Prevalence of Leptospira interrogans in free range domestic duck (Cairina moschata) from selected areas of Morogoro Municipality, Tanzania

G.G. Kaiki and A. E. Pereka

Department of Physiology Toxicology and Pharmacology, Sokoine University of Agriculture, P.O. Box 3017, Morogogoro Tanzania. Email: george.georgina16@yahoo.com

SUMMARY

Leptospirosis is described as the most common and universal zoonotic bacterial disease around the world caused by Leptospira interrogans which affects different species of domestic and wild animals. The bacteria may occurs worldwide especially in subtropical, tropical, and wet environment with slightly alkaline soil. The current study was conducted between November 2016 and July 2017 in the selected areas in Morogoro aimed at estimating the prevalence and establishes the common serovars of *Leptospira* in free range ducks. A total of 30 ducks from 12 households were used. Before blood sample collection, the duck biodata was recorded and the owner was asked on the general management system including the scavenging areas and any veterinary intervention in place. The ducks were restrained and blood samples were collected from the branchial vein and left to clot before harvesting the serum. Multiple Agglutination Tests was performed using four serovars namely Kenya, Grippotyphosa, Pomona and Hebdomadis as reference serovars. The results indicated that all the ducks were local breeds (Cairina moschata) which scavenged for fed around homestead. The ducks accessed dumping areas; stagnant water, animal houses and the mud around. No veterinary intervention to ducks was reported by all 12 respondents interviewed. Laboratory results indicated that 5 samples (16.7%) were positive at 1:20, 1:40 for Hebdomadis and Kenya serovars which implies that the ducks had been exposed to the specific Leptospira serovars. This study reports for the first time the seroprevelence of leptosirosis in ducks in Tanzania. Because of the zoonotic nature of Leptospira and sharing of common environment between ducks and humans, the diseases can easily get access to human. Therefore, in the efforts of surveillance and control of leptosisis in humans and animals, ducks should also be involved.

Key words: Leptospirosis, Leptospira interrogans, MAT, serovars, ducks

INTRODUCTION

Leptospirosis is the most common and universal zoonotic bacterial disease around the world caused by pathogenic Leptospira called Leptospira interrogans (Abela- Ridder et al, 2010). In Tanzania Leptospira infection has been reported in many different animal species including domestic animals such as cattle, sheep, pigs, horses and dogs (Machang'u et al, 1997; Mgode et al., 2006). Leptospirosis is transmitted from animal to human through direct and indirect contact with urine, abortion products and other material contaminated with fluids from infected animals (Bharti et al., 2003). A wide variety of wild animal hosts including rodents, bats, possums, deer, mongoose and small insectivores also have been reported to habour the infection (Bharti et al., 2003; Ellis, 2010). Ducks may come into contact with the Leptospira in infected stagnant water and in mud while swimming, passing through contaminated water, drinking contaminated water and coming into contact with urine from an infected animal (Mwachui et al., 2015) and outbreaks of disease mainly occur after heavy seasonal rainfall (Lau et al., 2010).

Leptospira are aerobic, Gram-negative spirochetes, with periplasmic flagella, resembling a question mark when viewed through a light microscope and are slow growing. Traditionally, Leptospira were divided into two groups; the pathogenic Leptospira were all classified as members of Leptospira interrogans, and the saprophytic Leptospira were classified as Leptospira biflexa, (Mohammed et al, 2011; Mety and Dikken, 1993). The first step in the pathogenesis of leptospirosis is penetration of tissue barriers to gain entrance to the body. Potential portals of entry include the skin via a cut, genital tract, and the mucous membranes of the conjunctivae or oral cavity swallowing while swimming in contaminated water (Corwin et a.l, 1990; Lingappa et al., 2004; Stern et al., 2010). Thereafter, there is hematogenous dissemination: Pathogenic leptospires make their way into the bloodstream and persist there during the leptospiremic phase of the illness (Stern et al., 2010) there after the spirochetes multiply in the organs mostly the central nervous system, kidney and liver. The symptoms of leptospirosis develop around 7 to 14 days after exposure to *Leptospira* with mild clinical signs namely; chills, high temperature, sudden headache, nausea, vomiting and loss of appetite, muscle pain and conjunctivitis (Beran *et al.*, 1994).

Diagnosis of leptospirosis in human is difficult based on clinical signs, such that may be misdiagnosed and mismanaged as being mistaken for malaria with similar clinical presentation that might contribute to a high rate of infection in the study area. Methods used in detection of leptospires depend on the availability of resources. Detection of *Leptopisra* infection can be done by the use of Microscopic Agglutination Test (MAT) where dark field microscope is used to detect the agglutination. Other methods are culture, Enzyme Linked Immunosorbent Assay (ELISA), staining method and the Polymerase Chain Reaction (PCR).

Morogoro is among the regions with many livestock in Tanzania. The region is also bordered with several wildlife conservations areas which give lot interactions between humans and animals especially in the interface areas. This gives some possibilities for transfer of different diseases causing agents from wildlife to the domestic environment where domestic animals and humans can easily be infected. The average annual rainfall varies between 600 mm and 1800 mm. The eastern part of Uluguru mountain receive high rainfall of about 2850 mm annually, the leeward side of the mountain are generally dry and receive rainfall of less than 600 mm per year (Msanya et al., 2001). The nature of the soil in the valleys of Morogoro is nearly neutral to alkaline (Msanya et al., 2001). Residents of Morogoro Municipality also keep a number of animals including ducks which scavenge all over the home environment in search for food. Studies on Leptospira infection in animals has been done in other animals (Machang`u et al, 1997; Mgode et al., 2006) but no any study in domestic birds. The purpose of this study was to estimate the prevalence and establish the common serovars of Leptospira infection in free range ducks reared in Morogoro Municipality, Tanzania.

MATERIALS AND METHOD

Study area and population

The study was conducted in Morogoro Municipality which is 190 km west of Dar es Salaam. The study flocks were from different suburbs namely Magadu, Falkland, Kididimo and Vibandani. Selection of study site was based on the convenience of accessibility from Sokoine University of Agriculture (SUA) laboratory and generally represented backyard farming of ducks in urban areas in Tanzania. The study population was Muscovy ducks (*Carina moschata*) managed in the backyard and allowed to scavenge freely during the day within the homestead.

Research design and sampling

The purposive sampling method was used during this cross sectional study design. Sampling involved only farmers who willingly agreed to participate in the study. A total of 12 backyard duck flocks were involved in this study. Before sampling, information on the duck biodata, general duck management system including the scavenging areas and any veterinary intervention in place were gathered from the owners. The ducks selected for sampling were manually restrained and 2 ml of blood sample was collected from the brachial vein on the inside of either wings using syringes and needles. The blood samples in the syringes were left at room temperature for one night to clot. The second day serum was harvested into cryovials and stored until analysis.

Laboratory diagnosis of the Leptospira interrogans

Microscopic Agglutination Test (MAT) which is considered as a gold standard method for leptospirosis serodiagnosis was carried out. This test was conducted at the Pest Management Center, SUA, and Morogoro, Tanzania. The MAT test was performed using standard laboratory procedure that aimed at detecting the antibodies reaction with the respective local antigens. Live leptospira including 4 serovars representing Kenva, Grippotyphosa local isolate from domestic animals and rodents found in Tanzania, were used. Other serovars that were used as references were Leptospira serovar Hebdomadis and Pomona. These serovars were cultured into fresh Leptospira EMJH (Ellinghausen and McCullough, modified by Johnson and Harris) culture medium incubated at 30 ^oC for 4 to 10 days before using as live antigen in MAT. Antigen density of 300×10^8 leptospires/ml was used for MAT.

Serum samples were diluted to 1:10 - 1:80 with phosphate buffered saline (pH 7.2) in U-bottomed microtiter plate. Live leptospires antigen (50 µl) were added to diluted serum to give final dilutions of 1:20-1:160. The plates with serum-antigen mixture were incubated at 30 °C for 2–4 hours

before examining for agglutination of leptospires and antibodies under dark field microscope.

RESULTS

General results

A total of 30 adult ducks from 12 households in four streets were sampled for analysis of *Leptospira* infection (Table 1). The ducks appeared apparently healthy. Interviews with owners indicated that all the ducks were local breeds (*Cairina moschata*). There was no formal feeding, rather ducks are left to scavenge for food and in rare occasions they were given kitchen leftovers. The ducks were scavenged for fed around homestead and accessed dumping areas; stagnant water, animal houses and any kind of muddy environment. No veterinary intervention to ducks was reported by all 12 respondents interviewed.

Serological results

The Microscopic Agglutination Test results are shown in Tables 1 and 2. The seropreplevelence of leptospirosis in ducks was 16.7%. Five sera samples

Table 2. Seroprevelence of *Leptospira* infection in ducks

Title	Number (%) of positive sera to different serovars				
	Hebdomadis	Kenya	Pomona	Grippotyphosa	Total number (%)
1:20	2 (6.7)	1 (3.3)	0 (0.0)	0 (0.0)	3 (10.0)
1:40	2 (6.7)	0 (0.0)	0 (0.0)	0 (0.0)	2 (6.7)
Total	4 (13.3)	1 (3.3)	0 (0.0)	0 (0.0)	5 (16.7)

DISCUSSION

This study was carried out to determine the prevalence of leptospirosis in ducks from Morogoro Municipality. The overall prevalence of leptospirosis in ducks was 16.7%. This shows that a relatively high number of ducks were infected with Leptospira in the study areas. There has been a belief that leptospirosis is a disease of mammals only but this study shows that birds are also infected. It is still not clear as to whether the ducks were sick from leptospirosis of were just carriers of the infection since all the screened ducks were apparently healthy. Nevertheless, Beran et al. (1994) reported that birds like ducks do not develop clinical leptospirosis when are infected with Leptospira but rather develop antileptospiral antibodies. Everand et al. (1985) reported a seroprevalence of 11% in chickens. This suggests that it is true that poultry are susceptible to leptospira infections. Whatever the case, ducks in Tanzania are always left to scavenge for food ducks were reactive at 1:20, 1:40 for Hebdomadis and Kenya serovars which implies that the ducks had been exposed to the specific *Leptospira* serovars.

Table 1. Number of ducks from each street

Street	Number of households visited	Number of ducks	Number (%) of positive ducks
Folkland	4	6	2 (6.7)
Kididimo	3	10	3 (10.0)
Vibandani	3	8	0 (0.0)
Magadu	2	6	0 (0.0)

around homestead areas and shed their faecal droppings all over and if infected, *Leptospira* can find their way into the food chain and affect humans. This study showed for the first time the role of ducks as reservoir hosts of *Leptospira* in Morogoro Municipality.

It was further found that 10% the agglutination was observed at the titre of 1:20 and 6.7% the agglutination observed at the titre of 1:40. The titres were extended up to 1:160 but the agglutination was not observed implying that the observed positive results indicated acute infection. The ducks had been recently infected or exposed to *Leptosira* and this shows that the pathogen exists either in the soil, other animals around or in humans. In this case, the ducks can be used as bioindicators of existence of *Leptospira* in the locality.

Of the four serovars tested, Hebdomadis and Kenya reacted positive to some samples. This is the

indication that ducks are more likely to be infected by Herbidomadis and Grippotyphosa serovas. Previous studies in Tanzania have reported the two serovars in humans, fish, domestic and wild animals (Machang`u et al., 1997; Mgode et al., 2006). Since the two serovars are already in the surroundings, different animal species will be exposed as has been with the case of ducks. The emergence and endemicity of *Leptospira* in Morogoro Municipality may have resulted from the regular high seasonal rain to the area and due to an increase of pastoral population over the recent years (Machang'u et al., 1997; Mgode et al., 2006). The nature of the soil in the valleys of Morogoro is nearly neutral to alkaline (Msanya et al., 2001) which give optimal condition for the survival of the Leptospira in the environment.

The questionnaire study further supports the laboratory observation on seropositivity of ducks on Leptospira. It was observed that ducks scavenge around areas of homestead and also easily access to dumping areas where every kind of rubbish is thrown including domestic animal manure. Ducks were exposed to muddy environment and wetland areas which may potentially serve as sources of Leptospira infections in these birds. Therefore, the intensive management system of ducks can help to minimize the unnecessary exposure of ducks to the contaminated areas which will make them to be safe but also for the affected birds, the chances of contaminating the environment becomes minimal.

It is concluded that ducks have been observed to be seropositive of Leptospira infection in Morogoro municipality which further give evidences of existence of this zoonotic pathogen. Efforts should be put in place to confine the ducks so that to minimize environmental shedding of the pathogens and also to protect the birds from *Leptospira* infection.

ACKNOWLEDGEMENTS

The authors acknowledge the financial support from the Higher Education Student Loan Board HESLB. Sincere thanks are extended to the farmers for accepting to use their ducks for this study samples. Also grateful to laboratory technicians at SUA Pest Management Center for advice and helping in analysis of duck sera samples.

REFERENCES

- Abela-ridder, B., Sikkema, R. and Hartskeerl, R. *Estimating* the burden of human leptospirosis. Intern J Antim Agents, 36, S5–S7, 2010.
- Balthazar M. Msanya, B.M., Kimaro, D.N., Kimbi, G.G., Kileo, E.P. and Mbogoni, J.J.D.J. Land resources inventory and suitability assessment for the major land use types in Morogoro Urban District, Tanzania, Vol 4. Department of Soil Science, Faculty of Agriculture, Sokoine University of Agriculture, Morogoro, Tanzania, 78PP, 2001.
- Beran, G.W., Steele, J.H., Benenson, A.S., Torten, M., Dresen, D.W., Ristic, M. and Pier, A.C. Handbook of zoonoses. Second edition. Section A: Bacterial, Rickettsial, Chlamydial and Mycotic. CRS Press, Washington, D.C., 544PP, 1994.
- Bharti AR, Nally JE, Ricaldi JN, Matthias MA, Diaz MM. Leptospirosis: a zoonotic disease of global importance. Lancet Infect Dis, 3: 757-771, 2003.
- Corwin A, Ryan A, Bloys W, Thomas R, Deniega B, Watts D. A. Waterborne outbreak of leptospirosis among United States military personnel in Okinawa, Japan. *Int J Epidemiol*. Rev 19:743–748, 1990.
- Ellis, W. A. Control of canine leptospirosis in Europe: time for a change? *Vet Rec*, 167,602-5, 2010.
- Everard CO, Fraser-Chanpong GM, James AC, Butcher LV. Serological studies on leptospirosis in livestock and chickens from Grenada and Trinidad. *Trans R Soc Trop Med Hyg*, 79(6):859-64, 1985.
- Lau, C. L., Smythe, L. D., Craig, S. B. and W, P. Climate change, flooding, urbanisation and leptospirosis: Trans Royal Soc Trop Med Hyg, 104,631–638, 2010.
- Lingappa J, Kuffner T, Tappero J, Whitworth W, Mize A, Kaiser R. Ingestion of contaminated water: *possible gene-environment interaction in an outbreak of leptospirosis*. Rev, 5:197–202, 2004.
- Machang'u, R. S., Mgode, G. and Mpanduji, D. Leptospirosis in animals and humans in selected areas of Tanzania. *Belg J Zool*, 127, 97–104, 1997.
- Mety, K E. and Dikken, H. Classification of the species of *Leptospira intrrogans* and history of its serovas. Groningen, the Netherlands: University Press Groningen, (OCoLC) 707944963, 1993.
- Mgode, GF Machang'u, RS Goris, MG Engelbert, M Sondij, S Hartskeerl RA. New Leptospira serovar Sokoine of serogroup Icterohaemorrhagiae from cattle in Tanzania. Intern J System Evolution Microb, 56(3) 593-597, 2006.
- Mohammed H, Nozha C, Hakim K, Abdelaziz F, Rekia B. *Leptospira*: Morphology, Classification and Pathogenesis. *J Bacteriol Parasitol* 2:120, 2011.
- Mwachui, M. A., Crump, L., HartskeerL, R., Zinsstag, J. and Hattendorf, J. Environmental and Behavioural Determinants of Leptospirosis Transmission: A Systematic Review. PLoS Negl Trop Dis, 9, e0003843, 2015.
- Stern EJ, Galloway R, Shadomy SV, Wannemuehler K, Atrubin D, Blackmore C, Wofford T, Wilkins PP, Ari MD, Harris L, Clark TA. Outbreak of leptospirosis among adventure race participants in Florida. Rev, 50:843–849, 2010.