

## Original Research Article

# Development of a high performance liquid chromatography method for simultaneous analysis of theophylline, guaifenesin and diphenhydramine in an elixir

Hayun\*, Yahdiana Harahap and Maria O Puspasari

Faculty of Pharmacy, Universitas Indonesia, Depok, 16424, West Java, Indonesia

\*For correspondence: **Email:** hayun.ms06@gmail.com; **Tel:** +62217270031; **Fax:** +62217863433

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### Abstract

**Purpose:** To develop and validate a new low-cost high performance liquid chromatography (HPLC) method for simultaneous analysis of theophylline (TH), guaifenesin (GF) and diphenhydramine hydrochloride (DH) in elixir dosage form.

**Methods:** Chromatographic conditions were an isocratic elution with C18-Kromasil® column (250 x 4.6 mm, 5 µm), methanol-water (1:1, v/v, pH 3.0) as mobile phase, flow rate 1.0 ml/min and UV detector at λ 218 nm. The method was validated for selectivity, linearity, LOD-LOQ, precision, and accuracy.

**Results:** Retention time of TH, GF and DH was 3.3, 5.3 and 9.1 min, respectively. The method showed good selectivity, calibration curves were linear over the concentration range of 1.000 – 10.002 µg/mL, 0.801 – 8.008 µg/mL, and 0.251 – 2.514 µg/mL ( $r^2 > 0.999$ ). LOD was 0.1093, 0.16520, and 0.0706 µg/mL, while LOQ was 0.3645, 0.5506, and 0.2354 µg/mL for TH, GF and DH, respectively. Recovery accuracy was 99.77 - 101.10, 100.50 - 101.95 and 99.20 - 100.13 % for TH, GF and DH, respectively; precision (RSD) was < 2.0.

**Conclusion:** The proposed method is highly selective, sensitive, precise, and accurate, and would suitable for the simultaneous analysis of TH, GF, and DH in elixir dosage form. Since methanol is cheaper than acetonitrile, the application of the method may reduce the cost of analysis.

**Keywords:** Simultaneous analysis, Theophylline, Guaifenesin, Diphenhydramine hydrochloride, Elixir

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## INTRODUCTION

Theophylline (TH), guaifenesin (GF) and diphenhydramine hydrochloride (DH) (Figure 1) is a combination of the active ingredients contained in elixir dosage form. This combination is used for a treatment of bronchial asthma and asthmatic bronchitis conditions [1,2]. The reversed phase high performance liquid chromatographic (HPLC) method has been widely used for the assay of one or two active ingredients simultaneously in dosages form containing TH, GF and or DH, or other active ingredients [3-8]. The HPLC methods have also

been used for the simultaneous analysis of these three active ingredients in a mixture with some other active substances or excipients using acetonitrile-phosphate buffer pH 3.2 as mobile phase [9,10].

Acetonitrile is an excellent solvent for mobile phase of reversed phase HPLC. However, it is relatively expensive solvent and sometimes difficult to obtain. Therefore it is necessary to develop an alternative HPLC method using another solvent as mobile phase. Methanol is the most suitable solvent replacement for acetonitrile. The solvent is cheaper, easier to

obtain and more polar than acetonitrile, reducing the risk of solid buffer precipitation. However, methanol has a weaker elution strength and a higher back pressure [11,12]. The objective of the present study was the development and validation of a new reversed-phase HPLC method for simultaneous analysis of TH, GF and DH in elixir dosage form, using a mixture of methanol-water as mobile phase.

## EXPERIMENTAL

### Materials and reagents

Reference standard of TH, GF and DH were obtained from the National Agency of Drug and Food Control (NA-DFC) of Republic of Indonesia. Methanol used was HPLC grade (Merck), phosphoric acid was of analytical grade (Merck), and double distilled water (Otsuka) and elixir sample containing TH, GF and DH (Tusapress<sup>®</sup>) were purchased from commercial source.

### Preparation of standard and sample solution

A stock solution of analytes was prepared by dissolving 50.0 mg TH, 40.0 mg GF, and 12.5 mg DH accurately weighed, in double distilled water up to 100.0 mL, series of dilution were performed with selected mobile phase to give working standard for individual and mixture of analytes with the concentrations of 5.0, 4.0 and 1.25 µg/mL, respectively. A sample solution was prepared by dissolving a certain amount of weight equivalent to approximately 5.0 mL of the sample in mobile phase to give 100.0 mL of solution, then performed filtration and a series of dilution to give sample solution with the concentration of 5.0, 4.0 and 1.25 µg/mL calculated of the labeled amount of TH, GF, and DH, respectively.

### HPLC conditions

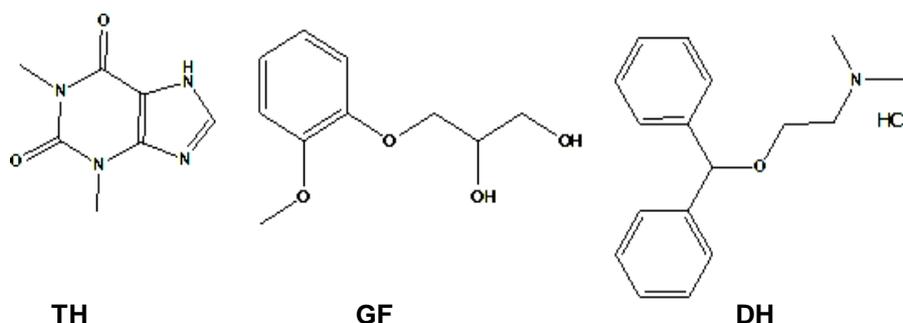
High performance liquid chromatography (HPLC) equipped with Kromasil<sup>®</sup>-C18 column (250 x 4.6

mm, 5 m), pump (Shimadzu LC-10AD), UV-Vis detector (Shimadzu SPD-10A), and syringe 20.0 µL (Hamilton Co. Nevada) were used in this study. The detector was set at 218 nm. The mobile phase of the various composition was prepared as mixture of water and methanol (4:1, 3:2 and 1:1 v/v), then adjusted by addition of phosphoric acid to a pH of 3.0, filtered using 0.45 µm membrane filter and degassed by sonication for 15 min. The flow rate was 1.0 ml/min isocratically. The retention time (tR), and capacity factor (k'), HETP, the number of theoretical plates (N), tailing factor (Tf), and separation (R) of the chromatogram peaks resulting from the injections of the mixture of three active ingredients solution into the HPLC were calculated. One of the compositions of the mobile phase which provided the best characteristics for separation was selected. The replicate injections into HPLC with selected condition were also performed to evaluate the precision of the instrument.

### Method validation

The analytical method validation was performed following International Conference on Harmonization (ICH) Q2 guidelines (2005) [13], covering the selectivity, linearity, limit of detection (LOD), limit of quantitation (LOQ), precision and accuracy. *The selectivity* was evaluated by injecting the blank sample solution, the sample solution, and the standard solutions into the chromatograph. The presence of interferences on the chromatogram peaks of analytes were observed [13].

The linearity of the proposed methods was evaluated by determining the correlation coefficient ( $r^2$ ) value of linear regression analysis ( $Y = a + bX$ ) of calibration curves of the analytes [13], in the concentrations range of 1.0 - 10.0 µg/mL for TH, 0.8 - 8.0 µg/mL for GF, and 0.25 - 2.5 µg/mL for DH, respectively.



**Figure 1:** Chemical structures of theophylline (TH), guaifenesin (GF), and diphenhydramine hydrochloride (DH)

The LOD and the LOQ were determined using data of standard deviation of the response ( $\sigma$ ) and the slope of the calibration curve (S).  $LOD = 3.3 \sigma/S$  and  $LOQ = 10 \sigma/S$ , respectively.

Precision was evaluated by performing repeatability and intermediate precision test. Repeatability (intra-day) was assessed by analyzing the analytes in the sample solutions at 100 % levels calculated from the concentration of the analytes in the label of the sample in 6 replicate. Intermediate precision was assessed by determining the analytes in the sample on three consecutive days in triplicate, respectively. The values of relative standard deviation (RSD) for the repeatability and the intermediate precision were reported [13].

The accuracy was tested by determining recovery values of the analytes in the sample addition [13]. Accurately weighed standard TH, GF and DH were spiked to the sample, diluted with mobile phase solvent to give three different concentration levels (80, 100, and 120 % of the labeled amounts of each analyte in the sample), and analyzed by the proposed methods in triplicate.

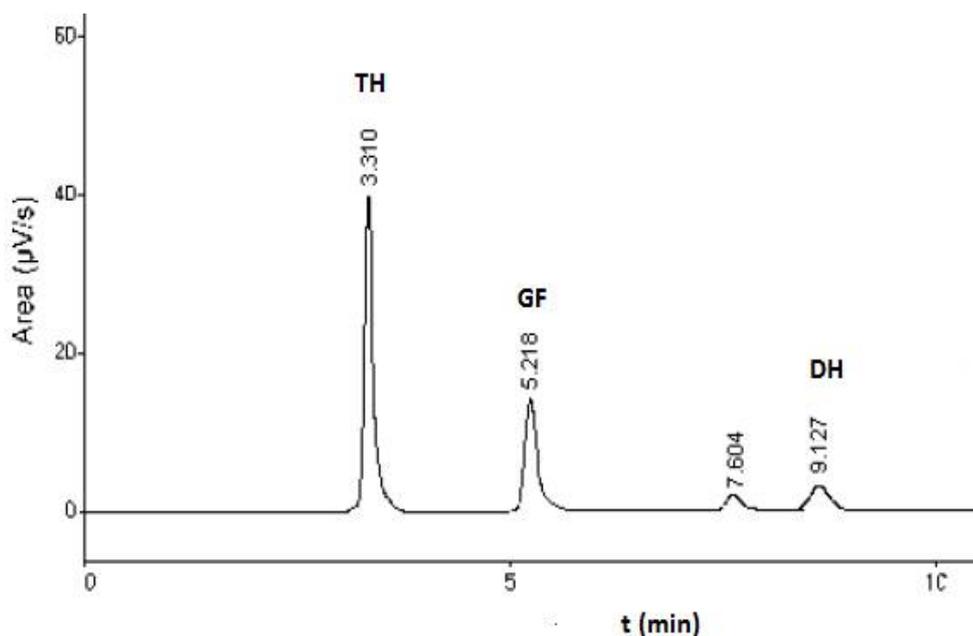
## RESULTS

Initial method development was the selection of mobile phase compositions. UV detector was set at  $\lambda$  218 nm. The wavelength is maximum  $\lambda$  of diphenhydramine HCl, which is the component

with the smallest concentration and absorption. In the  $\lambda$ , the three analytes are best detected. The mixture of methanol and water (1:1, v/v, pH 3.0) which provided the best chromatographic characteristics was selected as a mobile phase. Under this condition, retention time (tR) of TH, GF and DH were 3.3, 5.2, and 9.1 min, respectively (Figure 2). Five replicate injection consecutively of working standard solution gave RSD not more than 2.0 % for all analytes. The data indicates a good precision of the chromatographic systems.

The method showed a good selectivity. There was no observed interference from the sample components and others at Rf of TH, GF and DH. The calibration curves between the concentration of TH, GF and DH and their peaks area showed good linear relationship over the concentration range of 1.000 – 10.002  $\mu\text{g/mL}$ , 0.801 – 8.008  $\mu\text{g/mL}$ , and 0.251 – 2.514  $\mu\text{g/mL}$ , for TH, GF and DH, respectively ( $r^2 = 0.999$ ). The LOD values obtained were 0.1093, 0.16520, and 0.0706  $\mu\text{g/mL}$ , while the LOQ were 0.3645, 0.5506, and 0.2354  $\mu\text{g/mL}$  for TH, GF and DH, respectively (Table 1).

The precision and accuracy of the method were satisfactory. The RSD values of repeatability and intermediate precision obtained were all less than 2.0 % (Table 2), and the mean of recovery values obtained were 99.77 - 101.10 %, 100.50 - 101.95 % and 99.20 - 100.13 % for TH, GF and DH, respectively (Table 3).



**Figure 2:** HPLC chromatogram of 20  $\mu\text{L}$  injection of sample elixir solution containing theophylline (TH), guaifenesin (GF) and diphenhydramine hydrochloride (DH).

**Table 1:** Characteristics of the calibration curves, the limit of detection (LOD) and the limit of quantitation (LOQ) of HPLC method for the simultaneous analysis of theophylline (TH), guaifenesin (GF) and diphenhydramine hydrochloride (DH)

Parameter	TH	GF	DH
Range ( $\mu\text{g/mL}$ )	1.000–10.002	0.801–8.008	0.251–2.514
Regression Eq. (Y) <sup>1)</sup> :			
Intercept (a)	314.15	- 650.75	-1385.30
Slope (S) = (b)	53385	32871	36765
Standard dev of intercept ( $\sigma$ )	1946.001	1811.783	865.470
LOD ( $\mu\text{g/mL}$ )	0.1203	0.1819	0.0776
LOQ ( $\mu\text{g/mL}$ )	0.3645	0.5511	0.2354
Correlation coefficient (r)	0.999	0.999	0.999

<sup>1)</sup>Regression Eq.:  $Y = a + bX$ ; where  $Y = \text{peak area}$ ;  $X = \text{concentration } (\mu\text{g/mL})$

**Table 2:** Precision of the method

Analyte	Level (%)	Conc <sup>1)</sup> ( $\mu\text{g/mL}^1$ )	Repeatability (n=6) <sup>2)</sup>		Intermediate precision (n=9) <sup>2)</sup>	
			Mean conc ( $\mu\text{g/mL}$ )	RSD (%)	Mean conc ( $\mu\text{g/mL}$ )	RSD (%)
TH	100	5.283	5.324	0.55	5.303	0.93
GF	100	4.226	4.262	0.29	4.248	0.55
DH	100	1.321	1.325	0.76	1.317	1.05

<sup>1)</sup> Calculated from weight of the sample used, concentration of the analytes in the label and dilution; <sup>2)</sup> n = number of repetitive determinations.

**Table 3:** Accuracy of the method

Analyte	Level (%)	Analyte conc ( $\mu\text{g/mL}$ )		Results of analysis (n=3)		Mean spiked recovery ( $\mu\text{g/mL}$ ) (e-d)	Percent (%) recovery [(f/c)x100]
		Spiked	Sample <sup>1)</sup>	Mean total conc ( $\mu\text{g/mL}$ )			
a	b	c	d	e	f	g	
TH	80	2.005	2.008	4.035	2.027	101.10	
	100	2.503	2.510	5.012	2.502	99.96	
	120	3.008	3.012	6.013	3.001	99.77	
GF	80	1.601	1.608	3.223	1.615	100.87	
	100	2.001	2.010	4.050	2.040	101.95	
	120	2.401	2.412	4.825	2.413	100.50	
DH	80	0.502	0.499	0.997	0.498	99.20	
	100	0.626	0.623	1.245	0.622	99.36	
	120	0.751	0.748	1.500	0.752	100.13	

<sup>1)</sup> Calculated from weight of the sample used, the concentration of the analytes resulted from repeatability study and dilution

## DISCUSSION

Theophylline (TH), guaifenesin (GF) and diphenhydramine hydrochloride (DH) are a combination of the active ingredients in elixir dosage form for oral administration [1]. The UV absorption spectra of TH, GF and DH display considerable overlap. Moreover, oral solution generally contains flavouring, sweetening or coloring agent [3]. Several papers have reported the use of reverse phase HPLC methods for simultaneous analysis of these three active ingredients in a mixture with some other active substances or excipients [9,10]. However, due to cost or solvent availability, it is necessary to develop an alternative method using another solvent as a mobile phase.

The price of methanol is about 1/4 times that of acetonitrile [14]. In addition, methanol is easier to obtain and more polar than acetonitrile, reducing the risk of solid buffer precipitation [11]. Hence the proposed method would be less costly to perform routine determination of TH, GF and DH in elixir dosage form in the pharmaceutical industry.

The results of the method validation showed that all parameters are within acceptable limits. The presence of excipients in the formulations did not cause any interference with TH, GF, and DH peaks. Thus the method is selective for simultaneous determination of TH, GF, and DH. Good linearity was observed with either

concentration range exceeds the concentrations range employed in the test substance with a regression coefficient ( $r^2$ ) = 0.999, and the LOD-LOQ obtained indicating a high degree of sensitivity.

The RSD of intra-day measurements of TH, GF and DH were 0.55, 0.29 and 0.76 %, respectively, while the RSD of intermediate precision of TH, GF and DH were 0.93, 0.55 and 1.05 %. The RSD values obtained were less than RSD max, thus the method is precise. The recovery values obtained were 99.77 - 101.10 %, 100.50 - 101.95 % and 99.20 - 100.13 % for TH, GF and DH, respectively. Thus the method is accurate. Overall, the data indicate that the method is suitable as an alternative for the determination of TH, GF and DH in elixir dosage form simultaneously.

The proposed method consumed about 5 ml of methanol to run once analysis, while the previous method using acetonitrile-phosphate buffer [9] consumed about 3 ml of acetonitrile. Taking into account that methanol is cheaper than acetonitrile, this result could be an inspiration for the development of lower-cost HPLC methods for the determination of TH and/or GF in other dosage forms for asthma medications.

## CONCLUSION

An HPLC method for the simultaneous analysis of TH, GF and DH has been developed and validated. The method is selective, sensitive, precise and accurate, so is a suitable alternative for the simultaneous analysis of TH, GF and DH in elixir dosage form. Methanol is cheaper than acetonitrile, hence the use of methanol may reduce the cost of the analysis.

## DECLARATIONS

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### Conflict of Interest

No conflict of interest associated with this work.

### Contribution of Authors

The authors declare that this work was done by the authors named in this article and all liabilities

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