

Effects of bottom-up and top-down interactions on the productivity of Iyieke Lake, Afikpo North, Ebonyi State, south-east Nigeria

Nwonumara, G. N. and Idumah, O. O.

Department of Applied Biology, Faculty of Science, Ebonyi State University, P.M.B. 53, Abakaliki, Ebonyi State, Nigeria

*Corresponding author: nwonumara.godwin@ebsu-edu.net

Abstract

The bottom-up and top-down interactions of the abiotic and biotic components of Iyieke Lake were studied for eleven months covering a pre-flood, flood and post-flood periods. During the study, some water quality parameters were measured while plankton samples were collected for identification in the laboratory using standard method. The results of the water quality parameters measured showed that water temperature was highest (35.00R°C) in February. pH (7.10) and transparency (0.98 m) were highest in August. Highest TDS (28.0 mg/L) and conductivity (56.00 µS/cm) were recorded in March (pre-flood period) while dissolved oxygen level was lowest (4.00 mg/L) in the pre-flood period (February and March). Nitrate (0.18 mg/L) was highest in August (flood period) and phosphate (0.25 mg/L) at the two extreme periods (pre-flood and post flood). The depth of the lake was highest (3.2 m) in August which was the period of inundation. The phytoplankton (Bacillariophyta, Chlorophyta, Cyanobacteria) and zooplankton (Rotifera, Cladocera, Copepoda) of the lake were made of three divisions and taxa each, respectively. Bacillariophyta dominated among the phytoplankton while Rotifera was the most abundant among the zooplankton. The study revealed that nutrients and zooplankton grazing were the major factors that affected phytoplankton abundance and biomass during the study.

Keywords: Flood; phytoplankton; Iyieke; water quality; zooplankton.

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Introduction

Phytoplankton play key roles in lake productivity by fixing energy through photosynthetic activities (Akin-Oriola, 2003). They occupy mainly the euphotic zone of aquatic ecosystem, where they utilize energy derived from the sun to build up small inorganic molecules into complex organic compounds that are stored in their tissue, hence are found at the base of aquatic food chain. Phytoplankton serve as food for numerous aquatic fauna including zooplankton, herbivorous fish fingerlings and ensure the stability of trophic interaction in aquatic ecosystem. Hence, the productivity of any aquatic ecosystem depend largely on the inert capacity of the phytoplankton to fix energy by utilizing inorganic materials within such ecosystem. Therefore, the abundance, diversity and biomass production of phytoplankton in a lentic system depend largely on the interaction with the physico-chemical variables of the ecosystem, predation by zooplankton and other carnivorous animals that may be feeding on the zooplankton and herbivorous fishes (Reynolds, 1988; Reynolds, 2006; Salmaso and Braioni, 2008; Okogwu and Ugwumba, 2013).

Zooplankton grazing have been reported among the factors that affect the species composition, abundance, diversity and biomass of phytoplankton in lentic ecosystem

(Reynolds, 1998). Zooplankton play a vital role in energy flow in aquatic food web, serving both as consumer of most phytoplankton, bacteria and as prey for macro crustaceans, insects and fishes (Infante and Abella, 1985). Phytoplankton species, *Senedesmus acuminatus*, *Ankistrodesmus* have been reported to serve as suitable diet for *Moina micrura* and have been used as feed stock in the maintenance of some zooplankton species, *Anuraeopsis fissa* and *Daphnia* (Goulden *et al* 1982; Dumont *et al* 1995). Zooplankton also help to regulate phytoplankton and microbial productivities by enhancing clear water phase production due to its filtering activities (Mathivanan *et al* 2007). However, the trophic interactions between phytoplankton and zooplankton are regulated majorly by environmental and biological factors which affect the growth of the communities (Abdel-Aziz *et al* 2006). Hence, high flow rate, discharge, residence time, transparency, water temperature, nutrient concentrations, and suspended solids have been listed as the major regulatory factors of phytoplankton species composition, abundance, diversity and biomass in lentic and lotic ecosystems (Reynolds, 2006; Okogwu and Ugwumba, 2013). Also, zooplankton grazing and selective predation have been reported to have direct influence on the biomass and species composition of phytoplankton in lentic system



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(Sterner, 1989; Sommer *et al* 2001; Kagami *et al* 2002). Therefore, this research is designed to assess the effects of bottom-up and top-down interactions on the abundance and biomass accumulation of phytoplankton in Iyieke Lake within the study period.

Methodology

Study area

Iyieke Lake is a natural shallow floodplain lake (05° 50' 33.49 N, 007° 56' 40.88 E) located 50 m away from the Cross River channel (Okogwu and Ugwumba, 2012), near Ndibe beach. It is one of the numerous lakes within the drainage basin of the Cross River system (Nwonumara and Okogwu, 2013). Seasonal flooding of the lake by the Cross River divided the hydrology into three distinct periods which include the pre-flood period (January-May), when the lake remains completely separated from the river; the flood period (June-August), when the lake is inundated; and the post-flood period (September-December), when the flood recedes (Nwonumara and Okogwu, 2013). These periods cover both dry (November-April) and wet or rainy season (May-October). Major anthropogenic activities in and around the lake are fishing and farming.

Sample collection and field determinations

Water samples for physico-chemical and plankton analyses were collected monthly from the littoral and pelagic zones of the lake for eleven months, which covered the pre-flood (February-May), flood (June-August), and the post-flood (September-December) periods. Samples were pooled from different locations from each zone. Water temperature, dissolved oxygen (DO) and pH were measured *in situ* using Hanna digital thermometer, DO meter (Model HI 9142), and pH meter (Model HI 77700P), respectively. Total dissolved solids (TDS) and conductivity were also measured at the study site using Hanna TDS (Model HI 98108) and conductivity (Model HI 98303) metres, respectively. Depth was measured from calibrated poles permanently mounted in the lake while transparency was determined using a Secchi disc. Phosphate (PO₄-P) and nitrate (NO₃-N) were measured using standard procedures (APHA 2005). Plankton samples were collected from the same zones as in the physico-chemical variables using plankton net, pooled into a 1 litre plastic container holding 400 ml of buffered formalin (4 %) until the 1 litre mark is reached.

Laboratory and data analyses

Morphological identification of plankton was done to the species level with standard keys and guides of Gebrüder (1978), Prescott (1978), Jeje and Fernando (1986), Williams (1991), WRC, 2002 and Nwankwo (2004), using Olympus binocular microscope at x400 magnification (Model BHTU BH-2). The quantitative assessment of plankton abundance was carried out by identifying/counting individuals of each species deposited in Utermohl

chambers and presented as the number of individuals per litre (ind/L). Plankton biovolume (mm³) was estimated by measuring individual cells and their volumes were calculated according to geometrical solids (Ruttner-Kolisko, 1977; Hillebrand *et al* (1999). Biovolume (mm³) was, then, obtained by multiplication of the density of each species by the average volume of its cells. Specific biomass was expressed in µg (fresh weight)/L, assuming the specific density of a phytoplankton of 1 g/cm³ (Edler, 1979).

Variations in physico-chemical parameters, plankton abundance and biomass among pre-flood, flood and post-flood periods during the study were tested statistically using one sample t-test and one way analysis of variance (ANOVA). Values were considered significant at $p < 0.05$ levels. All statistical analyses were performed using statistical package for social science (SPSS) version 20.

Results

Physico-chemical variables

Water temperature was highest (35°C) in February, the pre-flood period and the water pH ranged from 6.50 to 7.10 (Figure 1). Both TDS and conductivity were highest in March (pre-flood period) with 28.0 mg/L and 56.0 µS/cm, respectively. Dissolved oxygen was lowest (4.00 mg/L) in the pre-flood period (February and March) and highest (1.00 mg/L) in June (flood period) (Figure 1). Nitrate (0.18 mg/L) was highest in August and phosphate (0.75 mg/L) in February and November. The depth of the lake (3.2 m) and transparency (0.98 m) were highest in August, the period of inundation (Figure 2). Temporal variation in the physico-chemical variables measured were not significant ($p > 0.05$) while water temperature, pH, transparency, conductivity and DO varied significantly ($p < 0.05$) between the study periods (Table 1).

Phytoplankton and zooplankton species composition

Three phytoplankton divisions were identified during the study (Table 2). The divisions are Chlorophyta, Bacillariophyta and Cyanobacteria with Bacillariophyta (309 ind/L) as the most abundant and the post-flood period (Table 2). Species with the highest frequency of occurrence among the Bacillariophyta were *Achnanthes* and *Gryosigma* spp. (Table 2). ANOVA result showed that difference in the abundance and biomass production of Cyanobacteria was significant ($p < 0.05$) during the study period.

The zooplankton species identified during the study were comprised of three broad taxonomic groups, namely Rotifera, Cladocera and Copepoda (Table 3). Copepoda (496 ind/L) was the most abundant while Rotifera (12 ind/L) was the least all in the post-flood period. *Collotheca* spp (16.66 %), *Diaphanosoma* spp (28.58 %) and *Thermocyclops oithonoides* (23.53 %) were the highest in percentage relative abundance and in the pre-flood and flood periods (Table 3). There was no significant difference ($p > 0.05$) in the abundance of the zooplankton taxa identified during the study period.

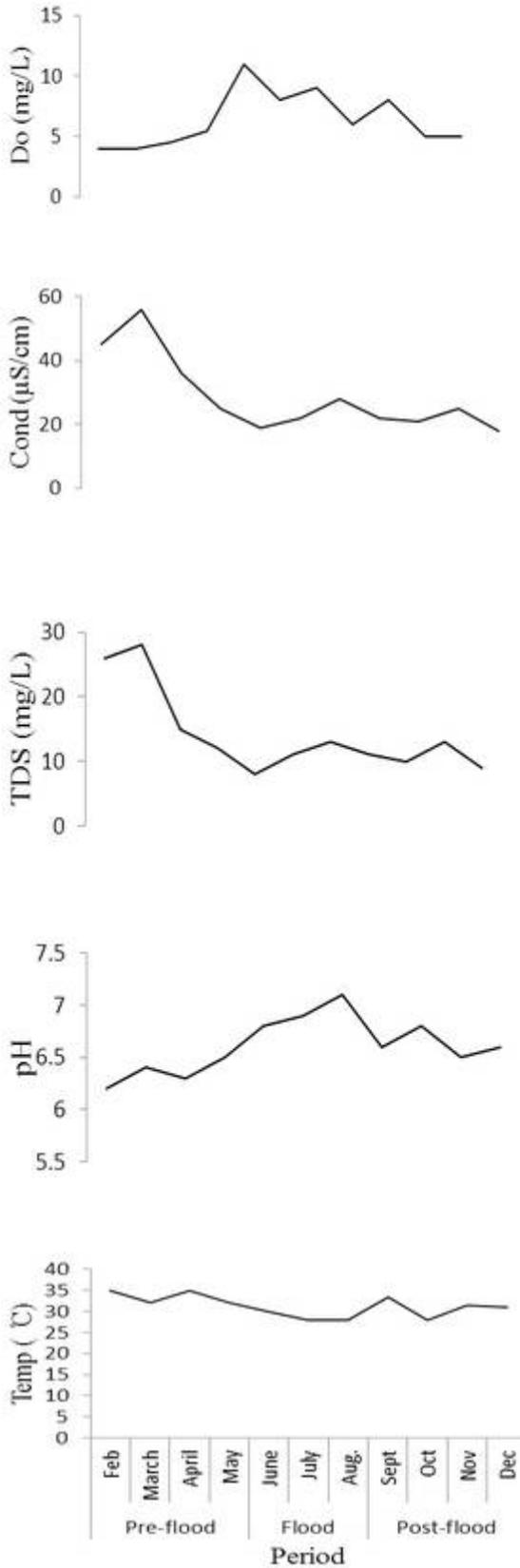


Figure 1: Temperature, pH, TDS, Conductivity and DO of the study site.

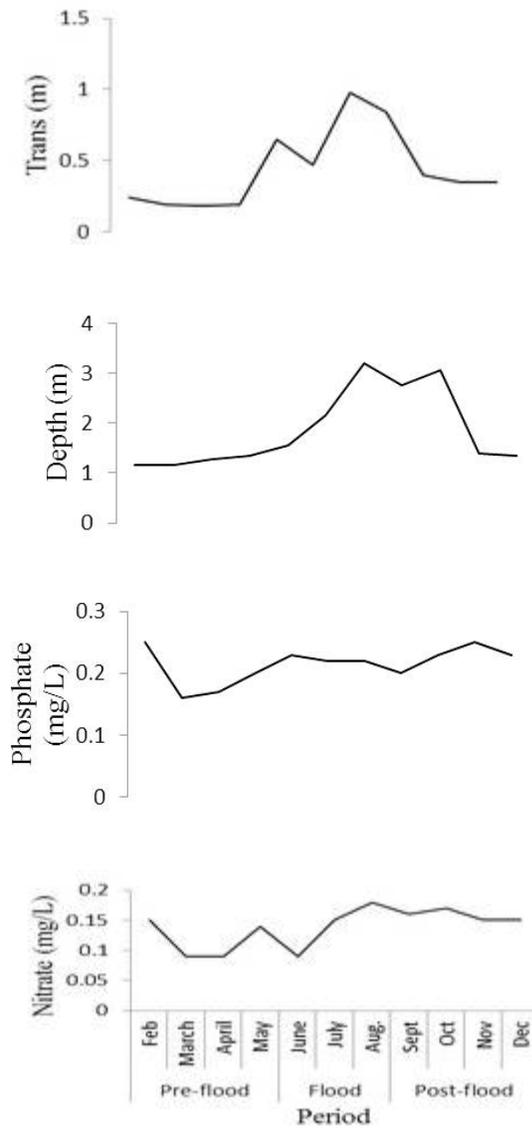


Figure 2: Nitrate, Phosphate, Depth and Transparency of the study site.

Phytoplankton biomass-zooplankton abundance relation

Total phytoplankton biomass had negative correlation (-0.21, $r = 0.53$) with total zooplankton abundance though not significant, thus indicating the negative effect of zooplankton grazing on phytoplankton biomass. At lower zooplankton abundance, phytoplankton biomass was observed to increase. However, when zooplankton abundance increased in August and November, phytoplankton biomass declined (Figure 10).

Principal coordinate (PC) analysis showed that dissolved oxygen (DO) was influenced by depth while temperature influenced conductivity and TDS (Figure 11). Nitrate, phosphate and zooplankton interactions affected phytoplankton biomass most (Figure 11).

Table 1: Statistical description of spatial and temporal variations in physico-chemical variables of Iyieke Lake during the study.

| Parameter | Pre – flood | Flood | Post - flood | p - value | Monthly | N | Range | P- value | t -statistic |
|-------------------------|-------------|------------|--------------|-----------|-------------|----|-------|----------|--------------|
| | Mean±SD | Mean±SD | Mean±SD | | Mean±SD | | | | |
| Temperature (°C) | 33.50±1.73 | 28.67±1.15 | 31.00±2.27 | 0.026 | 31.27±2.60 | 11 | 7.00 | 1.00 | 0.003 |
| pH | 6.35±0.13 | 6.93±0.15 | 6.63±0.13 | 0.002 | 6.61±0.27 | 11 | 0.90 | 0.99 | -0.01 |
| Transparency (m) | 0.20±0.03 | 0.70±0.26 | 0.49±0.24 | 0.028 | 0.44±0.27 | 11 | 0.80 | 1.00 | 0.00 |
| TDS (mg/L) | 20.25±7.93 | 10.67±2.52 | 10.75±1.71 | 0.051 | 14.18±6.65 | 11 | 20.00 | 0.99 | 0.001 |
| Conductivity (µS/cm) | 40.50±13.18 | 23.00±4.58 | 21.50±2.89 | 0.027 | 28.82±12.04 | 11 | 38.00 | 1.00 | -0.001 |
| Dissolved oxygen (mg/L) | 4.50±0.71 | 9.33±1.53 | 6.00±1.41 | 0.003 | 6.36±2.30 | 11 | 7.00 | 0.99 | 0.005 |
| Nitrate (mg/L) | 0.12±0.03 | 0.14±0.05 | 0.16±0.01 | 0.242 | 0.14±0.03 | 11 | 0.09 | 0.86 | -1.84 |
| Phosphate (mg/L) | 0.20±0.04 | 0.22±0.01 | 0.23±0.02 | 0.268 | 0.21±0.03 | 11 | 0.09 | 0.62 | 0.51 |
| Depth (m) | 1.23±0.10 | 2.30±0.84 | 2.13±0.89 | 0.135 | 1.85±0.79 | 11 | 2.05 | 0.99 | 0.01 |

Table 2: Percentage relative abundance of phytoplankton species of Iyieke lake during the study period (February-December, 2011).

| Phytoplankton Division | Pre-flood | Flood | Post-flood |
|----------------------------------|--------------------|--------------------|--------------------|
| Chlorophyta | 12 (4.32) | | 74 (12.84) |
| <i>Ankistrodesmus</i> spp. | 0.36 | * | * |
| <i>Pithophora</i> sp. | 1.44 | * | * |
| <i>Schizomeris</i> spp. | 1.80 | * | 0.17 |
| <i>Sticococcus</i> sp. | 0.72 | * | * |
| <i>Strauastrium laptocladium</i> | * | * | 0.17 |
| <i>Spirogyra</i> sp. | * | * | 12.50 |
| Bacillariophyta | 247 (88.85) | 235 (97.11) | 309 (53.65) |
| <i>Achmanthes</i> sp. | 5.04 | 20.66 | 15.23 |
| <i>Aulacoseira granulata</i> | 7.91 | 0.83 | 2.60 |
| <i>Bacillaria</i> spp. | 0.72 | * | * |
| <i>Bacteriastrium hyalinum</i> | 0.36 | * | * |
| <i>Cocconeis</i> spp. | 21.58 | 2.89 | 2.95 |
| <i>Coscinodiscus</i> spp. | 2.52 | 3.31 | 1.04 |
| <i>Gyrosigma balticum</i> | 46.40 | 69.01 | 30.03 |
| <i>Hemidiscus hardimanianus</i> | 0.36 | * | * |
| <i>Hyalodiscus</i> sp. | 0.36 | * | * |
| <i>Nitzschia</i> spp. | 2.16 | * | 2.08 |
| <i>Skeletonema</i> spp. | 0.72 | * | * |
| <i>Surirella linearis</i> | 0.36 | * | * |
| <i>Tabellaria</i> sp. | 0.36 | 0.41 | * |
| Cyanobacteria | 19 (6.83) | 7 (2.89) | 193 (33.51) |
| <i>Aphanizomenon flos-aquae</i> | 6.83 | 2.89 | 13.02 |
| <i>Glocotrichia</i> sp. | * | * | 20.49 |

Key: * indicate absence of species.

Table 3: Percentage relative abundance of zooplankton species of Iyieke Lake during the study period (February-December, 2011)

| Zooplankton species | Study period | | |
|--|-------------------|------------------|------------------|
| | Pre-flood | Flood | Post-flood |
| Rotifera | 72 (43.57) | 14 (4.82) | 12 (2.11) |
| <i>Anuraeopsis</i> sp. | * | 0.32 | 0.60 |
| <i>Brachionus</i> spp. | 1.92 | 0.49 | 0.30 |
| <i>Keratella cochlearis varrobusta</i> | 0.64 | * | * |
| <i>Notholca</i> spp. | 1.93 | 0.32 | 0.15 |
| <i>Collotheca</i> spp. | 16.66 | 2.56 | 0.15 |
| <i>Colurella</i> spp. | 1.92 | 0.16 | * |
| <i>Lepadella</i> sp. | 0.64 | 0.16 | * |
| <i>Proales</i> sp. | 3.20 | * | * |
| <i>Euchlanis deflexa deflexa</i> | 0.64 | * | 0.15 |

Table 3: Percentage relative abundance of zooplankton species of Iyieke lake during the study period (February-December, 2011) continued.

| Zooplankton species | Study period | | |
|----------------------------------|-------------------|--------------------|--------------------|
| | Pre-flood | Flood | Post-flood |
| <i>Hexarthra</i> spp. | 1.28 | * | * |
| <i>Lecane</i> spp. | 1.28 | 0.16 | * |
| <i>Testudinella species</i> | * | 0.16 | 0.61 |
| <i>Macrochaetus</i> sp. | 0.64 | * | 0.15 |
| <i>Trichocerca</i> spp. | 12.82 | 0.49 | * |
| Cladocera | 64 (41.03) | 223 (36.62) | 149 (22.68) |
| <i>Diaphanosoma</i> sp. | 12.82 | 28.58 | 11.85 |
| <i>Pseudosida bidentata</i> | 2.56 | 2.43 | 2.43 |
| <i>Ceriodaphnia cornuta</i> | * | 0.16 | 1.24 |
| <i>Daphnia longispina</i> | 5.77 | 0.16 | * |
| <i>Scapholeberis kingi</i> | * | * | 0.46 |
| <i>Simocephalus</i> spp. | 2.56 | 2.13 | 5.62 |
| <i>Moina micrura</i> | 7.35 | * | 0.46 |
| <i>Moinodaphnia macleayi</i> | 1.92 | 0.99 | 0.30 |
| <i>Bosmina longirostris</i> | 0.64 | * | * |
| <i>Bosminopsis deisteri</i> | 0.64 | 0.33 | * |
| <i>Echinisca</i> sp. | * | 0.49 | 0.30 |
| <i>Grimaldina brazzai</i> | 0.64 | * | * |
| <i>Guemella raphaelis</i> | * | 0.16 | * |
| <i>Ilyocypris spinifer</i> | 0.64 | * | * |
| <i>Macrothrix geoldi</i> | 0.64 | * | * |
| <i>Alona</i> spp. | 3.94 | 0.82 | * |
| <i>Chydorus</i> spp. | * | 0.32 | * |
| <i>Leydigia acanthocercoides</i> | 0.64 | * | * |
| <i>Pleuroxus hamatus hamatus</i> | 0.64 | * | * |
| Copepoda | 21 (13.46) | 372 (61.08) | 496 (75.49) |
| <i>Cryptocyclops bicolor</i> | * | 1.32 | 1.22 |
| <i>Ectocyclops sphaeratus</i> | 0.64 | * | 0.15 |
| <i>Eucyclops serrulatus</i> | 0.64 | 0.49 | 0.15 |
| <i>Halicyclops troglodytes</i> | 0.64 | * | 0.30 |
| <i>Mesocyclops leuckarti</i> | * | 0.16 | 0.15 |
| <i>Metacyclops minutus</i> | * | 0.49 | 5.78 |
| <i>Microcyclops varicans</i> | 1.92 | 13.90 | 18.69 |
| <i>Thermocyclops neglectus</i> | 0.64 | * | * |
| <i>Thermocyclops oithonodes</i> | 7.06 | 23.53 | 16.72 |

Key: * indicate absence of species.

(Adopted and modified from Nwonumara and Okogwu, 2013).

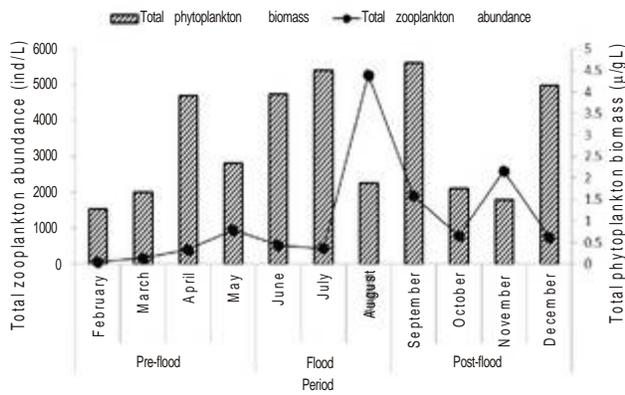


Figure 10: Phytoplankton biomass – zooplankton abundance relation.

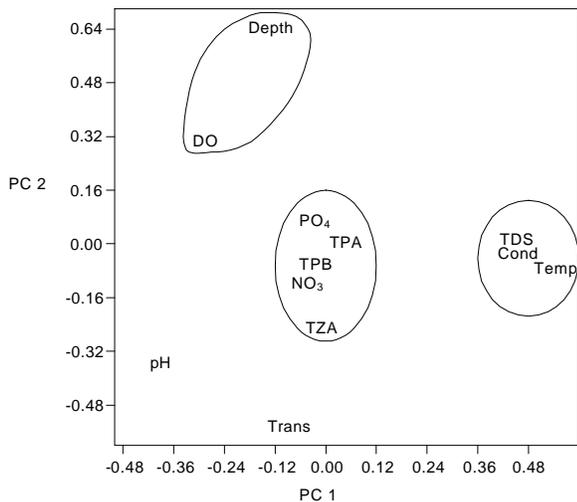


Figure 11: Principal coordinate of factors interaction in Iyieke Lake during the study.

Discussion

Iyieke Lake is one of the floodplain lakes along the Cross River Floodplain. The lake provides numerous services which include provision of water for irrigation farming, habitat for several aquatic flora and fauna, temperature moderation, regulation of river flow, flood control and recharging of ground water aquifer. Most importantly, it serves as spawning ground for different fish species (Nwonumara, 2012). Hence, study of the bottom-up and top-down interactions of the lake will provide basic knowledge of the functionality of the important ecosystem.

The study therefore revealed that water temperature, pH, transparency and DO varied significantly during the study period. Temperature variation could be linked to the influence of season on the variable. The pH of the lake was slightly acidic throughout the study period except in the flood period when it reached neutral point. Lower pH recorded in the pre-flood and post-flood periods could be linked to higher water temperature which enhanced microbial activity at the benthic zone (Atobatele and Olutona, 2013) that released more organic acid from

decomposition of organic matter at these periods. It could also be due to the decomposition of submerged plants which released more organic acids into the water in the post-flood period. Transparency of the lake was at the peak during flood period which, could be due to the lake sill height and the indirect connectedness of Cross River, preventing direct inflow of flood water. These according to Squires and Lesack, 2002, increase the progressive decantation of sediments as the distance from the connection point with the main river channel increases. Meanwhile, Okogwu *et al* (2009) have reported high transparency at the flood period in floodplain lakes. Inflow of flood water from Cross River increased the depth and volume of the lake. Thus, might have contributed to the higher dissolved oxygen (DO) concentration recorded in the flood period through water turbulence which facilitated air-water mixing and consequent increase in dissolved oxygen level (Nwonumara and Okogwu, 2013). Gautam *et al* (1993) as well as Sivakumar and Karuppasamy (2004) have also reported increase in DO level during flood period in lakes which they attributed to respiration from inundated vegetation. Phosphate concentration was low in the pre-flood period. This could be attributed to utilization by phytoplankton and aquatic plants (Wani and Subla, 1990; Sivakumar and Karuppasamy, 2004). However, higher values recorded in the flood and post-flood period could be due to the inflow of phosphate fertilizer from riparian farmland (Nwonumara and Okogwu, 2013) and decomposed plants submerged in water during flood (Lauridsen *et al* 1998).

The phytoplankton of the lake was represented by similar divisions that have been recorded in tropical waters such as Paranapanema River reservoirs (Nogueira *et al* 2010), Ikot Okpora, Obubra and Ejagham Lakes (Offem *et al* 2011a), Ikwori Lake (Offem *et al* 2011b), Nguru Lake (Mohammad *et al* 2012), Cross and Asu Rivers (Okogwu and Ugwumba, 2013), Shagari Reservoir (Magami *et al* 2014), Bhoj wetland (Bhat *et al* 2015), Khami River (Dzinomwa and Ndagurwa, 2017) and so on. However, the phytoplankton divisions at Iyieke Lake were fewer compared to some of the tropical waters mentioned above. This could be due to the nutrient status of the lake which may not have supported the proliferation of the phytoplankton divisions that are not recorded during the study. Bacillariophyta was the most frequently encountered among the three divisions recorded especially in the pre-flood period. This could be due to its exceptional ability to utilize silicate, an advantage that might have made it proliferate more than the other divisions (Akpan, 1997). However, low abundance of the Chlorophyta could be due to higher predation by zooplankton as some species (*Scenedesmus* spp.) among the division have been reported to serve as food for zooplankton (*Moina micrura*, *Anuraeopsis fissa*) (Dumont *et al* 1995). These zooplankton species were among the ones recorded during the study and could have been responsible for the low abundance of Chlorophyta due to their predatory characteristics. On the other hand, low abundance of the

Cyanobacteria may have been caused by the nutrient status of the water during the study period since most species of the group do not grow and proliferate in nutrient poor waters (Ambasht and Ambasht, 2005).

The limnological structure of Iyieke Lake during the study had positive effect on zooplankton production compared to phytoplankton. Among the factors that boosted zooplankton proliferation could be temperature at the pre-flood period and increased water level at the flood period. Increased temperature has been reported to be favourable to the parthenogenetic reproductive pattern of Rotifers and the attainment of peak reproductive period in fewer days (Akin-Oriola, 2003; Imoobe and Adeyinka, 2010). Similarly, increased water level to the littoral zone of the lake enhanced the hatching of resting Cladoceran eggs lying dormant in flooded areas (Okogwu, 2010). Increased water level also enhanced the influx of species from other water bodies which include Bob Eric Lake, Ehoma Lake, Egwebe Lake and Cross River which, connect to Iyieke lake during inundation (Okogwu, 2010; Nwonumara and Okogwu, 2013).

However, the negative correlation between the phytoplankton biomass and zooplankton abundance of the lake during the study could be attributed to grazing or massive predation by zooplankton and herbivorous fishes. The grazers exerted a top down effect on the phytoplankton, which reduced the proliferation and directly affected the biomass production.

In conclusion, the study revealed that phytoplankton abundance and biomass production were mainly controlled by eutrophic factor (bottom-up) and grazing by zooplankton (top-down) influence.

References

- Abdel Aziz, A. N. E., Gharib, S. M. and Dorgham, M.M. 2006. The interaction between phytoplankton and zooplankton in a Lake-Sea connection, Alexandria, Egypt. *International Journal of Oceans and Oceanography*, 1(1): 151-165.
- Akin-Oriola, G. A. 2003. Zooplankton associations and environmental factors in Ogunpa and Ona Rivers, Nigeria. *Revista de Biología Tropica*, 51: 99-106.
- Akpan, E. R. 1997. *Spatial and Seasonal distribution of phytoplankton in the Cross River estuary, Nigeria*. A paper delivered at the 6th Annual Conference of the Nigerian Society for Biological Conservation 26th-28th November, 1997, Calabar, Nigeria.
- Ambasht, R. S. and Ambasht, P. K. 2005. *Environment and Pollution*. Fourth Edition. CBS Publishers. New Delhi.
- APHA. 2005. *Standard Methods for the Examination of Water and Wastewater Analysis*. 21st Edition. American Public Health Association. Washington D.C., USA, 1099pp.
- Atobatele, E. O. and Olutona, G. O. 2013. Spatio-seasonal physico-chemistry of Aiba stream, Iwo, Nigeria. *African Journal of Biotechnology*, 12 (14): 1630-1635.
- Bhat, N. A., Wanganeo, A. and Raina, R. 2015. Variability in Water Quality and Phytoplankton Community during Dry and Wet Periods in the Tropical Wetland, Bhopal, Indian. *Journal of Ecosystem and Ecography*, 5(2): 2-8.
- Dumont, H., Sharma, S. S. S. and Jawahar, A. A. 1995. Laboratory studies on the population dynamics of *Anuraeopsis fissa* (Rotifera) in relation to food density. *Freshwater Biology*, 33: 39-46.
- Dzinomwa, T. and Ndagurwa, H. G. T. 2017. Effect of land use on water quality and phytoplankton community in the tropical Khami River in semi-arid south-west Zimbabwe. *African Journal of Aquatic Science*, 42 (1): 83-89.
- Edler L. 1979. Recommendations for marine biological studies in the Baltic Sea – phytoplankton and chlorophyll. *Baltic Marine Biologists*, 5: 1-39.
- Gautam, A., Joshi, V. P. and Sati, O. P. 1993. Physico-chemical characteristics of sewage and its impact on water quality of Alkananda at Srinagar (Garhwal). *Journal of Ecotoxicology and Environmental Monitoring*, 3: 61-63.
- Gebruder, B. 1978. *Rotatoria*. Druck Graphik, Muchen, Stuttgart, Berlin, 232pp.
- Goulden, C. E., Henry, L. L. and Tessier, A. J. 1982. Body size, energy reserves, and competitive ability in three species of Cladocera. *Ecology*, 63: 1780-1789.
- Hillebrand, H., Durselen, C. D., Kriechfeld, D., Pollinger, D. and Zohray, T. 1999. Biovolume calculation for pelagic and benthic microalgae. *Journal of Phycology*, 35: 403-424.
- Imoobe, T. O. T. and Adeyinka, M. L. 2010. Zooplankton-based assessment of the trophic state of a tropical forest river. *International Journal of Fisheries and Aquaculture*, 2(2): 64-70.
- Infante, A. and Abella, S. E. B. 1985. Inhibition of *Daphnia* by *Oscillatoria* in Lake Washington. *Limnology and Oceanography*, 30: 1046-1052.
- Jeje, C. Y. and Fernando, C. H. 1986. *A practical guide to the identification of Nigerian zooplankton* (Cladocera, Copepoda and Rotifera). Kanji Lake Research Institute, New Bussa, Nigeria Publication, Nigeria, 142pp.
- Kagami, M., Yoshida, T., Gurung, T. and Urabe, J. 2002. Direct and indirect effects of zooplankton on algal composition in *in situ* grazing experiments. *Oecologia*, 133: 356-363.
- Lauridsen, T., Jeppesen, E., Sondergaard, M. and Lodge, D. M. 1998. Horizontal migration of zooplankton: Predator-mediated use of macrophyte habitat. In: Jeppesen *et al* (Eds). *The Structuring Role of Submerged Macrophytes in Wetlands*. New York, NY: Springer Verlag, pp. 233-239.
- Magami, I. M., Adamu, T. and Aliero, A. A. 2014. Physicochemical flux and phytoplankton diversity in Shagari Reservoir, Sokoto, Nigeria. *Nigerian Journal of Basic and Applied Science*, 22(3&4): 67-72.
- Mathivanan, V. P., Vijayan, S. S. and Jeyachitra, O. 2007. An assessment of plankton population of Cauvery River with reference to pollution. *Journal of Environmental Biology*, 28(2): 523-526.
- Mohammad, M. A., Mohammad, L. B. and Auta, J. 2012. Effects of Physico-chemical factors on seasonal dynamics of the phytoplankton in Nguru Lake, north-eastern Nigeria. *Journal of Natural Sciences Research*, 2(8): 74-81.
- Nogueira, M. G., Ferrareze, M., Moreira, M. L. and Gouvêa,

- R. M. 2010. Phytoplankton assemblages in a reservoir cascade of a large tropical – subtropical river (SE, Brazil). *Brazilian Journal of Biology*, 70(3): 781-793
- Nwankwo, D. I. 2004. *Practical guide to the study of Algae*. JAS Publishers, Lagos, Nigeria. 99pp.
- Nwonumara, G. N. 2012. The physico-chemistry and plankton of Iyieke Lake, Southeast Nigeria. Master of Science thesis submitted to the Department of Applied Biology, Faculty of Science, Ebonyi State University, Abakaliki, Ebonyi State, Nigeria (Unpublished), 87pp.
- Nwonumara, G. N. and Okogwu, O. I. 2013. The impact of flooding on water quality, zooplankton composition, density and biomass in Lake Iyieke, Cross River-Floodplain, Southeastern Nigeria, *Zoology and Ecology*, 23(2): 138-146.
- Offem, B. O., Ayotunde, E. O., Ikpi, G. U., Ada, F. B. and Ochang, S. N. 2011a. Plankton-based assessment of the trophic state of three tropical lakes. *Journal of Environmental Protection*, 2: 304-315.
- Offem, B. O., Ayotunde, E. O., Ikpi, G. U., Ochang, S. N. and Ada, F. B. 2011b. Influence of seasons on water quality, abundance of fish and plankton species of Ikwori Lake, south-eastern Nigeria. *Fisheries and Aquaculture Journal*, 13: 1-18.
- Okogwu, O. I., Nwani, C. D., and Ugwumba, A. O. 2009. Seasonal variations in the abundance and biomass of microcrustaceans in relation to environmental variables in two shallow tropical lakes within the Cross River Floodplain, Nigeria. *Acta Zoologica Lituanica*, 19(3): 205-215.
- Okogwu, O. I. and Ugwumba, A. O. 2012. Response of phytoplankton functional groups in fluctuating water level in two shallow floodplain lakes in Cross River, Nigeria. *Inland Waters*, 2: 37-46.
- Okogwu, O. I. and Ugwumba, A. O. 2013. Seasonal dynamics of phytoplankton in two tropical rivers of varying size and human impact in Southeast Nigeria. *International Journal of Tropical Biology*, 61(4): 1827-1840
- Prescott, G. W. 1978. *How to know the freshwater algae*. W.C. Brown, McGraw-Hill, Boston, 285pp.
- Reynolds, C. S. 1988. What factors influence the composition of phytoplankton in lakes of different trophic status? *Hydrobiologia*, 369/370: 11-26.
- Reynolds, C. S. 2006. *The ecology of phytoplankton. First Edition*. Cambridge University Press, Edinburgh Building, Cambridge, UK, 551pp.
- Ruttner-Kolisko, A. 1977. Suggestions for biomass calculation of plankton rotifers. *Hydrobiologie–Beiheft Ergebnisse der Limnologie*, 8: 71-76.
- Salmaso, N. and Braioni, M. G. 2008. Factors controlling the seasonal development and distribution of the phytoplankton community in the lowland course of a large river in Northern Italy (River Adige). *Aquatic Ecology*, 42: 533-545.
- Sivakumar, K., and Karuppasamy, R. 2004. Factors affecting productivity of phytoplankton in a reservoir of Tamilnadu, India. *American-Eurasian Journal of Botany*, 1: 99-103.
- Sommer, M., Sommer, F., Santer, B., Jamieson, C., Boersma, M., Becker, C. and Hansen, T. 2001. Complementary impact of copepods and cladocerans on phytoplankton. *Ecology Letters*, 4: 545-550.
- Squires, M. and Lesack, L. F. W. 2002. Water transparency and nutrients as controls on phytoplankton along a flood-frequency gradient among lakes of the Mackenzie Delta, western Canadian Arctic. *Canadian Journal of Fisheries and Aquatic Sciences*, 59(8): 1339-1349.
- Sterner, R. W. 1989. The role of grazers in phytoplankton succession. In: *Plankton Ecology*: Sommer, U. (Ed.). Succession in Plankton Communities. Springer-Verlag, Berlin, pp. 70-107.
- Wani, I. A., and B. A. Subla. 1990. Physico-chemical features of two shallow Himalayan Lakes. *Bulletin of Environmental Science*, 8: 33-49.
- WRC. 2002. *Guide to the freshwater invertebrate of southern Africa*. Water Research Commission, South Africa, 300pp.
- Williams, C. E. 1991. Copepods. In: *Ecology and classification of north American freshwater invertebrates*, C. Thorp and N. Covich (Eds), Academic Press, New York, pp. 799-822.

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