Comparison of neutron scattering and DFM capacitance instruments in measuring soil water evaporation

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ABSTRACT

Soil water evaporation is an important parameter that needs to be accurately measured for the design of water-efficient agricultural systems. With this study, the abilities of the DFM capacitance probes and a neutron water meter (NWM) to measure evaporation from the soil surface were compared. Measured evaporation was compared to the control values measured with mini-lysimeters. Calibration of DFM capacitance probes and the NWM was done in the laboratory using the topsoil of a Bainsvlei soil form. Field measurements of soil water content were done on the same Bainsvlei soil. Calibration results indicated a good correspondence ($r^2 = 0.99$) between the measured values and known volumetric soil water contents. There was no significant difference ($p = 95\%$) between the DFM evaporation measurements and the control, whereas the NWM and control differed significantly. It was concluded that the DFM capacitance probe is a better tool than the NWM in measuring evaporation from the topsoil surface.

Keywords: neutron water meter, capacitance probes, evaporation, soil wetness

INTRODUCTION

In order to use the scarce water resource for agricultural production effectively, it is important that soil water content be measured as accurately as possible. The amount of water in the soil determines how strongly water molecules are retained by the soil matrix. Soil water can range from thin hygroscopic films tightly held around soil particles to saturation, where all of the soil macro-pores are filled with water. The amount of soil water determines the forces that control its movement. These forces in turn determine the availability of soil water for plant use and processes of water movement like evaporation and drainage. Thus, accurate measurement of soil water determines how accurately these components of the water balance can be quantified.

The conventional method of determining soil water content is described by Gardner (1986). With this method, a soil sample is taken and kept in an air-tight container until it is weighed; hereafter it is oven-dried at 105°C for 24 h and weighed again. The difference between the wet and dry mass expressed as a percentage of the dry mass provides the gravimetric soil water content. This method of measurement, although very accurate, is time consuming and costly. Furthermore, since the measurement employs destructive sampling, repeated measurements on the same spot are not possible. To overcome this constraint, indirect methods of soil water content measurement have been developed. One such method is the neutron water meter (NWM). This method was proposed in the 1940s and has been in use since with continual improvement (Bell, 1987). Despite the improvement, the NWM still has some drawbacks. The major concern is the health hazard of exposure to the radioactive source that is needed for neutron scattering. Besides this, although measurements can be done repeatedly, continuous logging of soil water content has not been possible. These drawbacks have been addressed with the introduction of capacitance-based soil water content measurements. The advancement of electronics has enabled capacitance probes to record continuous measurements of soil water content. The ease of use and the relatively cheaper availability of such probes make them a good candidate for use. To capitalise on these advantages due consideration should be given to measurement volumes and installation. Capacitance probes have smaller measurement volumes compared to those of NWM and require careful installation ensuring good contact with the soil.

Measuring the changes in soil water content to quantify evaporation from the soil surface has been done with various instruments. The NWM has been used to measure soil water content at different points in time and, if all the other components of the soil water balance equation are measured, it is possible to calculate the loss of soil water through evaporation from the soil surface (McGowan and Williams, 1980; Bennie et al., 1994; 1998). With the application of the same principle of monitoring change, some studies quantifying evaporation using capacitance probes have also been done (Verhoef et al., 2006; Mounzer et al., 2008). Comparison between the NWM and capacitance probes with regard to measuring soil water accurately has been done (Evett et al., 2002; Heng et al., 2002; Hossain, 2008). However, no comparative studies with regard to the measurement of evaporation have been done. The amount of water lost through evaporation from the top horizon of the soil profile can be considerable, although the change in soil water content induced by evaporation is small when considered on daily time intervals. The comparison with regard to the ability to capture the change in soil water content induced by evaporation is thus crucial.

The objective of this study was to compare the NWM and the DFM capacitance probe to measure evaporation from the soil surface against a control measured with another reliable method.

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MATERIALS AND METHODS

Description of the experiment

The field experiment was conducted on the Kenilworth-Bainsvlei ecotope, located in Bloemfontein, South Africa. The soil surface was bare and kept weed-free using herbicide. Soil water measurements were taken on rain-free days. Evaporation was taken as the change in soil water content between consecutive measurements. The micro-lysimeter, DFM capacitance probes and NWM were used to measure the soil water content. For most evaporation studies the micro-lysimeter has been used (Boast and Robertson, 1982; Wythers et al., 1999; Bremer, 2003). Micro-lysimeter measured evaporation served as a control when comparing the evaporation measured by the DFM capacitance probe and NWM (Paruelo et al., 1991).

The comparison of the DFM probe and NWM was done within a larger experiment where evaporation from 11 different sections of the micro-landscape of the in-field rainwater harvesting (IRWH) system was monitored (Zerizghy, 2012). The experiment was repeated in 5 different drying cycles (2009, DOY (day of year) 315–348; 2010, DOY 60–61).

Soil water measurement

DFM capacitance probes

DFM capacitance probes (DFM Software Solutions, South Africa) were used as one of the methods to measure the soil water content. The capacitance-based probes measure soil water by measuring the dielectric constant of the soil (Hossain, 2008). The dielectric constants of various materials differ; specifically, the dielectric constant of water is much higher than that of soil. Thus, depending on the amount of water in the soil the dielectric value changes and this can be quantified by capacitance sensors. The DFM probes (Fig. 1) are multilevel soil water and temperature measuring probes. The probes had a length of 1 200 mm with 6 sensors placed at 200, 400, 600, 800, 1 000 and 1 200 mm. The centre of the first sensor (80 mm in length) was kept 150 mm from the soil surface. The probes can take measurements at intervals set by the user.

Neutron water meter (NWM)

The NWM (Fig. 2) is a reliable method of measuring soil water content. This instrument uses fast neutron scattering to detect the amount of water in the soil. The hydrogen molecule in water slows down the scattering neutrons and results in thermalized neutrons, which are counted by the instrument and related to the water content. To use this instrument at different soil depths, an access tube is inserted into the soil (Fig. 2). A probe which has a source of neutrons and a counter that measures the thermalized neutrons is inserted through this access tube to a pre-determined depth. For this experiment a WaterMan NWM (Geoquip, South Africa) was used. The cable clamps of this machine were set to take measurements at soil depths of 150, 450, 750, 1 050, 1 350 and 1 650 mm.

Micro-lysimeters

The micro-lysimeters (Fig. 3) used were cylinders with a diameter of 70 mm and a length of 300 mm. Hoffman (1997) and Bennie et al. (1998) have used similar micro-lysimeters to measure evaporation. The soil cores in the lysimeters were prepared by pressing the micro-lysimeters into the soil surface. This was done to produce undisturbed soil columns which were representative of the natural packing. The lysimeters were fitted with stoppers to hold the soil column in place. For easy insertion and removal of the micro-lysimeter in the ground hole, where it is installed, a lining tube was put in place (Sharratt and McWilliams, 2005). This lining tube preserved the hole-integrity by protecting the soil wall from collapsing. The evaporative loss was determined by weighing the lysimeters and computing the difference between consecutive weighing.
This gravimetrically determined change in soil water was converted to depth units by dividing it with the surface area of the micro-lysimeters.

Measurement of soil water evaporation

Evaporation of soil water was measured as the difference between measurement days for the three soil water measuring methods. Evaporation measurement by the mini-lysimeter represented the 300 mm depth. The measurements by DFM probes and the NWM were also considered for the top 300 mm. Unrealistic values (resulting from operator error as measured against the background of ET) values for the day considered) were removed from the dataset and values corresponding to the missing values were not used during comparison of the means; hence the effect was not reflected in the statistical results.

Calibration of the soil water measuring instruments

A laboratory calibration of the DFM probes and NWM was done. For this purpose topsoil was collected from a field on a Kenilworth-Bainsvlei ecotope (Chimungu, 2009). The soil was air-dried and 5 samples were taken for determination of gravimetric soil water content. To prepare soil columns of known water content and bulk density the soil was packed into 210 ℓ plastic drums with a diameter of 540 mm and height of 920 mm. The size of the drums was large enough to contain the sphere of influence created by the probes. The largest radius of the sphere of influence is that of NWM in dry soil, which is about 250 mm (Hillel, 2004). These drums were air-tight containers (Fig. 4) with only 2 small openings on the top through which the soil was packed. An access tube was installed in the middle of the drum (Fig. 4) and the opening properly sealed. To provide the dry-end and wet-end of the soil 2 drums were used. The drum representing the dry-end was filled with a known mass of air-dried soil. Drums were well shaken to imitate the natural packing of the soil. The bulk density was determined by taking the mass of soil filled into the drum and the volume contained by the soil. The drums representing the wet-end were prepared in the same way except that water was added to bring the known soil mass to saturation. After adding water to the soil, it was left for a month to equilibrate. The mass of the water added to saturate the soil column was recorded. The volumetric water content of the soil in the drums was calculated by determining the gravimetric water content from the mass of soil and multiplying by the bulk density divided by the density of water.

Calibration of DFM capacitance probes

For the calibration of the DFM capacitance probes the probes were installed into the drums by pushing them into the soil column and remained in the soil for at least 6 h. Two probes were installed per drum through the openings on top of the drum (Fig. 4). After the probes were inserted, the holes were sealed using plastic tape to prevent water loss through evaporation. The installed probes measured the soil water content of the drums every 30 min. To provide acceptable replication 31 probes were used for calibration. There were 16 measuring sessions on each drum (2 probes being installed at a time). After each session, the drums were well shaken to minimise the disturbance caused by the installation of probes. Due to the limited height of the drums only 3 sensors on each probe recorded the water content. Since the water content of the soil column was homogenous each sensor on a probe was considered individually without any reference to depth. A DFM reading which did not indicate good contact between probe and the soil was discarded and 84 sensor readings from a total of 93 were provided. Split sample procedure of calibration and validation was followed (Chihobvu et al., 2011). The DFM readings were randomly divided into two datasets. Half of the data were used for calibration and the remaining half were used to validate the calibration equation.

Temperature correction for the sensors

It had been observed that readings of capacitance sensors were affected by temperature. Thus correcting for temperature is essential. For this purpose, monitoring of the DFM readings was conducted in a temperature-controlled room. A PVC cylinder filled with de-ionised water provided a homogenous medium into which the DFM probes were immersed. DFM readings were recorded at 5, 15, 25 and 35°C. Each temperature was maintained for a period of 48 h. The slope of the regression between temperature and DFM reading provided the rate of change. The slope was then used to derive a linear correction equation (Eq. (1)) similar to the one reported by Fares et al. (2009). The equation was designed to bring the readings taken at different temperatures to the same temperature of 25°C, a commonly-used standard ambient temperature (Wikipedia contributors, 2012). The equation is given below:

\[
DFM_{\text{corr}} = (25 – DFM_f)^* S + DFM_s
\]

where:

- \( DFM_{\text{corr}} \) = new transformed DFM reading (%)
- \( DFM_f \) = temperature reading from DFM probe (°C)
- \( DFM_s \) = water content reading from DFM probe (%)
- \( S \) = Slope determined from the relationship between temperature and DFM reading

The DFM readings recorded for the soil columns prepared in the drums were temperature corrected using Eq. (1). The slope (S) value found for the DFM probes was 0.04. A linear relationship between DFM readings and soil water content was assumed. The readings of the wet-end and dry-end plotted against their respective soil water contents were used for calibration. A linear regression equation between the DFM readings and known volumetric soil water content was developed.

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Calibration of NWM

The same drums that were used for the calibration of the DFM probes were also used to calibrate the NWM. Measurements were taken twice daily at a depth of 300 and 600 mm. Although the water content was homogenous throughout the drum, since NWM readings are affected by depth, calibration is done separately for the two depths. Four randomly-selected readings were used for calibration, while the remaining four were used to validate the calibration equation.

Statistical analysis

SAS software (SAS Institute Inc., 2006) was used to perform the regression for both DFM readings and NWM readings against the measured volumetric soil water content values. Validation of the calibration equation was done by using the model performance parameters suggested by Willmott (1982). These parameters include the index of agreement (D-index), mean absolute error (MAE), root mean square error (RMSE) with its systematic (RMSE_s) and unsystematic (RMSE_u) components, and the coefficient of determination ($r^2$). Willmott (1982) points out that in a ‘good’ model the RMSE should approach zero; a large RMSE indicates bias. The D-index should approach 1 and the RMSE should be as close as possible to the RMSE, indicating that the deviations of simulated from measured values are random. SAS software was also used for t-test comparison between evaporation values of the micro-lysimeter and DFM as well as the micro-lysimeter and NWM.

RESULTS AND DISCUSSION

Calibration results of DFM probes and NWM

A summary of information about the soil and the statistics for the readings from the capacitance probes and NWM probe are given in Table 1. For a given soil water content, the average reading by the DFM probes and the NWM with the observed variation are shown in Table 1. The coefficient of variation (CV) indicated there was much more variation in the readings taken for the dry soil compared to the wet soil. This trend was observed for both the DFM and NWM probes. The CV observed for the NWM readings of the dry topsoil was exceptionally high. There was also variation between readings taken at the two depths.

The linear regression line indicating the correspondence between DFM reading and volumetric soil water content is presented in Fig. 5. The DFM readings and the volumetric values are expressed as percentages. The regression line and equation are presented on the same figure. The resulting $r^2$ (0.99) indicated that the DFM probe readings strongly correlated with the volumetric soil water content. The linear regression had a prediction interval of 95% which is indicated by the dotted lines on both sides of the regression line. Most of the measured data points, as well as points predicted by the regression line, fall within this interval.

The linear regression between NWM reading and volumetric soil water content is presented in Fig. 6. An $r^2$ of 0.99 indicated there is a good correlation between the NWM readings and volumetric soil water content. Figure 6 also indicates the good performance of the linear regression fit.

Validation of the calibration equation of the DFM probes and NWM

The calibration equation for converting the DFM and NWM readings into volumetric soil water content was validated on separate datasets to those from which they were developed. A summary of the statistical performance of the calibration

<table>
<thead>
<tr>
<th>Soil column</th>
<th>Bulk density (kg·m⁻³)</th>
<th>Clay percentage</th>
<th>Volumetric water content (%)</th>
<th>DFM reading ($n = 84$)</th>
<th>NWM reading ($n = 8$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Air-dried</td>
<td>1 340</td>
<td>7.5</td>
<td>2.2</td>
<td>12.5</td>
<td>10.1</td>
</tr>
<tr>
<td>Saturated</td>
<td>1 340</td>
<td></td>
<td>40.5</td>
<td>67.0</td>
<td>3.4</td>
</tr>
</tbody>
</table>

$CV = \text{coefficient of variation}$
Comparison of DFM probes and NWM for measuring evaporation

The evaporation values computed from observations during rain-free days are presented in Tables 3 to 5. From the 21 measurement days, 5 drying cycles were identified. A drying cycle is a rain-free period of evaporation observation after an initial wetting by rain. The evaporation observation will vary

### Table 2

<table>
<thead>
<tr>
<th>Statistical parameter</th>
<th>Topsoil of the Kenilworth-Bainsvlei ecotope</th>
</tr>
</thead>
<tbody>
<tr>
<td>RMSE (%)</td>
<td>DFM</td>
</tr>
<tr>
<td>RMSE</td>
<td>1.077</td>
</tr>
<tr>
<td>MAE</td>
<td>0.389</td>
</tr>
<tr>
<td>RMSE</td>
<td>1.004</td>
</tr>
<tr>
<td>$r^2$</td>
<td>0.766</td>
</tr>
<tr>
<td>slope ($b$)</td>
<td>0.998</td>
</tr>
<tr>
<td>Y-intercept ($a$)</td>
<td>0.099</td>
</tr>
<tr>
<td>RMSE/RMSE</td>
<td>0.361</td>
</tr>
</tbody>
</table>

* Unrealistic values removed from dataset

### Table 3

<table>
<thead>
<tr>
<th>Evaporation (mm) values measured with the micro-lysimeter technique for bare in-field rainwater harvesting plot of Kenilworth-Bainsvlei ecotope</th>
</tr>
</thead>
<tbody>
<tr>
<td>Drying cycles</td>
</tr>
<tr>
<td>----------------</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>1</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>2</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>3</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>4</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>5</td>
</tr>
<tr>
<td></td>
</tr>
</tbody>
</table>

* Unrealistic values removed from dataset

### Table 4

<table>
<thead>
<tr>
<th>Evaporation (mm) values measured with DFM probes for bare in-field rainwater harvesting plot of Kenilworth-Bainsvlei ecotope</th>
</tr>
</thead>
<tbody>
<tr>
<td>Drying cycles</td>
</tr>
<tr>
<td>----------------</td>
</tr>
<tr>
<td></td>
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<tr>
<td>1</td>
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<tr>
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</tr>
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<td>2</td>
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<tr>
<td>5</td>
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<td></td>
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</tbody>
</table>

* Data removed due to unrealistic response

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depending on the evaporation measurement interval (daily or after 2 days). The drying cycles contained 8 daily evaporation values and 4 three-day cumulative evaporation values.

The paired t-test comparisons done on the evaporation values measured by DFM probes and NWM versus the control (micro-lysimeter) are presented in Table 6. The results showed that the DFM evaporation values were not significantly different at 95% probability. Contrary to the DFM results, all the comparisons between NWM evaporation and control were significant at 95% probability. Contrary to the DFM results, all the comparisons between NWM evaporation and control were significantly different.

The results confirmed that there was good correspondence between the changes in soil water content measured by the micro-lysimeter and the DFM capacitance probes. Since DFM probes are relatively new products in the market, a thorough evaluation of DFM probes with regards to their precision, accuracy and reliability over a wide range of soils is needed. Nevertheless, the use of the NWM to monitor the rather small water content changes in the topsoil is not an unavoidable error for the NWM and best practice is to keep the error as small as possible. This can hamper the NWM from sensing small changes in soil water content brought about by evaporation from the soil surface. Secondly, the NWM reading errors mostly occur in the top horizon of the soil profile where there is a possibility of radiation escaping through the surface and hence not giving a fully representative reading of the water content (McGowan and Williams, 1980). This failure of the NWM to observe daily change in water content, caused

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**TABLE 5**

<table>
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<tr>
<th>Drying cycles</th>
<th>Number of evaporation days</th>
<th>NWM access tubes</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
</tr>
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<tr>
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<td>1</td>
<td>NWM4 - Lys4</td>
<td>0.70</td>
<td>0.23</td>
<td>0.23</td>
<td>0.00</td>
<td>0.93</td>
<td>0.00</td>
<td>0.00</td>
<td>0.23</td>
<td>0.23</td>
<td>0.70</td>
<td>-</td>
<td>0.00</td>
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<tr>
<td></td>
<td>1</td>
<td></td>
<td>0.23</td>
<td>1.40</td>
<td>0.47</td>
<td>-</td>
<td>-</td>
<td>0.47</td>
<td>0.23</td>
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<td>0.23</td>
<td>0.00</td>
<td>0.00</td>
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<td>0.93</td>
<td>0.70</td>
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<td>0.47</td>
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<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>3</td>
<td>1</td>
<td>NWM1 - Lys1</td>
<td>1.16</td>
<td>0.70</td>
<td>0.93</td>
<td>0.70</td>
<td>0.70</td>
<td>0.23</td>
<td>0.70</td>
<td>0.23</td>
<td>0.23</td>
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<td>0.00</td>
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<td>0.23</td>
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<tr>
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<td>NWM6 - Lys6</td>
<td>2.79</td>
<td>0.70</td>
<td>2.56</td>
<td>2.33</td>
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<td>1.16</td>
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<td>0.93</td>
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<td>2.33</td>
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<td>5</td>
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<td>NWM7 - Lys7</td>
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<td>1.13</td>
<td>-</td>
<td>1.32</td>
<td>0.21</td>
<td>NWM3 - Lys3</td>
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<td>9</td>
<td>-7.41</td>
<td>&lt;0.001**</td>
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<td></td>
<td>2.04</td>
<td>10</td>
<td>1.88</td>
<td>0.09</td>
<td>NWM4 - Lys4</td>
<td>-1.02</td>
<td>8</td>
<td>-3.79</td>
<td>0.005**</td>
<td></td>
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<td>-1.05</td>
<td>9</td>
<td>-1.28</td>
<td>0.23</td>
<td>NWM5 - Lys5</td>
<td>-2.38</td>
<td>8</td>
<td>-6.02</td>
<td>&lt;0.001**</td>
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**TABLE 6**

<table>
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<tr>
<th>Difference</th>
<th>Mean</th>
<th>DF</th>
<th>t-value</th>
<th>Pr &gt; [t]</th>
<th>Difference</th>
<th>Mean</th>
<th>DF</th>
<th>t-value</th>
<th>Pr &gt; [t]</th>
</tr>
</thead>
<tbody>
<tr>
<td>DFM1 - Lys1</td>
<td>-0.39</td>
<td>8</td>
<td>-0.44</td>
<td>0.67</td>
<td>NWM1 - Lys1</td>
<td>-1.54</td>
<td>7</td>
<td>-4.37</td>
<td>0.003**</td>
</tr>
<tr>
<td>DFM2 - Lys2</td>
<td>0.82</td>
<td>7</td>
<td>0.57</td>
<td>0.59</td>
<td>NWM2 - Lys2</td>
<td>-2.56</td>
<td>7</td>
<td>-7.40</td>
<td>&lt;0.001**</td>
</tr>
<tr>
<td>DFM3 - Lys3</td>
<td>-1.13</td>
<td>11</td>
<td>-1.32</td>
<td>0.21</td>
<td>NWM3 - Lys3</td>
<td>-1.87</td>
<td>9</td>
<td>-7.41</td>
<td>&lt;0.001**</td>
</tr>
<tr>
<td>DFM4 - Lys4</td>
<td>2.04</td>
<td>10</td>
<td>1.88</td>
<td>0.09</td>
<td>NWM4 - Lys4</td>
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<td>8</td>
<td>-3.79</td>
<td>0.005**</td>
</tr>
<tr>
<td>DFM5 - Lys5</td>
<td>-1.05</td>
<td>9</td>
<td>-1.28</td>
<td>0.23</td>
<td>NWM5 - Lys5</td>
<td>-2.38</td>
<td>8</td>
<td>-6.02</td>
<td>&lt;0.001**</td>
</tr>
<tr>
<td>DFM6 - Lys6</td>
<td>-0.99</td>
<td>11</td>
<td>-1.70</td>
<td>0.12</td>
<td>NWM6 - Lys6</td>
<td>-2.17</td>
<td>9</td>
<td>-6.12</td>
<td>&lt;0.001**</td>
</tr>
<tr>
<td>DFM7 - Lys7</td>
<td>-0.43</td>
<td>11</td>
<td>-0.73</td>
<td>0.48</td>
<td>NWM7 - Lys7</td>
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<td>9</td>
<td>-4.87</td>
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</tr>
<tr>
<td>DFM8 - Lys8</td>
<td>-0.63</td>
<td>11</td>
<td>-1.24</td>
<td>0.24</td>
<td>NWM8 - Lys8</td>
<td>-0.60</td>
<td>9</td>
<td>-2.61</td>
<td>0.028**</td>
</tr>
<tr>
<td>DFM9 - Lys9</td>
<td>-0.90</td>
<td>9</td>
<td>-1.94</td>
<td>0.08</td>
<td>NWM9 - Lys9</td>
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<td>-3.08</td>
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</tr>
<tr>
<td>DFM11 - Lys11</td>
<td>-0.27</td>
<td>10</td>
<td>-0.33</td>
<td>0.75</td>
<td>NWM11 - Lys11</td>
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<td>8</td>
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</tr>
<tr>
<td>DFM12 - Lys12</td>
<td>-0.81</td>
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<td>-1.39</td>
<td>0.19</td>
<td>NWM12 - Lys12</td>
<td>-2.74</td>
<td>8</td>
<td>-4.82</td>
<td>0.001**</td>
</tr>
</tbody>
</table>

* Data removed due to unrealistic response
§ Not measured due to technical problems

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http://dx.doi.org/10.4314/wsa.v39i2.1
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by evaporation from bare soil, is also validated by other data
collected by the authors (not reported here). Daily NWM water
content measurements done on Glen-Bonheim, Glen-Swartland
and Paradyss-Tukulu ecotopes in 2008 showed that no reliable
daily evaporation measurements could be taken using this
method. Hensley et al. (2000) and Botha (2006) have reported
evaporation values from the water content monitoring con-
ducted by NWM. However, in their studies they calculated
evaporation values from the water balance equation, where all
the other variables were known or measured. Their measure-
ments were also done over longer periods and not based on
daily water losses due to evaporation.

Various other studies comparing capacitance probes and
the NWM have been done (Evett et al., 2002; Heng et
al., 2002; Mwale et al., 2005; Mounzer et al., 2008; Vera et
al., 2009). These studies confirmed the observation that the
capacitance probes are sensitive enough to measure small
changes in the soil water content due to water losses through
evaporation from the soil surface. However, the presence of
air-pockets could result in incorrect readings (Mwale et al.,
2005). Thus, to increase the reliability of capacitance probes,
there is a need to ensure proper installation of the probes. On
the other hand, the abovementioned studies confirmed that
the NWM gave consistent readings over a wide range of soil
textures and soil water contents. This was attributed to the
ability of the NWM to measure larger volumes of soil and its
relatively lower sensitivity to soil heterogeneity (Heng et al.,
2002; Vera et al., 2009). These studies have also confirmed the
accuracy of the NWM in measuring the soil water content of
the entire root zone; however, the evaporation-sensitive zone
usually forms only a small fraction of the total root zone.
Hoffman (1997) computed the evaporation-sensitive zone to
generally be the top 300 mm. As discussed above, sensing the
soil water content change in the evaporation-sensitive zone of
the soil profile can be less accurate with NWM. Mounzer et al.
(2008) reported that both capacitance probes and the NWM
performed well in measuring the evapotranspiration from a
cropped field and that the readings obtained with the capaci-
tance probes correlated better with the values predicted by
the FAO methodology.

CONCLUSIONS AND RECOMMENDATIONS

The measurement of soil water evaporation with DFM capaci-
tance probes and NWM was compared. Measurements with
the DFM probes and NWM provided good agreement with the
gravimetrically determined soil water content. Both instru-
ments can be used to monitor the change in soil water content
and the multi-depth measuring capabilities provide a good
tool to measure the soil water content of a soil profile. From
the results it can be concluded that DFM probes provide more
accurate measurements of evaporation from the topsoil than
the NWM. Based on the observations made during the study it
is recommended that proper care be taken when installing the
DFM probes to provide good contact between the probe and the
soil.

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