild edema accompanied with few degenerated neurons.

Table 3: Effect of RESV on spinal inflammatory markers in sham control and experimental rats

Group	TNF-α (ng/mg protein)	IL-1β (ng/mg protein)	IL-6 (pg/mg protein)	NF-p65 (pg/mg protein)
Sham control	124.56±13.30	68.65±5.78	91.23±9.41	79.47±8.34
SCIR	290.74±19.10 a**	181.52±15.06 a**	215.38±23.02 a**	195.84±20.02 a**
RESV 20 + SCIR	206.24±21.16 b**	122.63±11.10 b*	146.67±14.07 b*	115.75±11.03 b**
RESV 40 + SCIR	144.28±17.33 b**	86.24±10.23 b**	105.11±10.09 b**	91.36±10.73 b**
RESV 60 + SCIR	163.23±20.23 b**	99.45±10.00 b*	116.56±11.07 b*	102.35±9.03 b**
MP 50 + SCIR	151.68±17.34 b**	91.24±9.23 b**	103.46±8.09 b**	99.93±09.82 b**

Values are expressed as mean  $\pm$  S.D (n = 8); \*\*p < 0.01, \*p < 0.05; (a) SCIR group compared with the sham control group, (b) Treatment groups (RESV 20, 40, 60 and MP 50) compared with SCIR-injured group



**Plate 1:** The effect of RESV on the spinal section stained with H & E n in sham control and experimental rats. A transection of control showed normal neuronal architecture (Fig 2A, 100x magnification) (A); transaction of SCIR induced rats showed more r degenerated neurons (hemorrhage and congestion) with widespread edema, increased cytoplasmic vacuolization and disintegrated nucleus (B). Fewer degenerated neurons (hemorrhage and congestion) were observed in the RESV-administered rats (C to E) than in the SCIR-injured animals, confirming the neuroprotective effects of RESV. Similarly, MP treatment (F) also restored neuronal structure significantly, but mild E accompanied by a few degenerated neurons remained. Scale = 50 μm

#### DISCUSSION

Paraplegia is a fatal complication resulting from SCII during thoracic-abdominal aortic aneurysm surgery [1]. Several pathophysiological events, including excitotoxicity (glutamate-mediated), energy failure, inflammation, apoptosis, and oxidative stress, have been implicated in SCIR injury [4,5]. Among these, oxidative stress and inflammation are the major contributors and initiators of SCIR injury [4-6]. Therefore, in the present study, we examined the protective effects of RESV against SCIR injury in a dose-dependent manner by evaluating various oxidative stress and inflammatory markers using MP as a standard.

Assessment of locomotor behavior (hind limb movement) is an important measure of long-term functional recovery after SCIR injury and is a pivotal tool for determining the neuroprotective efficacy of various drugs. Our results clearly show the neuroprotective efficacy of both RESV and MP based on markedly increased BBB scores. This result reflects improved hind limb movement by preventing neuronal loss and

mitigating the incidence of neurological deficits. Liu *et al* also suggested that RESV can significantly lower neurological deficits in a SCIR-injured rabbit model [15].

Morphological abnormalities of spinal tissue are indirectly assessed by evaluating E based on dry and wet weights. During SCIR induction, a substantial reduction in blood flow was observed (due to aortic occlusion), followed by reperfusion (increased blood flow), which alters the electrolytic balance (pumps) and various metabolic processes in spinal tissue, leading to elevated water movement and eventually resulting in E [13,15]. Due to the vasoprotective effects of RESV [16], the electrolytic balance was significantly maintained, thus attenuating E. Our results are in agreement with those of Yang et al, who showed that RESV treatment can revert E successfully in the SCIR-injured rat model [17]. Similarly. Ren et al also inferred that RESV can effectively reduce E under ischemic conditions [18].

Considerable evidence indicates that overproduction of free radicals (oxidative stress)

and the subsequent inflammatory response under ischemic conditions are major factors pathophysiological contributing to various changes during SCIR injury [13,19]. Neural tissues are highly susceptible to free-radical damage due to an increased metabolic rate and lipid content. During SCIR, many free radicals are generated; to counter the excess free radicals the endogenous antioxidant defense system, which includes SOD, CAT and reduced GSH, is upregulated by various factors to trigger the Nrf2-dependent pathway. However, high levels of free radicals suppress normal antioxidant levels and ultimately contribute to oxidative stress. Therefore, SOD, CAT and GSH levels were significantly lower in SCIR-injured

Moreover, levels of the lipid peroxidation product MDA were elevated in the SCIR group. oxidative contributing to stress. RESVsupplemented animals exhibited improved SOD and CAT activities and higher GSH levels, as well as substantially reduced MDA levels, in spinal tissue. Overall, RESV 40 showed better antioxidant activity compared with other RESV dosages or the MP group. Due to the free hydroxyl group, RESV may act as an antioxidant and trigger endogenous antioxidant levels to revert oxidative stress conditions by maintaining a balance between antioxidants and free radicals and activating the Nrf2/HO-1 pathway [20]. Furthermore, RESV has been reported to be a xanthine oxidase inhibitor and halt the synthesis of hypoxanthine and free radicals (ROS) under ischemic conditions [21].

Inflammation is the crucial event that is responsible for the neurological damage (SCI) due to ischemic reperfusion and thus has been proven to be a promising target for a therapeutic approach after SCIR injury [21]. Neutrophil and neuroglial activation, mobilization and infiltration are well-documented key factors during SCIR that initiate the inflammatory cascade by triggering NF-kB and subsequently increasing various pro-inflammatory cytokines such as tumor necrosis factor-α, IL-1β and IL-6 [22]. Hence, the inflammatory status of spinal tissue was indirectly evaluated by measuring MPO activity. Several reports have indicated a strong correlation between MPO levels and the severity of ischemic reperfusion injury [23]. Lafci et al also showed that MPO activity was significantly increased after SCIR injury in a rat model [24]. Similarly, in the present study, MPO activity in spinal cord tissue was significantly increased in response to ischemic reperfusion. The increased MPO activity was markedly attenuated by the RESV and MP treatments, indicating the antioxidant and anti-inflammatory activities of RESV. Renaud and Martinoli argued that treatment with RESV substantially reduces microglial activation and significantly inhibits the infiltration of neutrophils, thus diminishing MPO levels under ischemic conditions [25].

Overexpression of NF-kB has been positively correlated with a higher degree of ischemiainduced neuronal injury [15]. Levels of proinflammatory cytokines such as TNF-α, IL-1β, IL-6, and nuclear factor NF-p65 were dramatically increased in the SCIR rats due to excessive production of free radicals which, in turn, triggered the neuroinflammatory cascade by activating various cytokines. Compared with the SCIR-injured rats, the RESV and MP treatments substantially suppressed the concentrations of these pro-inflammatory cytokines, indicating their anti-inflammatory potential. Our results are supported by those of Smith et al., who evaluated inflammatory chemokine concentrations (interleukin-1 $\dot{\beta}$ , IL-6 and TNF- $\alpha$ ) and reported elevated levels of these inflammatory cytokines after SCIR [26]. Furthermore, RESV can effectively improve cerebral blood flow by crossing the blood-brain barrier and exhibits protective effects against ischemic injury by decreasing glial cell activation and thus positively alleviating oxidative stress and the inflammatory cascade.

RESV can also influence SIRT1 expression indirectly at the transcriptional level, block the translocation of NF- $\kappa$ B (p65) from the cytosol to the nucleus, and inhibit the production of various pro-inflammatory cytokines, especially IL-1 $\beta$ , IL-6 and TNF- $\alpha$ . Moreover, RESV augments endogenous antioxidant activities, thus inhibiting activation of the inflammatory cascade [15].

Sham-operated control rats showed normal neuronal architecture with prominent nuclei. However, SCIR-injured rats had higher numbers of degenerated neurons (hemorrhage and congestion) with widespread E, increased cytoplasmic vacuolization and disintegrated nuclei due to increased oxidative stress and cell damage by lipid peroxidation (MDA). As previously, explained RESV and concomitantly improved the antioxidant status and attenuated MDA levels substantially, indicated by the fact that RESV- and MPadministered rats displayed significantly fewer neurons degenerated (hemorrhage and than SCIR-injured congestion) animals. validating the neuroprotective effects of RESV.

Liu et al [15] also demonstrated that RESV treatment normalized the neuronal count and

function due to antioxidant, anti-inflammatory and anti-apoptotic activities. RESV (40 mg) exhibited the highest neuroprotection compared with MP due to its free radical-quenching activity. Our results are in agreement with the findings of Ates et al and Yang et al [12,17]. The major limitation of the current study is that bioenergetics parameters and apoptosis were not evaluated to confirm secondary energy failure and its association with RESV treatment. Additionally, the reason for the improved activity of RESV (40 mg) should be explored using different molecular techniques.

#### CONCLUSION

Both RESV and MP exhibit neurotherapeutic properties, reflected in various aspects, including substantially reduced E, oxidative stress and inflammation. Furthermore, RESV and MP exhibit spinal protective properties concomitantly reverting histopathological abnormalities and improving the number of neurons. Thus, it is proposed here that RESV (40 mg) can be used as a neurotherapeutic agent to treat SCIR injury and lower the risk of paraplegia significantly.

#### **ACKNOWLEDGEMENT**

Financial support provided by the Chest Hospital of Henan Province, Zhengzhou, Henan 450008, PR China is appreciated.

#### **DECLARATIONS**

### Conflict of Interest

No conflict of interest associated with this work.

#### **Contribution of Authors**

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

#### **REFERENCES**

- Zhu JW, Chen T, Guan J, Liu WB, Liu J. Neuroprotective effects of allicin on spinal cord ischemia-reperfusion injury via improvement of mitochondrial function in rabbits. Neurochem Int 2012; 61(5): 640-648.
- Bicknell CD, Riga CV, Wolfe JH. Prevention of paraplegia during thoracoabdominal aortic aneurysm repair. Eur J Vasc Endovasc Surg 2009; 37: 654-660.
- Toumpoulis IK, Papakostas JC, Matsagas MI, Malamou-Mitsi VD, Pappa LS, Drossos GE, De Rose JJ,

- Anagnostopoulos CE. Superiority of early relative to late ischemic preconditioning in spinal cord protection after descending thoracic aortic occlusion. J Thorac Cardiovasc Surg 2004; 128(5): 724-730.
- Kaplan S, Bisleri G, Morgan JA, Cheema FH, Oz MC. Resveratrol, a Natural Red Wine Polyphenol, Reduces Ischemia-Reperfusion—Induced Spinal Cord Injury. Ann Thorac Surg 2005; 80(6): 2242-2249.
- Fu J, Fan HB, Guo Z, Wang Z, Li XD, Li J, Pei GX. Salvianolic acid B attenuates spinal cord ischemiareperfusion-induced neuronal injury and oxidative stress by activating the extracellular signal-regulated kinase pathway in rats. J Surg Res. 2014; 188(1): 222-230.
- Pettiford JN, Bikhchandani J, Ostlie DJ, Peter SD, Sharp RJ, Juang D. A review: the role of high dose methylprednisolone in spinal cord trauma in children. Ped Surg Int. 2012; 28(3): 287-294.
- Liu BL, Zhang X, Zhang W, Zhen HN. New enlightenment of French Paradox: resveratrol's potential for cancer chemoprevention and anti-cancer therapy. Cancer Biol Ther. 2007; 12: 1833-1836.
- Yousuf S, Atif F, Ahmad M, Hoda N, Ishrat T, Khan B, Islam F. Resveratrol exerts its neuroprotective effect by modulating mitochondrial dysfunctions and associated cell death during cerebral ischemia. Brain Res. 2009; 1250: 242-253.
- Karalis F, Soubasi V, Georgiou T, Nakas CT, Simeonidou C, Guiba-Tziampiri O, Spandou E. Resveratrol ameliorates hypoxia/ischemia-induced behavioral deficits and brain injury in the neonatal rat brain. Brain Res. 2011; 1425: 98-110.
- Meng Z, Li J, Zhao H, Liu H, Zhang G, Wang L, Hu H, Li D, Liu M, Bi F, Wang X, Tian G, Liu Q and Buren B. Resveratrol relieves ischemia induced oxidative stress in the hippocampus by activating SIRT1. Exp Ther Med.2015; 10(2): 525-530.
- 11. Kodali M, Parihar VK, Hattiangady B, Mishra V, Shuai B, and Shetty AK. Resveratrol prevents age-related memory and mood dysfunction with increased hippocampal neurogenesis and microvasculature, and reduced glial activation. Sci Rep. 2015; 8075: 5.
- Ates O, Cayli S, Altinoz E, Gurses I, Yucel N, Kocak A, Yologlu S, Turkoz Y. Effects of resveratrol and methylprednisolone on biochemical, neurobehavioral and histopathological recovery after experimental spinal cord injury. Acta pharma Sinic. 2006; 27(10): 1317-1325.
- Hwang JY, Min SW, Jeon YT, Hwang JW, Park SH, Kim JH, Han SH. Effect of coenzyme Q10 on spinal cord ischemia-reperfusion injury. J Neurosurg: Spine. 2015; 22(4): 432-438.
- Kaplan S, Bisleri G, Morgan JA, Cheema FH, Oz MC. Resveratrol, a Natural Red Wine Polyphenol, Reduces Ischemia-Reperfusion-Induced Spinal Cord Injury. Ann Thorac Surg. 2005; 80(6): 2242-2249.
- 15. Liu C, Shi Z, Fan L, Zhang C, Wang K, Wang B. Resveratrol improves neuron protection and functional

- recovery in rat model of spinal cord injury. Brain Res. 2011; 1374: 100-109.
- Mdzinarishvili A, Kiewert C, Kumar V, Hillert M, Klein J. Bilobalide prevents ischemia-induced edema formation in vitro and in vivo. Neurosci. 2007; 144: 217-222.
- Yang YB, Piao YJ. Effects of resveratrol on secondary damages after acute spinal cord injury in rats. Acta Pharma Sinic. 2003; 24(7): 703-710.
- Ren J, Fan C, Chen N, Huang J and Yang Q. Resveratrol pretreatment attenuates cerebral ischemic injury by upregulating expression of transcription factor Nrf2 and HO-1 in rats. Neurochem Res. 2011; 36: 2352-2362.
- Guven M, Akman T, Yener AU, Sehitoglu MH, Yuksel Y, Cosar M. The Neuroprotective Effect of Kefir on Spinal Cord Ischemia/Reperfusion Injury in Rats. J Kor Neurosurg Soc. 2015; 57(5): 335-341
- Das S, Fraga CG and Das DK. Cardioprotective effect of resveratrol via HO-1 expression involves p38 map kinase and PI-3-kinase signaling, but does not involve NFkappaB. Free Radic Res. 2006; 40: 1066-1075.
- Ege E, Ilhan A, Gurel A, Akyol O, Ozen S. Erdosteine ameliorates neurological outcome and oxidative stress due to ischemia/reperfusion injury in rabbit spinal cord. Eur J Vasc Endovasc Surg. 2004; 28: 379-386.

- 22. Pallas M, Porquet D, Vicente A and Sanfeliu C. Resveratrol: new avenues for a natural compound in neuroprotection. Curr Pharm Des. 2013; 38: 6726-6731.
- 23. Simao F, Matte A, Matte C, Soares FM, Wyse AT, Netto CA, Salbego CG. Resveratrol prevents oxidative stress and inhibition of Na(+)K(+)-ATPase activity induced by transient global cerebral ischemia in rats. J Nutr Biochem. 2011; 10: 921-928.
- 24. Lafci G, Gedik HS, Korkmaz K, Erdem H, Cicek OF, Nacar OA, Yildirim L, Kaya E, Ankarali H. Efficacy of iloprost and montelukast combination on spinal cord ischemia/reperfusion injury in a rat model. J Cardiothorac Surg 2013; 8(1): 64
- Zhu P, Li J, Fujino M, Zhuang J, Li XK. Development and treatments of Inflammatory cells and cytokines in spinal cord ischemia-reperfusion injury. Mediators Inflamm. 2013; 2013: 1-7.
- Smith PD, Puskas F, Meng X, Lee JH, Cleveland JC (Jr), Weyant MJ, Fullerton DA, Reece TB. The evolution of chemokine release supports a bimodal mechanism of spinal cord ischemia and reperfusion injury. Circ. 2012; 126: 110-117.